

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property
Organization
International Bureau



(43) International Publication Date
4 March 2004 (04.03.2004)

PCT

(10) International Publication Number
WO 2004/018627 A3

(51) International Patent Classification⁷: C12N 7/00

Rahway, NJ 07065-0907 (US). MORSY, Manal [US/US];
126 East Lincoln Avenue, Rahway, NJ 07065-0907 (US).

(21) International Application Number:

PCT/US2003/026145

(74) Common Representative: MERCK & CO., INC.; 126
East Lincoln Avenue, Rahway, NJ 07065-0907 (US).

(22) International Filing Date: 21 August 2003 (21.08.2003)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

60/405,182	22 August 2002 (22.08.2002)	US
60/455,234	17 March 2003 (17.03.2003)	US
60/455,312	17 March 2003 (17.03.2003)	US
60/458,825	28 March 2003 (28.03.2003)	US

(81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.

(84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, RO, SE, SI, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

(71) Applicant (*for all designated States except US*): MERCK & CO., INC. [US/US]; 126 East Lincoln Avenue, Rahway, NJ 07065-0907 (US).

(72) Inventors; and

(75) Inventors/Applicants (*for US only*): BETT, Andrew, J. [CA/US]; 126 East Lincoln Avenue, Rahway, NJ 07065-0907 (US). CHASTAIN, Michael [US/US]; 126 East Lincoln Avenue, Rahway, NJ 07065-0907 (US). SANDIG, Volker [DE/US]; 126 East Lincoln Avenue, Rahway, NJ 07065-0907 (US). EMINI, Emilio, A. [US/US]; 126 East Lincoln Avenue, Rahway, NJ 07065-0907 (US). SHIVER, John, W. [US/US]; 126 East Lincoln Avenue, Rahway, NJ 07065-0907 (US). CASIMIRO, Danilo, R. [PH/US]; 126 East Lincoln Avenue, Rahway, NJ 07065-0907 (US). KASLOW, David, C. [US/US]; 126 East Lincoln Avenue,

Published:

— with international search report
— before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments

(88) Date of publication of the international search report:
2 September 2004

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: METHODS FOR PROPAGATING ADENOVIRUS AND VIRUS PRODUCED THEREBY

(57) Abstract: Various methods for propagating and rescuing multiple serotypes of replication-defective adenovirus in a single adenoviral E1-complementing cell line are disclosed. Typically, replication-defective adenovirus vectors propagate only in cell lines which express E1 proteins of the same serotype or subgroup as the vector. The disclosed methods offer the ability to propagate vectors derived from multiple adenoviral serotypes in a single production cell line which expresses E1 proteins from a single serotype. Propagation in this manner is accomplished by providing all or a portion of an E4 region *in cis* within the genome of the replication-defective adenovirus. The added E4 region or portion thereof is cloned from a virus of the same or highly similar serotype as that of the E1 gene product(s) of the complementing cell line. Interaction between the expressed E1 of the cell line and the heterologous E4 of the replication-defective adenoviral vectors enables their propagation and rescue. The invention bypasses a need in the art to customize specific cell lines to the serotype or subgroup of the adenoviral vector being propagated and enables one to easily and rapidly develop alternative adenoviral serotypes as gene delivery vectors for use as vaccines or as a critical component in gene therapy.

WO 2004/018627 A3

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US03/26145

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : C12N 7/00
US CL : 435/235.1

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/5, 235.1; 424/93.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

PubMed

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X — Y	US 5,837,511 A (FALCK-PEDERSEN et al.) 17 November 1998 (17.11.1998), column 7, lines 47-55, 62-67, column 8, lines 6-18, column 9, lines 11-15, 54-57, column 12, lines 35-67, column 16, lines 23-49	1, 2, 4-10, 12, 13, 17-21, 24, 27-37, 43-50, 56-65, 71-78 3, 11, 14-16, 22, 23, 25, 26, 38-42, 51-55, 66-70, 79-83
Y	US 6,200,798 B1 (YEH et al.) 13 March 2001 (13.03.2001), column 5, lines 20-30	3, 14-16, 22, 23, 25, 26, 38-42, 51-55, 66-70, 79-83
Y	US 6,391,612 B1 (BRUDER et al.) 21 May 2002 (21.05.2002), column 5, lines 60-67, column 6, lines 1-10	11

☐ Further documents are listed in the continuation of Box C.☐ See patent family annex.

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Date of the actual completion of the international search

19 March 2004 (19.03.2004)

Date of mailing of the international search report

12 JUL 2004

Name and mailing address of the ISA/US

Mail Stop PCT, Attn: ISA/US
Commissioner for Patents
P.O. Box 1450
Alexandria, Virginia 22313-1450

Facsimile No. (703) 305-3230

Authorized officer

Dorthea J. Lawrence for
Tim Brown

Telephone No. (571) 272-0773



(51) International Patent Classification⁷: C12N
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PCT/US2003/026145
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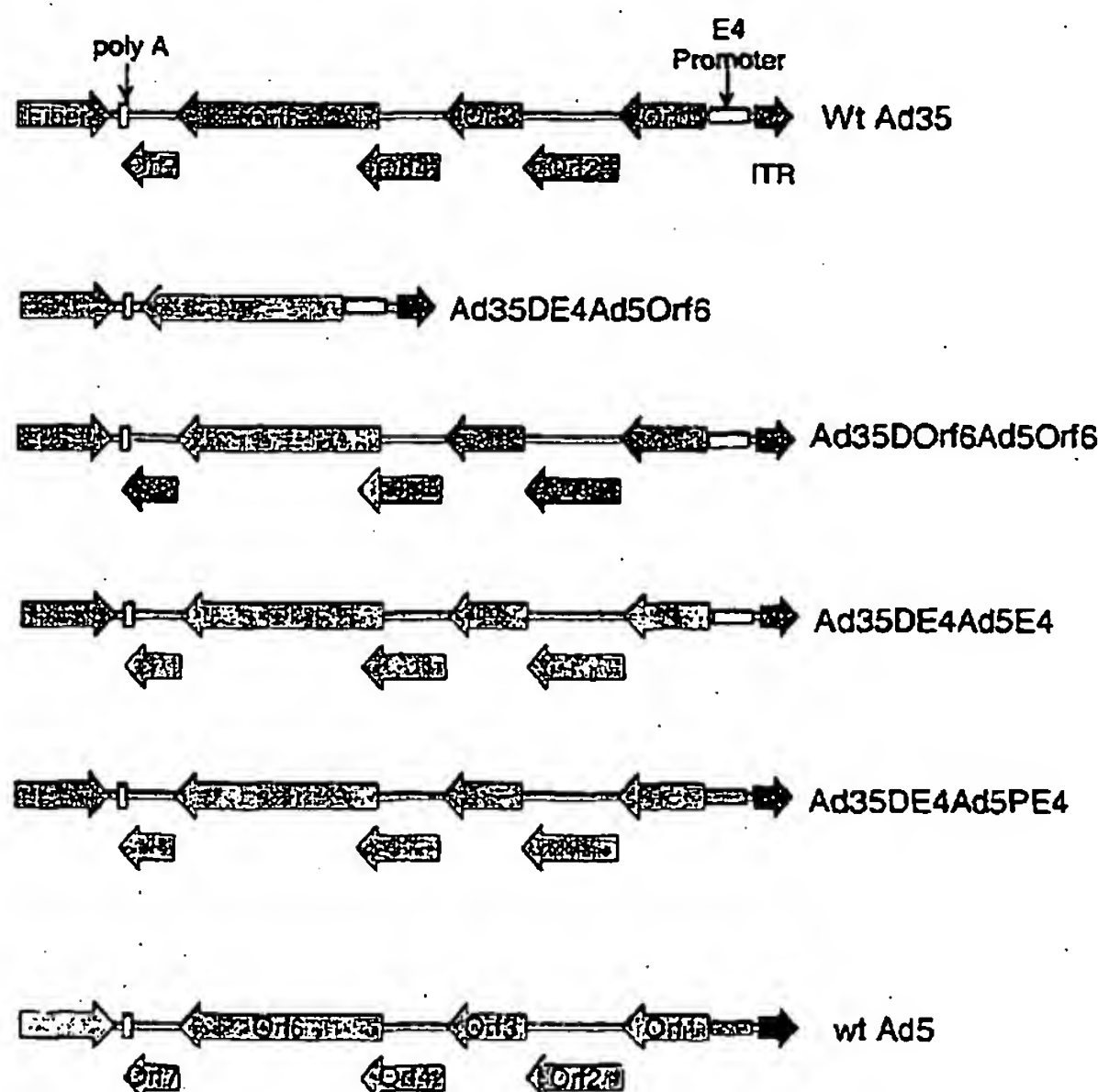
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(57) Abstract: Various methods for propagating and rescuing multiple serotypes of replication-defective adenovirus in a single adenoviral E1-complementing cell line are disclosed. Typically, replication-defective adenovirus vectors propagate only in cell lines which express E1 proteins of the same serotype or subgroup as the vector. The disclosed methods offer the ability to propagate vectors derived from multiple adenoviral serotypes in a single production cell line which expresses E1 proteins from a single serotype. Propagation in this manner is accomplished by providing all or a portion of an E4 region *in cis* within the genome of the replication-defective adenovirus. The added E4 region or portion thereof is cloned from a virus of the same or highly similar serotype as that of the E1 gene product(s) of the complementing cell line. Interaction between the expressed E1 of the cell line and the heterologous E4 of the replication-defective adenoviral vectors enables their propagation and rescue. The invention bypasses a need in the art to customize specific cell lines to the serotype or subgroup of the adenoviral vector being propagated and enables one to easily and rapidly develop alternative adenoviral serotypes as gene delivery vectors for use as vaccines or as a critical component in gene therapy.



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Published:

— without international search report and to be republished upon receipt of that report

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TITLE OF THE INVENTION

METHODS FOR PROPAGATING ADENOVIRUS AND VIRUS PRODUCED THEREBY

CROSS-REFERENCE TO RELATED APPLICATIONS

5 The present application claims the benefit of application serial nos. 60/458,825, filed March 28, 2003; 60/455,312, filed March 17, 2003; 60/455,234, filed March 17, 2003; and 60/405,182, filed August 22, 2002.

FIELD OF THE INVENTION

10 The present invention concerns various methods to propagate and rescue multiple serotypes of replication-defective adenovirus in a single adenoviral E1-complementing cell line. Typically, replication-defective adenovirus vectors propagate only in cell lines which express E1 proteins of the same serotype or subgroup as the vector. The methods disclosed herein offer the ability to propagate vectors derived from multiple serotypes in a single cell line expressing E1
15 proteins from a single serotype. Such propagation of a wide range of vectors in one cell line is accomplished by providing all or a portion of an E4 region *in cis* within the genome of the replication-defective adenovirus. The added E4 region or portion thereof is cloned from a virus of the same or highly similar serotype as that of the E1 gene product(s) of the complementing cell line. Interaction between the E1 gene products of the cell line and the heterologous E4 gene
20 products of the replication-defective adenoviral vector enables the propagation and rescue of the recombinant replication-defective adenovirus vectors. The invention, therefore, bypasses an existing need in the art to customize complementing cell lines to the specific serotype or subgroup of the adenoviral vector being propagated or, alternatively, to have to transfect a cell line with an E4 region and then regulate the expression *in trans* of the E4 region within the E1
25 complementing cell line.

BACKGROUND OF THE INVENTION

Beginning with the first human adenoviruses (Ads) isolated over four decades ago (Rowe *et al.*, *Proc. Soc. Exp. Biol. Med.*, 84:570-579, 1953), over 100 distinct serotypes of
30 adenovirus have been isolated which infect various mammalian species, 51 of which are of human origin (Straus, Adenovirus infections in humans. In *The Adenoviruses*. 451-498, 1984; Hierholzer *et al.*, *J. Infect. Dis.*, 158: 804-813, 1988; Schnurr and Dondero, *Intervirology*, 36: 79-83, 1993; Jong *et al.*, *J Clin Microbiol.*, 37:3940-3945:1999). The human serotypes have been categorised into six subgenera (A-F) based on a number of biological, chemical,
35 immunological and structural criteria; criteria which include hemagglutination properties of rat

and rhesus monkey erythrocytes, DNA homology, restriction enzyme cleavage patterns, percentage of G+C content and oncogenicity (Straus, Adenovirus infections in humans. In *The Adenoviruses*. 451-498, 1984; Horwitz, Adenoviridae and their replication, *In Virology*: 1679-172, 1990).

5 Deletion of an essential E1 region common to the various adenovirus serotypes has enabled the use of adenovirus vectors as gene transfer vectors for vaccine and gene therapy purposes. Resultant replication-defective vectors are propagated in cell lines that provide the deleted E1 gene products *in trans*. Supplementation of the essential E1 gene products *in trans* in this manner works well when the E1 gene products are from the same or a highly similar
10 serotype. As such, E1-deleted group C serotypes (Ad1, Ad2, Ad5 and Ad6) grow well in 293 or PER.C6 cells which contain and express the Ad5 E1 region. In contrast, E1-deleted serotypes other than group C, for example those from subgroups A, B, D, E, and F (e.g., Ad3, Ad4, and Ad7 to Ad51), do not replicate efficiently in 293 or PER.C6 cells. The Ad5 E1 sequences in 293 and PER.C6 cells do not fully complement the replication of these alternative serotypes. This
15 presents a challenge due to the fact that the most characterized and studied complementing cell lines available for growth and propagation of adenovirus are based on E1 sequence from adenovirus serotype 5.

This inability to fully complement the replication of serotypes other than group C adenovirus in Ad5 E1 complementing cell lines has been attributed to the inability of Ad5 (group
20 C) E1b 55K gene product to functionally interact with the E4 gene products of non-group C serotypes. While the interaction is conserved within members of the same subgroup, it is not well conserved between subgroups.

Hence, cell lines expressing both Ad5 E1 and ORF6 were generated and proved useful in complementing alternative adenovirus serotypes; *see, e.g., Abrahamsen et al., 1997 J. Virol.* 8946-8951. Such incorporation of E4 (or ORF6) into Ad 5 complementing cell lines as
25 was done in Abrahamsen *et al., supra*, is known.

U.S. Patent No. 5,849,561 discloses complementation of an E1-deleted non-group C adenovirus vector in an Ad5-E1 complementing cell line which also expresses portions of the Ad5-E4 gene.

30 U.S. Patent No. 6,127,175, issued to Vigne, *et al.*, discloses a stably transfected mammalian cell line which expresses a portion of the E4 region of adenovirus, preferably ORF6 or ORF6/7. Such a cell line is useful for complementation of recombinant Ad genomes deficient in the E4 region.

European Application EP 1 054 064 A1 discloses recombinant, replication
35 deficient adenovirus 35 (Ad35) vectors and cell lines which complement *in trans* the growth of

these vectors. A cell line which expresses Ad5E1A and E2A genes (PER.C6) was shown to complement an Ad35-E1 deleted vector upon co-expression of Ad35-E1B proteins.

U.S. Patent No. 6,270,996, issued to Wilson, *et al.*, discloses E1/E4 deleted adenovirus vectors and E1/E4(ORF6) cell lines which complement *in trans* virus growth without
5 resulting in cell toxicity.

U.S. Patent No. 6,202,060, issued to Mehtali, *et al.*, discloses adenoviral vectors wherein portions of the early genes are under control of an inducible promoter. The '060 patent also discloses complementing cell lines which may be used in tandem with these Ad vectors.

The generation of serotype-specific cell lines providing a complementing
10 serotype-specific E1 gene product(s) *in trans* is known as well.

Although Ad5-based vectors have been used extensively in a number of gene therapy trials, there may be limitations on the use of Ad5 and other group C adenoviral vectors due to preexisting immunity in the general population due to natural infection. Ad5 and other group C members tend to be among the most seroprevalent serotypes. Immunity to existing
15 vectors may develop as a result of exposure to the vector during treatment. These types of preexisting or developed immunity to seroprevalent gene delivery vectors may limit the effectiveness of gene therapy or vaccination efforts. Alternative adenovirus serotypes, thus, constitute very important targets in the pursuit of gene delivery systems capable of evading the host immune response.

20 There remains both a practical and commercial need for an adenovirus-based vaccine and/or gene therapy delivery system which allows for the production of multiple serotype recombinant adenovirus vectors in a single source complementing mammalian cell line. The present invention addresses and overcomes this deficiency in the art by disclosing novel methods for propagating multiple serotype recombinant Ad vectors in a single complementing
25 cell line where the required serotype-specific sequences are provided *in cis*.

SUMMARY OF THE INVENTION

The present invention relates to an enhanced means for propagating replication-defective adenovirus in an E1-complementing cell line(s) where the E1 gene product(s) being
30 expressed is not native to the adenovirus being propagated. The method is based on Applicants' finding that supply, *in cis*, of a nucleic acid sequence encoding all or a portion of a heterologous adenoviral E4 region which is native to a virus of the same or highly similar serotype as the E1 gene product(s) of the complementing cell line enables the growth of adenoviral vectors of varying serotype in any single complementing cell line, despite the fact the cell line is not
35 customized for the particular serotype of vector being propagated. This is of particular

importance given that existing and settled adenoviral E1-complementing cell lines (such as PER.C6™ and 293) are based on one of the most prominent adenovirus serotypes (Ad5) and are not suited for the large-scale propagation and rescue of alternative serotypes.

5 The basic steps involved in the propagation of adenoviral vectors in accordance with the methods of the instant invention are as follows: First, all or a portion of a heterologous adenoviral E4 region comprising nucleic acid sequence encoding at least open reading frame 6 (ORF6) is inserted into a replication-defective adenoviral vector. By "heterologous", Applicants mean that the nucleic acid sequence is not native to the viral vector being propagated, *i.e.*, not normally present within a virus of the same or highly similar serotype. As will be described, the
10 adenoviral E4 region or portion thereof can be either a nucleic acid sequence encoding ORF 6 or any larger portion of the E4 region, and includes nucleic acid comprising the complete E4 region with E4 promoter. The region into which the nucleic acid is incorporated is not limited, *i.e.*, the insertion can be made into the complete E4 region with E4 promoter or into a smaller portion narrowing into the ORF6 region. Alternatively, the heterologous E4 region or portion thereof
15 can be inserted into different areas of the genome such as the E1 or E3 regions. Further, the native E4 region or portion thereof can be deleted and replaced, or left intact. This is not deemed a critical element of the instant invention. What is a critical element is that the heterologous E4 region or portion thereof being inserted is native to a virus of the same or highly similar serotype as the E1 gene product(s) expressed by the complementing cell line.

20 Following the modification of the adenoviral vector of interest, the recombinant adenovirus is then introduced into an adenoviral E1-complementing cell line and allowed to propagate. The adenovirus is subsequently harvested and rescued from the complementing cell line.

The resultant virus can be studied and used in various gene therapy and vaccine
25 efforts. The virus, therefore, forms an important aspect of the instant invention.

BRIEF DESCRIPTION OF THE DRAWINGS

FIGURE 1 illustrates a transcription map for adenovirus serotype 5. The linear genome is divided into 100 map units as well as into r- and l- strands which designate the
30 direction of transcription. Early transcription units are designated with an E and are active prior to viral DNA replication. Late transcription units are designated with and L and are active primarily after DNA replication. Promoters are represented as brackets and polyadenylation sites as arrowheads. The tripartite leader is designated 1, 2, and 3.

FIGURES 2A-1 through 2A-10 illustrate the nucleic acid sequence for the wild-
35 type adenovirus 35 (SEQ ID NO: 1) utilized in the Examples.

FIGURE 3 illustrates the homologous recombination scheme utilized to recover pAd35ΔE1.

FIGURE 4 illustrates the various configurations of the E4 regions (or portions) within the alternative serotype recombinants.

FIGURE 5 illustrates the homologous recombination scheme utilized to recover pAd35ΔE1ΔE4Ad5Orf6.

FIGURE 6 illustrates the nucleic acid sequence encoding the gag expression cassette (SEQ ID NO: 2). The various regions of the figure are as follows: (1) a first underlined segment of nucleic acid sequence encoding the immediate early gene promoter region from human cytomegalovirus; (2) a first segment of lowercase letters which is not underlined, which segment of DNA contains a convenient restriction enzyme site; (3) a region in caps which contains the coding sequence of HIV-1 gag; (4) a second segment of lowercase letters which is not underlined, which segment of DNA contains a convenient restriction enzyme site; and (5) a second underlined segment, this segment containing nucleic acid sequence encoding a bovine growth hormone polyadenylation signal sequence.

FIGURE 7 illustrates the nucleic acid sequence encoding the SEAP expression cassette (SEQ ID NO: 3). The various regions of the figure are as follows: (1) a first underlined segment of nucleic acid sequence encoding the immediate early gene promoter region from human cytomegalovirus; (2) a first segment of lowercase letters which is not underlined, which segment of DNA contains a convenient restriction enzyme site; (3) a region in caps which contains the coding sequence of the human placental SEAP gene; (4) a second segment of lowercase letters which is not underlined, which segment of DNA contains a convenient restriction enzyme site; and (5) a second underlined segment, this segment containing nucleic acid sequence encoding a bovine growth hormone polyadenylation signal sequence.

FIGURE 8 illustrates *in vivo* expression of SEAP in C3H/HeN mice using 10^{10} vp doses of Ad35 vectors. This experiment was designed to address any effects of E3 deletion. The vectors were injected intramuscularly and the levels of SEAP expression were determined from the serum samples. Shown are geometric means for each cohort of 5 mice.

FIGURE 9 illustrates *in vivo* expression of SEAP in C3H/HeN mice using 10^{10} vp doses of Ad35 vectors. This experiment was designed to address any effects of Ad5 sequence insertion into the Ad35 genome. The vectors were injected intramuscularly and the levels of SEAP expression were determined from the serum samples. Two extra cohorts received 10^{10} vp and 10^9 vp of Ad5 vector. Shown are geometric means for each cohort of 5 mice.

FIGURES 10A-B illustrate *in vivo* SEAP expression using MRKAd5-based (A) and Ad35ΔE1ΔE4Ad5Orf6-based (B) vector in rhesus macaques. Shown are the serum antigen

levels for individual monkeys following a single intramuscular (i.m.) injection of 10^{11} vp MRKAd5SEAP (filled circles), 10^9 vp MRKAd5SEAP (open boxes) or 10^{11} vp Ad35 Δ E1SEAP Δ E4Ad5Orf6.

FIGURE 11 illustrates *in vivo* SEAP expression in African green monkeys using Ad5- and Ad35-based vectors. Shown are the antigen levels for each animal in serum samples collected two days after the treatment.

FIGURE 12 illustrates the homologous recombination scheme utilized to recover pAd24 Δ E1.

FIGURE 13 illustrates the homologous recombination scheme utilized to recover pAd24 Δ E1Ad5Orf6.

FIGURE 14 illustrates the configuration of E4 regions in the Ad24 recombinants generated.

FIGURE 15 illustrates the growth kinetics of the Ad24-based vectors in PER.C6 cells.

FIGURES 16A-1 through 16A-10 illustrate the nucleic acid sequence for wild-type adenovirus serotype 24 (SEQ ID NO: 5). The ATCC product number for Ad24 is VR-259.

FIGURE 17 illustrates, in tabular format, gag-specific T cell responses in monkeys immunized with MRKAd5-HIVgag and Ad24 HIV vectors. Shown are the numbers of spot-forming cells per million PBMC following incubation in the absence (mock) or presence of Gag peptide pool. The pool consisted of 20-aa peptide overlapping by 10 aa and encompassing the entire gag sequence.

FIGURE 18 illustrates, in tabular format, the characterization of the gag-specific T cells in monkeys immunized with 10^{11} vp of MRKAd5-HIV1gag and Ad24 Δ E1gag Δ Orf6Ad5Orf6. Shown are the percentages of CD3+ T cells that are either gag-specific CD4+ or gag-specific CD8+ cells. These values were corrected for mock values (<0.03%).

FIGURE 19 illustrates individual anti-p24 titers (in mMU/mL) in macaques immunized with gag-expressing adenovirus vectors.

FIGURE 20 illustrates *in vivo* expression of SEAP in C3H/HeN mice using 10^{10} vp doses of Ad24 vectors. The vectors were injected intramuscularly and the levels of SEAP expression were determined from the serum samples. Two extra cohorts received 10^{10} vp and 10^9 vp of Ad5 vector. Shown are geometric means for each cohort of 5 mice.

FIGURE 21 illustrates *in vivo* SEAP expression using MRKAd5 and Ad24 vectors in rhesus macaques. Shown are the geometric means of the SEAP levels for cohorts of 3 monkeys. In bars are the standard errors of the geometric means.

FIGURE 22 illustrates a homologous recombination scheme to be utilized to recover pAd24ΔE1ΔE4Ad5Orf6.

FIGURE 23 illustrates gag-specific T cell responses in rhesus macaques immunized following a heterologous Ad5/Ad6 prime-Ad24 boost regimen. a: Mock, no peptide: gag, 20-mer peptide pool encompassing entire gag sequence; b: Peak response after 2 or 3 doses of the priming vaccine; c: 3 wks prior to boost; d: 4 wks after boost; e: ND, not determined.

FIGURE 24 illustrates, in tabular format, the percentages of CD3⁺ T lymphocytes that are gag-specific CD8⁺ cells or gag-specific CD4⁺ cells determined after the Ad24 Boost Immunization (wk 60). Numbers reflect the percentages of circulating CD3⁺ lymphocytes that are either gag-specific CD4⁺ or gag-specific CD8⁺ cells. Mock values (equal to or less than 0.01%) have been subtracted.

FIGURE 25 illustrates gag-specific T cell responses in rhesus macaques immunized following a heterologous Ad 24 prime-Ad5 boost regimen. a: Mock, no peptide: gag, 20-mer peptide pool encompassing entire gag sequence; b: Peak response after 2 doses of the priming vaccine; c: Wk 24; d: 4 wks after boost; e: ND, not determined.

FIGURE 26 illustrates the homologous recombination scheme utilized to recover pAd34ΔE1ΔE4Ad5Orf6.

FIGURE 27 illustrates the homologous recombination scheme utilized to recover pMRKAd34ΔE1ΔE4Ad5Orf6.

FIGURES 28A-1 to 28A-9 illustrate a nucleic acid sequence for wild-type adenovirus serotype 34 (SEQ ID NO: 12). The ATCC product number for Ad34 is VR-716.

FIGURE 29 illustrates the time course of SEAP expression using MRKAd5 and Ad34 vectors in rhesus macaques. Data represent cohort geometric means.

FIGURE 30 illustrates, in tabular format, T cell responses induced using MRKAd5 and Ad34 vectors expressing HIV-1 gag. Data are expressed in numbers of spot-forming cells per million PBMC (SFC/10⁶ PBMC). "a" refers to a 20-mer peptide pool with 10-aa overlap and encompassing the entire HIV-1 CAM1 gag.

FIGURE 31 illustrates, in tabular format, the levels of CD4⁺ and CD8⁺ Gag-specific T cells in Ad34-immunized macaques at week 12. "a" refers to a 20-mer peptide pool with 10-aa overlap and encompassing the entire HIV-1 CAM1 gag.

FIGURE 32 illustrates, in tabular format, T cell responses induced using a heterologous Ad34 prime/Ad35 boost regimen in macaques. "a" refers to a 20-mer peptide pool with 10-aa overlap and encompassing the entire HIV-1 CAM1 gag.

FIGURE 33 illustrates, in tabular format, the levels of CD4+ and CD8+ Gag-specific T cells in Ad34 primed/Ad35 boosted macaques at week 28. "a" refers to a 20-mer peptide pool with 10-aa overlap and encompassing the entire HIV-1 CAM1 gag.

5 DETAILED DESCRIPTION OF THE INVENTION

The present invention details an efficient strategy for the propagation and rescue of alternative adenoviral serotypes utilizing available adenovirus production cell lines, nullifying the need to customize available cell lines for a specific serotype of interest. This is enabled by the incorporation of a critical E4 region into the adenovirus to be propagated.

10 The critical E4 region in the instant invention comprises, in the minimum, nucleic acid sequence encoding E4 ORF6 and can comprise the entire region of E4, inclusive of the promoter region. An important characteristic of the imported E4 region is that it is native to a virus of the same or highly similar serotype as the E1 gene product(s) (particularly E1B 55K) of the E1-complementing cell line, but heterologous to (*i.e.*, non-native to a virus of the same
15 serotype as) the adenoviral vector being propagated. As will be detailed below, the heterologous E4 region or portion thereof can be varied and can be inserted into the vector backbone at numerous locations.

The heterologous E4 region or portion thereof can, for instance, be a nucleic acid sequence encoding the entire open reading frame of the non-native E4. This segment of nucleic
20 acid sequence can, in turn, be incorporated into the "native" entire E4 open reading frame of the recipient virus. In such an embodiment, the promoter native to the adenoviral vector would drive the expression of the non-native E4 region within the recombinant replication-defective adenoviral vector. Alternatively, the nucleic acid sequence encoding the entire open reading frame can be inserted into a different region of the adenoviral vector genome, such as for
25 example the E1 or E3 regions. In this latter embodiment, the native E4 region or portion thereof can be deleted or left intact.

In another embodiment, the heterologous E4 region comprises a nucleic acid sequence encoding the entire open reading frame of E4 and includes a non-native E4 promoter. In this type of embodiment, the E4 region can be inserted into the location of the combined
30 native E4 and E4 promoter region. The non-native E4 region in this embodiment would be driven by expression of the non-native E4 promoter. Alternatively, the nucleic acid sequence encoding the entire open reading frame and the non-native E4 promoter can be inserted into a different region of the adenoviral vector genome, such as for example the E1 or E3 regions. In this latter embodiment, the native E4 region or portion thereof can be deleted or left intact.

An alternative and further embodiment exists wherein the heterologous E4 region or portion thereof comprises nucleic acid sequence encoding a partial E4 region comprising ORF6 (one aspect of which is a region solely encoding ORF6). In this particular aspect of the invention, the heterologous non-native E4 protein can, in certain embodiments, replace the non-native ORF6 region or the entire E4-encoding region of the native virus. In the latter situation, the promoter driving expression of the non-native ORF6 can either be the native E4 promoter or a heterologous, non-native promoter operatively linked to the non-native ORF6, while in the latter, the expression of the non-native ORF6 would generally be driven by the native E4 promoter. Alternatively, the nucleic acid sequence encoding a partial E4 region comprising ORF6 can be inserted into a different region of the adenoviral vector genome, such as for example the E1 or E3 regions. In this latter embodiment, the native E4 region or portion thereof can be deleted or left intact.

As one of skill in the art can appreciate, there are various ways in which one can envision the supply of a heterologous E4 nucleic acid sequence *in cis* to an adenoviral vector and thereby enable its growth based on Applicants' novel findings herein. Moreover, as one of skill in the art can appreciate, either native or non-native promoters can be utilized to drive expression of the heterologous E4 region or portion thereof.

Adenovirus pre-plasmids (plasmids comprising the genome of the replication-defective adenovirus with desired deletions and insertions) can be generated by homologous recombination using adenovirus backbones and an appropriate shuttle vector (designed to target in specific deletions and incorporate desired restriction sites into the resultant plasmid). Shuttle vectors of use in this process can be generated using general methods widely understood and appreciated in the art, *e.g.*, PCR of the adenoviral terminal ends taking into account the desired deletions, and the sequential cloning of the respective segments into an appropriate cloning plasmid. The adenoviral pre-plasmid can then be digested and transfected into the complementing cell line via calcium phosphate co-precipitation or other suitable means. Virus replication and amplification then occurs, a phenomenon made evident by notable cytopathic effect. Infected cells and media are then harvested after viral replication is complete (generally, 7-10 days post-transfection).

It is to be noted that various alternative adenoviral serotypes can be developed in accordance with the disclosed methods and, particularly, alternative adenoviral serotype vectors that were previously unable to be propagated or very inefficiently propagated utilizing existing adenoviral production cell lines based on subgroup C complementing E1 sequence. The various adenoviral vectors that can be developed in accordance with the instant methods include adenoviral vectors of subgroups A-F (for instance, serotypes of subgroups A, B (*e.g.*, serotypes

11, 14, 16, 21, 34 and 35), C (e.g., serotypes 2 and 5), D (e.g., serotypes 24, 26 and 36), E (e.g., serotype 4) and F.

In preferred embodiments, the various non-group C family members can be developed with heterologous E4 supplied from a subgroup C member such as adenovirus serotype 5. Particular embodiments of the instant invention utilize a development scheme wherein the heterologous E4 protein is derived from a wildtype adenovirus serotype 5 sequence; see, e.g., a viral sequence which has been deposited with the American Type Culture Collection ("ATCC") under ATCC Deposit No. VR-5 (for which a transcription map can be found in Figure 1). A particular example of this type of embodiment is wherein an adenovirus of subgroup B (or any non-C subgroup) comprising heterologous E4 proteins *in cis* from Ad5 is propagated in Ad5 E1-complementing cell lines, for instance, PER.C6™ or 293. Applicants have, in fact, successfully propagated E1- serotypes 10, 24, 34, and 35 via use of this particular embodiment.

One of skill in the art can readily identify alternative adenovirus serotypes (e.g., alternative serotypes of subgroups A, B (e.g., serotypes 11, 14, 16, 21, 34 and 35), C, (e.g., serotypes 2 and 5), D (e.g., serotypes 24, 26 and 36), E (e.g., serotype 4) and F) for the supply of the heterologous E4 protein. As long as the heterologous E4 region (or portion thereof comprising ORF6) of the vector is native to a virus of the same or highly similar serotype as the E1 region of the complementing cell line, the methods of the instant invention are widely applicable to the propagation and rescue of adenovirus of all serotypes. In light of the present disclosure, one can readily envision, for instance, how a complementing cell line based on a non-subgroup C adenovirus (e.g., the Ad35 cell line of EP 1 054 064 A1) can be utilized to propagate a virus of an adenoviral vector of subgroup C (e.g., adenovirus serotype 5) provided that the appropriate nucleic acid sequence encoding an E4 protein provided *in cis* is native to a virus of the same or highly similar serotype as that of the E1 expressed by the complementing cell line (i.e., an Ad35 E4 protein).

Complementing cell lines of use in the instant invention are available in the art and are not limited to any specific type. The critical feature, again, is that the heterologous segment of E4-encoding nucleic acid sequence provided *in cis* to the replication-defective vector being propagated be native to a virus of the same or highly similar serotype as the E1 expressed by the complementing cell line. One aspect of the instant invention employs E1-complementing cell lines wherein the expressed E1 is of serotype 5; e.g., PER.C6™ and 293 cell lines. Both these cell lines express the adenoviral E1 gene product. PER.C6™ is described in Fallaux *et al.*, 1998 *Human Gene Therapy* 9:1909-1917, hereby incorporated by reference. 293 cell lines are described in Graham *et al.*, 1977 *J. Gen. Virol.* 36:59-72, hereby incorporated by reference.

Another aspect of the instant invention are the adenoviral vectors of any serotype falling with adenoviral subgroups A, B, C, D, E and F (for instance, alternative serotypes of subgroups A, B (*e.g.*, serotypes 11, 14, 16, 21, 34 and 35), C (*e.g.*, serotype 2), D (*e.g.*, serotypes 24, 26 and 36), E (*e.g.*, serotype 4) and F) which are modified to contain a non-native E4-
5 encoding nucleic acid sequence *in cis* which comprises, in whole or in part, nucleic acid sequence encoding open reading frame 6 (ORF6). Virus in accordance with this description can be propagated in accordance with the above-described methods and rescued using any suitable means known in the art.

Another aspect of the instant invention is a vector in accordance with the instant
10 invention which comprises a heterologous passenger gene in addition to that of the heterologous E4 nucleic acid sequence. In specific embodiments, the passenger gene encodes an antigen.

As one of ordinary skill in the art will appreciate, the instant methods are not limited by the heterologous gene that can be incorporated. The instant invention relates generally to a means by which to propagate multiple serotypes of adenovirus in a single
15 complementing cell line and the recombinant virus that make the process possible. In preferred embodiments, the passenger gene is incorporated into the E1 deletion. In alternatively preferred embodiments, the passenger gene is inserted in an E3-deleted region. The position of the passenger gene, as one of ordinary skill in the art will appreciate, can be varied according to the specific complementing cell utilized and the specific deletions present within the replication-
20 defective adenovirus genome.

In specific embodiments the passenger gene can encode an HIV-1 antigen, and in more preferred embodiments selected from the group consisting of genes encoding HIV-1 gag, pol, nef and env. In alternative embodiments, the passenger gene can be a reporter gene, such as secreted alkaline phosphatase (SEAP).

The passenger gene preferably exists in the form of an expression cassette. A
25 gene expression cassette preferably comprises (a) a nucleic acid sequence encoding a protein of interest; (b) a promoter operatively linked to the nucleic acid sequence encoding the protein; and (c) a transcription termination sequence. The transcriptional promoter of the adenoviral vector is preferably recognized by an eukaryotic RNA polymerase. In a preferred embodiment, the
30 promoter is a "strong" or "efficient" promoter. An example of a strong promoter is the immediate early human cytomegalovirus promoter (Chapman *et al.*, 1991 *Nucl. Acids Res.* 19:3979-3986), which is hereby incorporated by reference), in certain embodiments without intronic sequences. Those skilled in the art, however, will appreciate that any of a number of other known promoters, such as the strong immunoglobulin, or other eukaryotic gene promoters

may also be used, including the EF1 alpha promoter, the murine CMV promoter, Rous sarcoma virus (RSV) promoter, SV40 early/late promoters and the beta-actin promoter.

The promoter may comprise a regulatable sequence such as the Tet operator sequence. This is extremely useful, for example, in cases where the gene products are affecting a result other than that desired and repression is sought.

Transcription termination sequences can also be utilized within the gene expression cassettes. Preferred termination sequences are, for instance, the bovine growth hormone terminator/polyadenylation signal (bGHpA) and the short synthetic polyA signal (SPA) of 50 nucleotides in length, defined as follows:

AATAAAAGATCTTTATTTTCATTAGATCTGTGTGTTGGT-TTTTGTGTG (SEQ ID NO:4).

Further embodiments incorporate a leader or signal peptide into the transgene. A preferred leader is that from the tissue-specific plasminogen activator protein, tPA.

The following non-limiting Examples are presented to better illustrate the invention.

EXAMPLE 1

Construction and Rescue

An E1- Ad35-based pre-adenovirus plasmid was constructed in order to determine whether an E1- Ad35 vector (a representative group B serotype) could be propagated in a group C E1-complementing cell line. The general strategy used to recover Ad35 as a bacterial plasmid is illustrated in Figure 3. Cotransformation of BJ5183 bacteria with purified wild-type Ad35 viral DNA and a second DNA fragment termed the Ad35 ITR cassette resulted in the circularization of the viral genome by homologous recombination. The ITR cassette contains sequences from the right (bp 34419 to 34793) and left (bp 4 to 456 and bp 3403 to 3886) end of the Ad35 genome (see Figures 2A-1 to 2A-10) separated by plasmid sequences containing a bacterial origin of replication and an Ampicillin resistance gene. The ITR cassette contains a deletion of E1 sequences from Ad5 457 to 3402 with a unique *Swa* I site located in the deletion. The Ad35 sequences in the ITR cassette provide regions of homology with the purified Ad35 viral DNA in which recombination can occur. The ITR cassette was also designed to contain unique restriction enzyme sites (*Pme* I) located at the end of the viral ITR's so that digestion will release the Ad35 genome from plasmid sequences. Potential clones were screened by restriction analysis and one clone was selected as pAd35ΔE1. Pre-Adenovirus plasmid pAd35ΔE1 contains Ad35 sequences from 4 to 456 and bp 3403 to 34793.

To determine if pre-adenovirus plasmid pAd35ΔE1 could be rescued into virus and propagated in a group C E1 complementing cell line, the plasmid was digested with *Pme* I and transfected into a T-25 flask of PER.C6 cells using the calcium phosphate co-precipitation technique. *Pme* I digestion releases the viral genome from the plasmid sequences allowing viral replication to occur after entry into 293 cells. Viral cytopathic effect (CPE), indicating that virus replication and amplification is occurring, was never observed. Cells and media from the transfection were harvested at 14 days post transfection, freeze-thawed three times, clarified by centrifugation and used to infect new PER.C6 cells but no virus was ever amplified. Following multiple attempts, we have been unable to rescue and amplify pAd35ΔE1 in PER.C6 cells.

EXAMPLE 2

Insertion of Ad5 Orf 6 and Ad5 E4 into the Ad5 Genome

To refine the strategy of including Ad5 Orf6 in the genome of an alternative serotype so that propagation could take place in a Ad5/group C complementing cell line four additional strategies were developed. In the first strategy, the entire alternative serotype E4 region (not including the E4 promoter) was deleted and replaced with Ad5 Orf6. In the second strategy, just the alternative serotype Orf6 gene was deleted and replaced with Ad5 Orf6. In the third strategy, the entire alternative serotype E4 coding region (not including the E4 promoter) was deleted and replaced with the Ad5 E4 coding region (not including the Ad5 E4 promoter) and, in the final strategy, the entire alternative serotype E4 coding and promoter region was deleted and replaced with the Ad5 E4 promoter and coding region. The configuration of the E4 regions generated by the four strategies is diagramed in Figure 4. For each of these strategies the desired pre-Adenovirus plasmid was generated by bacterial recombination. Cotransformation of BJ 5183 bacteria with purified wild-type viral DNA and the appropriately constructed ITR cassette resulted in the circularization of the viral genome by homologous recombination. The construction of each pre-Ad plasmid, based on Ad35, is outlined below:

To construct pAd35ΔE1ΔE4Ad5Orf6 (An Ad35 pre-Ad plasmid containing an E1 deletion and an E4 deletion substituted with Ad5 Orf6), an Ad35 ITR cassette was constructed containing sequences from the right (bp 31599 to 31913 and bp 34419 to 34793) and left (bp 4 to 456 and bp 3403 to 3886) end of the Ad35 genome separated by plasmid sequences containing a bacterial origin of replication and an ampicillin resistance gene. These four segments were generated by PCR and cloned sequentially into pNEB193, generating pNEBAd35-4. Next the Ad5 Orf6 open reading frame was generated by PCR and cloned between Ad35 bp 31913 and 34419 generating pNEBAd35-4Ad5Orf6 (the ITR cassette). PNEB193 is a commonly used commercially available cloning plasmid (New England Biolabs cat# N3051S) containing a

bacterial origin of replication, ampicillin resistance gene and a multiple cloning site into which the PCR products were introduced. The ITR cassette contains a deletion of E1 sequences from Ad35 bp 457 to 3402 with a unique *Swa* I restriction site located in the deletion and an E4 deletion from Ad35 bp 31912 to 34418 into which Ad5 Orf6 was introduced in an E4 parallel orientation. In this construct, Ad5Orf6 expression is driven by the Ad35 E4 promoter. The Ad35 sequences (bp 31599 to 31913 and bp 3403 to 3886) in the ITR cassette provide regions of homology with the purified Ad35 viral DNA in which bacterial recombination can occur following cotransformation into BJ 5183 bacteria (Figure 5). The ITR cassette was also designed to contain unique restriction enzyme sites (*Pme*I) located at the end of the viral ITR's so that digestion will release the recombinant Ad35 genome from plasmid sequences. Potential clones were screened by restriction analysis and one clone was selected as pAd35 Δ E1 Δ E4Ad5Orf6. Pre-Adenovirus plasmid pAd35 Δ E1 Δ E4Ad5Orf6 contains Ad35 sequences from bp 4 to 456; bp 3403 to bp 31913 and bp 34419 to bp 34793 with Ad5Orf6 cloned between bp 31913 and bp 34419.

To construct pAd35 Δ E1 Δ Orf6Ad5Orf6 (An Ad35 pre-Ad plasmid containing an E1 deletion and a deletion of E4 Orf6 substituted with Ad5 Orf6), an Ad35 ITR cassette was constructed containing sequences from the right (bp 31599 to 32081 and bp 32990 to 34793) and left (bp 4 to 456 and bp 3403 to 3886) end of the Ad35 genome separated by plasmid sequences containing a bacterial origin of replication and an ampicillin resistance gene. These four segments were generated by PCR and cloned sequentially into pNEB193, generating pNEBAd35-10. Next the Ad5 Orf6 open reading frame was generated by PCR and cloned between Ad35 bp 32081 and 32990 generating pNEBAd35-10Ad5Orf6 (the ITR cassette). PNEB193 is a commonly used commercially available cloning plasmid (New England Biolabs cat# N3051S) containing a bacterial origin of replication, ampicillin resistance gene and a multiple cloning site into which the PCR products were introduced. The ITR cassette contains a deletion of E1 sequences from Ad35 bp 457 to 3402 with a unique *Swa* I restriction site located in the deletion and a deletion of E4 Orf6 from Ad35 bp 32082 to 32989 into which Ad5 Orf6 was introduced in an E4 parallel orientation. In this construct, Ad5Orf6 expression is driven by the Ad35 E4 promoter. The Ad35 sequences (bp 31599 to 32081 and bp 3403 to 3886) in the ITR cassette provide regions of homology with the purified Ad35 viral DNA in which bacterial recombination can occur following cotransformation into BJ 5183 bacteria. The ITR cassette was also designed to contain unique restriction enzyme sites (*Pme* I) located at the end of the viral ITR's so that digestion will release the recombinant Ad35 genome from plasmid sequences. Potential clones were screened by restriction analysis and one clone was selected as pAd35 Δ E1 Δ Orf6Ad5Orf6. Pre-Adenovirus plasmid pAd35 Δ E1 Δ Orf6Ad5Orf6 contains Ad35

sequences from bp 4 to 456; bp 3403 to bp 32081 and bp 32990 to bp 34793 with Ad5Orf6 cloned between bp 32081 and bp 32990.

To construct pAd35 Δ E1 Δ E4Ad5E4 (An Ad35 pre-Ad plasmid containing an E1 deletion and a deletion of E4 substituted with Ad5 E4), an Ad35 ITR cassette was constructed containing sequences from the right (bp 31599 to 31838 and bp 34419 to 34793) and left (bp 4 to 456 and bp 3403 to 3886) end of the Ad35 genome separated by plasmid sequences containing a bacterial origin of replication and an ampicillin resistance gene. These four segments were generated by PCR and cloned sequentially into pNEB193, generating pNEBAd35-7. Next the Ad5 E4 coding region was generated by PCR and cloned between Ad35 bp 31838 and 34419 generating pNEBAd35-7Ad5E4-2 (the ITR cassette). PNEB193 is a commonly used commercially available cloning plasmid (New England Biolabs cat# N3051S) containing a bacterial origin of replication, ampicillin resistance gene and a multiple cloning site into which the PCR products were introduced. The ITR cassette contains a deletion of E1 sequences from Ad35 bp 457 to 3402 with a unique *Swa* I restriction site located in the deletion and an E4 deletion from Ad35 bp 31839 to 34418 into which the Ad5 E4 coding region was introduced in an E4 parallel orientation. In this construct, the Ad5 E4 region is expressed using the Ad35 E4 promoter. The Ad35 sequences (bp 31599 to 31838 and bp 3403 to 3886) in the ITR cassette provide regions of homology with the purified Ad35 viral DNA in which bacterial recombination can occur following cotransformation into BJ 5183 bacteria. The ITR cassette was also designed to contain unique restriction enzyme sites (*Pme* I) located at the end of the viral ITR's so that digestion will release the recombinant Ad35 genome from plasmid sequences. Potential clones were screened by restriction analysis and one clone was selected as pAd35 Δ E1 Δ E4Ad5E4. Pre-Adenovirus plasmid pAd35 Δ E1 Δ E4Ad5E4 contains Ad35 sequences from bp 4 to 456; bp 3403 to bp 31838 and bp 34419 to bp 34793 with the Ad5 E4 coding region (Ad 5 bp 32914 to bp 35523) cloned between bp 31838 and bp 34419.

To construct pAd35 Δ E1 Δ E4Ad5PE4 (An Ad35 pre-Ad plasmid containing an E1 deletion and a deletion of E4 coding region and promoter substituted with Ad5 E4 coding region and promoter), an Ad35 ITR cassette was constructed containing sequences from the right (bp 31599 to 31838 and bp 34660 to 34793) and left (bp 4 to 456 and bp 3403 to 3886) end of the Ad35 genome separated by plasmid sequences containing a bacterial origin of replication and an ampicillin resistance gene. These four segments were generated by PCR and cloned sequentially into pNEB193, generating pNEBAd35-8. Next the Ad5 E4 promoter and coding region was generated by PCR and cloned between Ad35 bp 31838 and 34660 generating pNEBAd35-8Ad5E4PC (the ITR cassette). PNEB193 is a commonly used commercially available cloning plasmid (New England Biolabs cat# N3051S) containing a bacterial origin of replication,

ampicillin resistance gene, and a multiple cloning site into which the PCR products were introduced. The ITR cassette contains a deletion of E1 sequences from Ad35 bp 457 to 3402 with a unique *Swa* I restriction site located in the deletion and an E4 deletion from Ad35 bp 31839 to 34659 into which the Ad5 E4 promoter and coding region was introduced in an E4 parallel orientation. In this construct, the Ad5 E4 region is expressed using the Ad5 E4 promoter. The Ad35 sequences (bp 31599 to 31838 and bp 3403 to 3886) in the ITR cassette provide regions of homology with the purified Ad35 viral DNA in which bacterial recombination can occur following cotransformation into BJ 5183 bacteria. The ITR cassette was also designed to contain unique restriction enzyme sites (*Pme* I) located at the end of the viral ITR's so that digestion will release the recombinant Ad35 genome from plasmid sequences. Potential clones were screened by restriction analysis and one clone was selected as pAd35ΔE1ΔE4Ad5PE4. Pre-Adenovirus plasmid pAd35ΔE1ΔE4Ad5PE4 contains Ad35 sequences from bp 4 to 456; bp 3403 to bp 31838 and bp 34660 to bp 34793 with the Ad5 E4 promoter and coding region (Ad 5 bp 32914 to bp 35826) cloned between bp 31838 and bp 34660.

EXAMPLE 3

Rescue of pAd35ΔE1ΔE4Ad5Orf6, pAd35ΔE1ΔOrf6Ad5Orf6, pAd35ΔE1ΔE4Ad5E4 and pAd35ΔE1ΔE4Ad5PE4 into Virus

In order to determine if pre-adenovirus plasmids pAd35ΔE1ΔE4Ad5Orf6, pAd35ΔE1ΔOrf6Ad5Orf6, pAd35ΔE1ΔE4Ad5E4 and pAd35ΔE1ΔE4Ad5PE4 could be rescued into virus and propagated in a group C E1 complementing cell line, the plasmids were each digested with *Pme* I and transfected into T-25 flasks of PER.C6 cells using the calcium phosphate co-precipitation technique; Cell Pect Transfection Kit, Amersham Pharmacia Biotech Inc. *Pme*I digestion releases the viral genome from plasmid sequences allowing viral replication to occur after cell entry. Viral cytopathic effect (CPE), indicating that virus replication and amplification was occurring, was observed for all construct. When CPE was complete, approximately 7-10 days post transfection, the infected cells and media were harvested, freeze/thawed three times and the cell debris pelleted by centrifugation. Approximately 1 ml of the cell lysate was used to infect a T-225 flasks of PER.C6 cells at 80-90% confluence. Once CPE was reached, infected cells and media were harvested, freeze/thawed three times and the cell debris pelleted by centrifugation. Clarified cell lysates were then used to infect 2-layer NUNC cell factories of PER.C6 cells. Following complete CPE the virus was purified by ultracentrifugation on CsCl density gradients. In order to verify the genetic structure of the rescued viruses, viral DNA was extracted using pronase treatment followed by phenol chloroform extraction and ethanol precipitation. Viral DNA was then

digested with *Hind*III and treated with Klenow fragment to end-label the restriction fragments with P33-dATP. The end-labeled restriction fragments were then size-fractionated by gel electrophoresis and visualized by autoradiography. The digestion products were compared with the digestion products of the corresponding pre-Adenovirus plasmid (that had been digested with *Pme*I/*Hind*III prior to labeling) from which they were derived. The expected sizes were observed, indicating that the viruses had been successfully rescued.

EXAMPLE 4

Insertion of an Expression Cassette into pAd35ΔE1ΔE4Ad5Orf6, pAd35ΔE1ΔOrf6Ad5Orf6, pAd35ΔE1ΔE4Ad5E4 and pAd35ΔE1ΔE4Ad5PE4

In order to introduce a gag or SEAP expression cassette into the E1 region of the various Ad35 pre-Adenovirus plasmids described above (pAd35ΔE1ΔE4Ad5Orf6, pAd35ΔE1ΔOrf6Ad5Orf6, pAd35ΔE1ΔE4Ad5E4 and pAd35ΔE1ΔE4Ad5PE4) bacterial recombination was again used. A gag expression cassette consisting of the following: 1) the immediate early gene promoter from the human cytomegalovirus, 2) the coding sequence of the human immunodeficiency virus type 1 (HIV-1) gag (strain CAM-1; 1526 bp) gene, and 3) the bovine growth hormone polyadenylation signal sequence (Figure 6), was cloned into the E1 deletion in Ad35 shuttle plasmid, pNEBAd35-2 (a precursor to the Ad35 ITR cassettes described above), generating pNEBAd35CMVgagBGHPA. pNEBAd35-2 contains Ad35 sequences from the left end of the genome (bp 4 to 456 and bp 3403 to 3886) with a unique *Swa*I site between bp 456 and 3403 at the position of the deletion. The gag expression cassette was obtained from a previously constructed shuttle plasmid by *Eco*RI digestion. Following the digestion the desired fragment was gel purified, treated with Klenow to obtain blunt ends and cloned into the *Swa*I site in pNEBAd35-2. This cloning step resulted in the gag expression cassette being cloned into the E1 deletion between bp 456 and 3403 in the E1 parallel orientation. The shuttle vector containing the gag transgene was digested to generate a DNA fragment consisting of the gag expression cassette flanked by Ad35 bp 4 to 456 and bp 3403 to 3886 and the fragment was purified after electrophoresis on an agarose gel. Cotransformation of BJ 5183 bacteria with the shuttle vector fragment and one of the Ad35 pre-Ad plasmids (pAd35ΔE1ΔE4Ad5Orf6, pAd35ΔE1ΔOrf6Ad5Orf6, pAd35ΔE1ΔE4Ad5E4, pAd35ΔE1ΔE4Ad5PE4), linearized in the E1 region by digestion with *Swa* I, resulted in the generation of corresponding Ad35 gag-containing pre-Adenovirus plasmids (pAd35ΔE1gagΔE4Ad5Orf6, pAd35ΔE1gagΔOrf6Ad5Orf6, pAd35ΔE1gagΔE4Ad5E4, and pAd35ΔE1gagΔE4Ad5PE4) by homologous recombination. Potential clones were screened by restriction analysis.

A similar strategy was used to generate Ad35 pre-Ad plasmids containing a SEAP expression cassette. In this case a SEAP expression cassette consisting of: 1) the immediate early gene promoter from the human cytomegalovirus, 2) the coding sequence of the human placental SEAP gene, and 3) the bovine growth hormone polyadenylation signal sequence (Figure 7) was cloned into the E1 deletion in Ad35 shuttle plasmid, pNEBAd35-2, generating pNEBAd35CMVSEAPBGHPA. The SEAP expression cassette was obtained from a previously constructed shuttle plasmid by EcoRI digestion. Following the digestion the desired fragment was gel purified, treated with Klenow to obtain blunt ends and cloned into the SmaI site in pNEBAd35-2. The transgene was then recombined into the various Ad35 backbones generating pAd35ΔE1SEAPΔE4Ad5Orf6, pAd35ΔE1SEAPΔOrf6Ad5Orf6, pAd35ΔE1SEAPΔE4Ad5E4, and pAd35ΔE1SEAPΔE4Ad5PE4 as described above for the gag transgene. All pre-Ad plasmids were rescued into virus and expanded to prepare CsCl purified stocks as described above.

15 EXAMPLE 5

In vivo Transgene Expression

A. Immunization

Female mice were between 4-10 weeks old. The total dose of each vaccine was suspended in 0.1 mL of buffer. The vectors were given to both quadriceps of each animal with a volume of 50 μL per quad and using 0.3-mL 28G1/2 insulin syringes (Becton-Dickinson, Franklin Lakes, NJ). The rhesus macaques and African green monkeys were between 2-5 kg in weight. For the primates, the total dose of each vaccine was suspended in 1 mL of buffer. The monkeys were anesthetized (ketamine/xylazine mixture) and the vaccines were delivered i.m. in 0.5-mL aliquots into two muscle sites using tuberculin syringes (Becton-Dickinson, Franklin Lakes, NJ). Serum samples were collected at defined intervals and stored frozen until the assay date. All animal care and treatment were in accordance with standards approved by the Institutional Animal Care and Use Committee according to the principles set forth in the *Guide for Care and Use of Laboratory Animals*, Institute of Laboratory Animal Resources, National Research Council.

B. SEAP Assay

Serum samples were analyzed for circulating SEAP levels using TROPIX phospho-light chemiluminescent kit (Applied Biosystems Inc). Duplicate 5 μL aliquots of each serum were mixed with 45 μL of kit-supplied dilution buffer in a 96-well white DYNEX plate.

Serially diluted solutions of a human placental alkaline phosphatase (Catalog no. M5905, Sigma, St. Louis, MO) in 10% naïve monkey or mouse serum served to provide the standard curve. Endogenous SEAP activity in the samples was inactivated by heating the well for 30 minutes at 65 °C. Enzymatic SEAP activities in the samples were determined following the procedures described in the kit. Chemiluminescence readings (in relative light units) were recorded using DYNEX luminometer. RLU readings are converted to ng/mL SEAP using a log-log regression analyses.

C. Rodent Results

In the first mouse experiment, cohorts of 5 C3H/HeN mice were given single intramuscular injections of one of the following vectors: (1) 10^{10} vp Ad35 Δ E1SEAP Δ E4Ad5Orf6; (2) 10^{10} vp Ad35 Δ E1SEAP Δ E3 Δ E4Ad5Orf6; or (3) 10^{10} vp Ad35 Δ E1SEAP. Serum samples prior to and after the injection were analyzed for circulating SEAP activities and the results are shown in Figure 8. Results indicate that (1) the Ad35 constructs are all capable of expressing the SEAP transgene and that (2) the introduction of Ad5Orf6 sequence where the deleted Ad35E4 was did not significantly affect the transgene expression relative to Ad35 Δ E1SEAP. Ad35 Δ E1SEAP Δ E3 Δ E4Ad5Orf6 also yielded a similar expression profile as Ad35 Δ E1SEAP. The levels of SEAP in the serum dropped after day 2 and were at background levels by day 12.

The second mouse experiment evaluates the effect of a full Ad5E4 replacement instead of an Ad5Orf6 substitution for the Ad35 E4 cassette. Here, cohorts of 5 C3H/HeN mice were given single intramuscular injections of one of the following vectors: (1) 10^{10} vp MRKAd5-SEAP; (2) 10^9 vp MRKAd5-SEAP; (3) 10^{10} vp Ad35 Δ E1SEAP Δ E4Ad5Orf6; (4) 10^{10} vp Ad35 Δ E1SEAP Δ E4Ad5E4; or (5) 10^{10} vp Ad35 Δ E1SEAP Δ E4Ad5PE4. The introduction of Ad5E4 or Ad5PE4 resulted in comparable if not, slightly improved expression levels compared to the vector with the Ad5Orf6 insertion (Figure 9). The peak levels for the Ad35 constructs are lower than those produced by Ad5SEAP (at least 10-fold).

D. Primate Results

Cohorts of 3 rhesus macaques were given single intramuscular injections of one of the following vectors: (1) 10^{11} vp MRKAd5-SEAP; (2) 10^9 vp MRKAd5-SEAP; or (3) 10^{11} vp Ad35 Δ E1SEAP Δ E4Ad5Orf6. Serum samples prior to and after the injection were analyzed for circulating SEAP activities and the results for the individual monkeys are shown in Figures 10A-B. Results indicate that the peak level of SEAP product produced by the alternative adenovirus serotype was lower than but were within 3-fold of that of MRKAd5SEAP at the same

high dose level of 10^{11} vp. The levels observed from the Ad35 vector were about 50-fold higher than those observed using 10^9 vp of MRKAd5SEAP. The levels of SEAP in the serum dropped after day 10 and were close to background as early as day 15.

A separate experiment using African green monkeys was conducted to examine the effect of the additional E3 deletion or the full Ad5E4 substitution on in vivo gene expression. In here, cohorts of 2-3 African green macaques were given single intramuscular injections of one of the following vectors: (1) 10^{11} vp MRKAd5-SEAP; (2) 10^{10} vp MRKAd5-SEAP; (3) 10^9 vp MRKAd5-SEAP; (4) 10^{10} vp Ad35 Δ E1SEAP Δ E4Ad5Orf6; (5) 10^{10} vp Ad35 Δ E1SEAP Δ E3 Δ E4Ad5Orf6; or (6) 10^{10} vp Ad35 Δ E1SEAP Δ E4Ad5E4. Results (Figure 11) indicate that the peak levels of SEAP product produced by Ad35 Δ E1SEAP Δ E3 Δ E4Ad5Orf6 and Ad35 Δ E1SEAP Δ E4Ad5E4 were comparable if not, slightly improved compared to Ad35 Δ E1SEAP Δ E4Ad5Orf6.

EXAMPLE 6

In vivo Immunogenicity

A. Immunization

Cohorts of 3-6 animals were given intramuscular injections at wk 0 and wk 4 of either of the following constructs: (1) 10^{11} vp MRKAd5-HIV1 gag; or (2) 10^{11} vp of Ad35 Δ E1gag Δ E4Ad5Orf6. Rhesus macaques were between 3-10 kg in weight. In all cases, the total dose of each vaccine was suspended in 1 mL of buffer. The macaques were anesthetized (ketamine/xylazine) and the vaccines were delivered i.m. in 0.5-mL aliquots into both deltoid muscles using tuberculin syringes (Becton-Dickinson). Sera and peripheral blood mononuclear cells (PBMC) were prepared from blood samples collected at several time points during the immunization regimen. All animal care and treatment were in accordance with standards approved by the Institutional Animal Care and Use Committee according to the principles set forth in the *Guide for Care and Use of Laboratory Animals*, Institute of Laboratory Animal Resources, National Research Council.

B. ELISPOT Assay

The IFN- γ ELISPOT assays for rhesus macaques were conducted following a previously described protocol (Allen *et al.*, 2001 *J. Virol.* 75(2):738-749), with some modifications. For antigen-specific stimulation, a peptide pool was prepared from 20-aa peptides that encompass the entire HIV-1 gag sequence with 10-aa overlaps (Synpep Corp., Dublin, CA). To each well, 50 μ L of $2-4 \times 10^5$ peripheral blood mononuclear cells (PBMCs)

were added; the cells were counted using Beckman Coulter Z2 particle analyzer with a lower size cut-off set at 80 femtoliters ("fL"). Either 50 μ L of media or the gag peptide pool at 8 μ g/mL concentration per peptide was added to the PBMC. The samples were incubated at 37°C, 5% CO₂ for 20-24 hrs. Spots were developed accordingly and the plates were processed using custom-built imager and automatic counting subroutine based on the ImagePro platform (Silver Spring, MD); the counts were normalized to 10⁶ cell input.

C. Intracellular Cytokine Staining

To 1 ml of 2 x 10⁶ PBMC/mL in complete RPMI media (in 17x100mm round bottom polypropylene tubes (Sarstedt, Newton, NC)), anti-hCD28 (clone L293, Becton-Dickinson) and anti-hCD49d (clone L25, Becton-Dickinson) monoclonal antibodies were added to a final concentration of 1 μ g/mL. For gag-specific stimulation, 10 μ L of the peptide pool (at 0.4 mg/mL per peptide) were added. The tubes were incubated at 37 °C for 1 hr., after which 20 μ L of 5 mg/mL of brefeldin A (Sigma) were added. The cells were incubated for 16 hr at 37 °C, 5% CO₂, 90% humidity. 4 mL cold PBS/2%FBS were added to each tube and the cells were pelleted for 10 min at 1200 rpm. The cells were re-suspended in PBS/2%FBS and stained (30 min, 4 °C) for surface markers using several fluorescent-tagged mAbs: 20 μ L per tube anti-hCD3-APC, clone FN-18 (Biosource); 20 μ L anti-hCD8-PerCP, clone SK1 (Becton Dickinson, Franklin Lakes, NJ); and 20 μ L anti-hCD4-PE, clone SK3 (Becton Dickinson). Sample handling from this stage was conducted in the dark. The cells were washed and incubated in 750 μ L 1xFACS Perm buffer (Becton Dickinson) for 10 min at room temperature. The cells were pelleted and re-suspended in PBS/2%FBS and 0.1 μ g of FITC-anti-hIFN- γ , clone MD-1 (Biosource) was added. After 30 min incubation, the cells were washed and re-suspended in PBS. Samples were analyzed using all four color channels of the Becton Dickinson FACSCalibur instrument. To analyze the data, the low side- and forward-scatter lymphocyte population was initially gated; a common fluorescence cut-off for cytokine-positive events was used for both CD4⁺ and CD8⁺ populations, and for both mock and gag-peptide reaction tubes of a sample.

D. Results

PBMCs collected at regular 4-wk intervals were analyzed in an ELISPOT assay. Results (Table 1) indicate that the Ad35 Δ E1gag Δ E4Ad5Orf6 is able to induce in non-human primates significant levels of gag-specific T cells. After a single dose (wk 4), the Ad35-induced responses were about 5-fold lower than that of MRKAd5-HIV1 gag. After the second dose (wk

8), the responses between both cohorts were comparable; the differences became pronounced in the succeeding time points.

5 Table 1. Gag-specific T cell response in monkeys immunized with MRKAd5-HIV1 gag and Ad35ΔE1gagΔE4Ad5Orf6. Shown is the number of spot-forming cells per million PBMC following incubation in the absence (mock) or presence of Gag H peptide pool. The H pool consisted of 20-aa peptide overlapping by 10 aa and encompassing the entire gag sequence.

Grp	Vaccine Wk 0, Wk 4	Monkey ID	Pre		Wk 4		Wk 8		Wk 12		Wk 16	
			Mock	Gag H	Mock	Gag H	Mock	Gag H	Mock	Gag H	Mock	Gag H
1	MRKAd5-HIV1 gag 10 ¹¹ vp	00C018	1	5	13	1025	0	824	3	753	1	533
		00C034	0	4	5	219	5	404	0	491	1	350
		00C058	4	4	3	1086	0	440	0	439	0	599
2	Ad35ΔE1gagΔE4Ad5Orf6 10 ¹¹ vp	00D045	1	1	3	168	5	645	4	178	0	91
		00D067	1	4	5	89	0	103	0	76	0	19
		00D068	1	4	10	34	5	365	3	143	0	95
		00D054	3	15	10	195	0	501	3	350	0	124
		00D075	3	5	18	275	13	716	3	158	0	103
		00D073	14	26	1	241	3	485	3	278	0	148
3	Naïve	00D087	1	1	3	3	8	54	3	5	3	1

10 Intracellular IFN-γ staining analyses of PBMC collected at wk 8 suggest that the Ad35-based vaccine is able to induce both HIV-specific CD4+ and CD8+ T cells (Table 2).

15 Table 2. Characterization of the gag-specific T cells in monkeys immunized with MRKAd5-HIV1 gag and Ad35ΔE1gagΔE4Ad5Orf6. Shown are the percentages of CD3+ T cells that are either gag-specific CD4+ or gag-specific CD8+ cells. These values were corrected for mock values (<0.02%).

Grp	Vaccine Wk 0, Wk 4	Monkey ID	Wk 8	
			%CD4+CD3+	%CD8+CD3+
1	MRKAd5-HIV1 gag 10 ¹¹ vp	00C018	0.08	0.37
		00C034	0.09	0.06
		00C058	0.03	0.21
2	Ad35ΔE1gagΔE4Ad5Orf6 10 ¹¹ vp	00D045	0.06	0.08
		00D067	0.02	0.02
		00D068	0.15	0.02
		00D054	0.05	0.08
		00D075	0.08	0.05
		00D073	0.09	0.06

20 In a separate experiment, 3 different Ad35 constructs expressing HIV-1 gag were evaluated for their immunogenicity in macaques. Here, cohorts of 3 macaques were given immunizations at wk 0 and 4 of either of the following vectors: (1) 10¹⁰ vp Ad35ΔE1gagΔE4Ad5Orf6; (2) 10¹⁰

vp Ad35ΔE1gagΔE3ΔE4Ad5Orf6; or (3) 10^{10} vp Ad35ΔE1gagΔE4Ad5E4. The levels of T cell immunity induced by all 3 vectors were comparable at this stage (Table 2), suggesting that the additional E3 deletion or full Ad5E4 substitution does not appear to impair the immunogenic properties of the vector.

5

Table 3. Gag-specific T cell response in monkeys immunized with several Ad35ΔE1ΔE4-based vectors. Shown is the number of spot-forming cells per million PBMC following incubation in the absence (mock) or presence of Gag H peptide pool. The H pool consisted of 20-aa peptide overlapping by 10 aa and encompassing the entire gag sequence.

10

Grp	Vaccine Wk 0, Wk 4	Monkey ID	Pre		Wk 4		Wk 8	
			Mock	Gag H	Mock	Gag H	Mock	Gag H
1	Ad35ΔE1gagΔE4Ad5Orf6 10^{10} vp	00C047	4	1	0	20	0	189
		00C157	8	5	1	81	1	833
		00C078	3	1	0	46	4	349
2	Ad35ΔE1gagΔE3ΔE4Ad5Orf6 10^{10} vp	00C091	1	1	1	118	3	315
		00C122	3	0	0	31	1	138
		00D177	3	3	1	45	1	64
3	Ad35ΔE1gagΔE4Ad5E4 10^{10} vp	00D018	3	19	29	120	23	193
		00D046	8	5	1	21	10	143
		00D063	3	4	0	63	4	371
Naïve	none	00D363	0	5	ND	ND	0	0

EXAMPLE 7

Construction and Rescue of pAd24ΔE1.

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An E1- Ad24-based pre-adenovirus plasmid was constructed in order to determine whether an E1- Ad24 vector (a representative group D serotype) could be propagated in an Ad5/group C E1-complementing cell line. Since at the time the vector construction was initiated the complete sequence of Ad24 (*see* Figures 16A-1 through 16A-10; subject of copending application serial no. 60/455, 312, filed March 17, 2003) was unknown we took advantage of some sequence homology between Ad24 and Ad17. The general strategy used to recover Ad24 as a bacterial plasmid is illustrated in Figure 12 and described below.

20

Cotransformation of BJ5183 bacteria with purified wild-type Ad24 viral DNA and a second DNA fragment termed the Ad17 ITR cassette resulted in the circularization of the viral genome by homologous recombination. The ITR cassette contains sequences from the right (bp 34469 to 35098) and left (bp 4 to 414 and bp 3373 to 4580) end of the Ad17 genome (Accession No. AF108105) separated by plasmid sequences containing a bacterial origin of replication and an Ampicillin resistance gene. The ITR cassette contains a deletion of E1 sequences from Ad17

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(bp 415 to 3372) with a unique *Swa* I site located in the deletion. The Ad17 sequences in the ITR cassette provide regions of homology with the purified Ad24 viral DNA in which recombination can occur. The ITR cassette was also designed to contain unique restriction enzyme sites (*Pme* I) located at the end of the viral ITR's so that digestion will release the Ad24 genome from plasmid sequences. Potential clones were screened by restriction analysis and one clone was selected as pAd24ΔE1. pAd24ΔE1 contains Ad17 sequences from bp 4 to 414 and from bp 3373 to 4580, Ad24 bp 4588 to 34529, and Ad17 bp 34469 to 35098 (bp numbers refer to the wt sequence for both Ad17 and Ad24). PAd24ΔE1 contains the coding sequences for all Ad24 virion structural proteins that constitute its serotype specificity. This approach can be used to circularize any group D serotype into plasmid form which has sufficient homology to Ad17.

To determine if pre-adenovirus plasmid pAd24ΔE1 could be rescued into virus and propagated in a group C E1 complementing cell line, the plasmid was digested with *Pme* I and transfected into a 6 cm dish of 293 cells using the calcium phosphate co-precipitation technique. *Pme* I digestion releases the viral genome from the plasmid sequences allowing viral replication to occur after entry into 293 cells. Viral cytopathic effect (CPE), indicating that virus replication and amplification is occurring, was very slow to arise. Following multiple attempts, we were successful at rescuing and amplifying Ad24ΔE1 but the virus grew to lower titers and took more passages to amplify than a similar Ad5 based vector. In order to verify the genetic structure of the virus, viral DNA was extracted using pronase treatment followed by phenol chloroform extraction and ethanol precipitation. Viral DNA was then digested with *Hind*III and treated with Klenow fragment to end-label the restriction fragments with P33-dATP. The end-labeled restriction fragments were then size-fractionated by gel electrophoresis and visualized by autoradiography. The digestion products were compared with the digestion products from the pre-plasmid (that had been digested with *Pme*I/*Hind*III prior to labeling). The expected sizes were observed, indicating that the virus had been successfully rescued.

EXAMPLE 8

Insertion of Ad5 Orf 6 into the E1 region of Ad24

In order to determine if the insertion of Ad5 E4 Orf6 into the Ad24 genome would allow more efficient propagation in a group C E1 complementing cell line we constructed an Ad24 based pre-adenovirus plasmid containing Ad5 Orf6 in the E1 region. In order to introduce Ad5 Orf6 in to the E1 region of pAd24ΔE1, bacterial recombination was used. An Ad5 Orf6 transgene consisting of the Ad5 Orf6 coding region flanked by the HCMV promoter and pA was cloned into the E1 deletion in an Ad17 shuttle vector (a precursor to the Ad17 ITR cassette). The Ad5 Orf6 transgene was cloned between bp 414 and 3373 in the E1 anti-parallel

orientation. The shuttle vector containing the Ad5 Orf6 transgene was digested to generate a DNA fragment consisting of the transgene flanked by Ad17 sequences (bp 4 to 414 and bp 3373 to 4580) and the fragment was purified after electrophoresis on an agarose gel.

Cotransformation of BJ 5183 bacteria with the shuttle vector fragment and pAd24ΔE1, which
5 had been linearized in the E1 region by digestion with *Swa*I, resulted in the generation of pAd24ΔE1Ad5Orf6 by homologous recombination (Figure 13). Potential clones were screened by restriction analysis and one clone was selected as pre-adenovirus plasmid pAd24ΔE1Ad5Orf6.

In order to determine if pre-adenovirus plasmid pAd24ΔE1Ad5Orf6 could be
10 rescued into virus and propagated in an Ad5/group C E1 complementing cell line, pAd24ΔE1Ad5Orf6 was digested with *Pme* I and transfected into a 6 cm dish of 293 cells using the calcium phosphate co-precipitation technique. *Pme*I digestion releases the viral genome from plasmid sequences allowing viral replication to occur after entry into 293 cells. Once
15 complete viral cytopathic effect (CPE) was observed at approximately 7-10 days post transfection, the infected cells and media were freeze/thawed three times and the cell debris pelleted. The virus was amplified in two additional passages in 293 cells and then purified from the final infection by ultracentrifugation on CsCl density gradients. In order to verify the genetic
20 structure of the virus, viral DNA was extracted using pronase treatment followed by phenol chloroform extraction and ethanol precipitation. Viral DNA was then digested with *Hind*III and treated with Klenow fragment to end-label the restriction fragments with P33-dATP. The end-labeled restriction fragments were then size-fractionated by gel electrophoresis and visualized by
25 autoradiography. The digestion products were compared with the digestion products from the pre-plasmid (that had been digested with *Pme*I/*Hind*III prior to labeling). The expected sizes were observed, indicating that the virus had been successfully rescued.

EXAMPLE 9

Insertion of Ad5 Orf 6 into the E4 region of Ad24

To refine the strategy of including Ad5 Orf6 in the genome of an alternative serotype so that propagation could take place in an Ad5/group C complementing cell line two
30 additional strategies were developed. In the first strategy, the entire alternative serotype E4 region (not including the E4 promoter) was deleted and replaced with Ad5 Orf6. In the second strategy, just the alternative serotype Orf6 gene was deleted and replaced with Ad5 Orf6. The configuration of the E4 regions generated by the two strategies is diagramed in Figure 14. For each of these strategies the desired pre-Adenovirus plasmid was generated by bacterial
35 recombination. Cotransformation of BJ 5183 bacteria with pAd24ΔOrf6BstZ17I and the

appropriately constructed Ad24 E4 shuttle plasmid resulted in the generation of the desired Ad24 based pre-Ad plasmid. PAd24 Δ Orf6BstZ17I, a derivative of pAd24 Δ E1, was constructed so that the E4 region in the Ad24 pre-Ad plasmid could be easily modified using bacterial recombination. PAd24 Δ Orf6BstZ17I contains a deletion in the E4 region from Ad24 bp 32373 to bp 33328 with a unique BstZ17I site located at the position of the deletion. The complete sequence of pAd24 Δ Orf6BstZ17I consists of Ad17 sequences from bp 4 to 414 and from bp 3373 to 4580, Ad24 bp 4588 to 32372 and from 33329 to 34529, and Ad17 bp 34469 to 35098 (bp numbers refer to the wt sequence for both Ad17 and Ad24).

To construct pAd24 Δ E1 Δ E4Ad5Orf6 (An Ad24 pre-Ad plasmid containing an E1 deletion and a deletion of E4 substituted with Ad5 Orf6), an Ad24 E4 shuttle plasmid was constructed by digesting pAd24 Δ E1 with *PmeI* and *BsrGI* and cloning the restriction fragment representing the E4 region (bp 31559 to bp 35164) into pNEB193, generating pNEBAd24E4. PNEBAd24E4 was then digested with *AccI* and *EcoNI* to remove the E4 coding sequences and ligated with an oligo designed to contain *BglII* and *XhoI* sites (underlined) (5' ACTCGAGATGTATAGATCT (SEQ ID NO: 6); 5' CTAGATCTATACATCTCGAG (SEQ ID NO: 7)), generating pNEBAd24 Δ E4. PNEBAd24 Δ E4 was then digested with *BglII* and *XhoI* and ligated with the Ad5 Orf6 gene, which was PCR amplified, generating pNEBAd24 Δ E4Ad5Orf6. The PCR primers used to amplify the Ad5 Orf6 gene (5' GCACAGATCTTTGCTTCAGGAATATG (SEQ ID NO: 8); 5' GAGAACTCGAGGCCTACATGGGGGTAGAG (SEQ ID NO: 9)) were designed to contain *BglII* and *XhoI* sites (underlined above) for ligation with the pNEBAd24 Δ E4 fragment. In the final step pNEBAd24 Δ E4Ad5Orf6 E4 shuttle plasmid was digested with *PvuI* and *PmeI*, the restriction fragments were size fractionated by agarose gel electrophoresis and the desired fragment containing Ad5Orf6 flanked by Ad24 sequences was gel purified. Cotransformation of BJ 5183 bacteria with E4 shuttle fragment and pAd24 Δ Orf6BstZ17I, which had been linearized in the E4 region by digestion with BstZ17I, resulted in the generation of pAd24 Δ E1 Δ E4Ad5Orf6 by homologous recombination. Potential clones were screened by restriction analysis and one clone was selected as pre-adenovirus plasmid pAd24 Δ E1 Δ E4Ad5Orf6.

To construct pAd24 Δ E1 Δ Orf6Ad5Orf6 (An Ad24 pre-Ad plasmid containing an E1 deletion and a deletion of E4 Orf6 substituted with Ad5 Orf6), an Ad24 E4 shuttle plasmid was constructed in which the Ad24 Orf6 gene was replaced by Ad5 Orf6. To do this the *EcoRI* restriction fragment representing bp 32126 to bp 33442 of the Ad24 genome (encompassing the E4 Orf6 coding region), was subcloned into the *EcoRI* site in pNEB193, generating pNEBAd24Orf6. In order to delete the E4 Orf6 gene in pNEBAd24Orf6 and replace it with Ad5 Orf6, pNEBAd24Orf6 was digested with *SylI* and treated with Klenow to blunt the ends and then

digested with to *EagI*. The desired pNEBAd24Orf6 fragment was then ligated with a PCR product representing the Ad5 Orf6 gene from Ad5 bp 33193 to bp 24125, generating pNEBAd24ΔOrf6Ad5Orf6. The PCR primers used to generate the Ad5 Orf6 fragment (5'CGAGACCGGCCGACGCAGATCTGTTTG (SEQ ID NO: 10);

5 5'GAAGTCCCCGGGCTACATGGGGGTTAG (SEQ ID NO: 11)) were designed to contain *EagI* and *SmaI* sites (underlined above) for ligation with the pNEBAd24Orf6 fragment. In the final step pNEBAd24ΔOrf6Ad5Orf6 was digested with *EcoRI*, the restriction fragments were size fractionated by agarose gel electrophoresis and the desired fragment containing Ad5Orf6 flanked by Ad24 sequences was gel purified. Cotransformation of BJ 5183 bacteria with the *EcoRI* fragment and pAd24ΔOrf6BstZ17I, which had been linearized in the E4 region by digestion with *BstZ17I*, resulted in the generation of pAd24ΔE1ΔOrf6Ad5Orf6 by homologous recombination. Potential clones were screened by restriction analysis and one clone was selected as pre-adenovirus plasmid pAd24ΔE1ΔOrf6Ad5Orf6.

15 EXAMPLE 10

Rescue of pAd24ΔE1ΔE4Ad5Orf6, pAd24ΔE1ΔOrf6Ad5Orf6, into Virus

In order to determine if pre-adenovirus plasmids pAd24ΔE1ΔE4Ad5Orf6, pAd24ΔE1ΔOrf6Ad5Orf6, could be rescued into virus and propagated in a group C E1 complementing cell line, the plasmids were each digested with *PmeI* and transfected into T-25
20 flasks of PER.C6 cells using the calcium phosphate co-precipitation technique; (Cell Pfect Transfection Kit, Amersham Pharmacia Biotech Inc.). *PmeI* digestion releases the viral genome from plasmid sequences allowing viral replication to occur after cell entry. Viral cytopathic effect (CPE), indicating that virus replication and amplification was occurring, was observed for both constructs. When CPE was complete, approximately 7-10 days post transfection, the
25 infected cells and media were harvested, freeze/thawed three times and the cell debris pelleted by centrifugation. Approximately 1 ml of the cell lysate was used to infect T-225 flasks of PER.C6 cells at 80-90% confluence. Once CPE was reached, infected cells and media were harvested, freeze/thawed three times and the cell debris pelleted by centrifugation. Clarified cell lysates were then used to infect 2-layer NUNC cell factories of PER.C6 cells. Following
30 complete CPE the virus was purified by ultracentrifugation on CsCl density gradients. In order to verify the genetic structure of the rescued viruses, viral DNA was extracted using pronase treatment followed by phenol chloroform extraction and ethanol precipitation. Viral DNA was then digested with *HindIII* and treated with Klenow fragment to end-label the restriction fragments with P33-dATP. The end-labeled restriction fragments were then size-fractionated by
35 gel electrophoresis and visualized by autoradiography. The digestion products were compared

with the digestion products of the corresponding pre-Adenovirus plasmid (that had been digested with *Pme*I/*Hind*III prior to labeling) from which they were derived. The expected sizes were observed, indicating that the viruses had been successfully rescued.

5 EXAMPLE 11

Comparison of the Growth Kinetics of Ad24 based vectors.

In order to compare the growth kinetic of Ad24ΔE1, Ad24ΔE1Ad5Orf6, Ad24ΔE1ΔE4Ad5Orf6 and Ad24ΔE1ΔOrf6Ad5Orf6 one step growth curves were preformed (Figure 15). PER.C6 cells in 60 mm dishes were infected at 1 vp per cell with either Ad24ΔE1, Ad24ΔE1Ad5Orf6, Ad24ΔE1ΔE4Ad5Orf6 or Ad24ΔE1ΔOrf6Ad5Orf6. Cells and media were then harvested at various times post infection, freeze thawed three times and clarified by centrifugation. The amount of virus present in the samples was determined by quantitative PCR and is illustrated in Figure 15. This study demonstrates that Ad24 vectors that incorporate Ad5 Orf6 have a distinct growth advantage over Ad24ΔE1 in PER.C6 cells. The instant invention can be practiced with recombinant Ad24 vectors absent a heterologous Orf 6 region where the E1-complementing cell line expresses an Ad24 E1 region or, alternatively, E1 and E4 regions of the same serotype (such as Ad5E1/E4-expressing cell lines).

EXAMPLE 12

Insertion of an Expression Cassette into pAd24ΔE1ΔE4Ad5Orf6, pAd24ΔE1ΔOrf6Ad5Orf6,

In order to introduce a gag or SEAP expression cassette (see Figures 6 and 7, respectively) into the E1 region of the Ad24 pre-Adenovirus plasmids described above (pAd24ΔE1ΔE4Ad5Orf6, pAd24ΔE1ΔOrf6Ad5Orf6) bacterial recombination was used. A gag expression cassette consisting of the following: 1) the immediate early gene promoter from the human cytomegalovirus, 2) the coding sequence of the human immunodeficiency virus type 1 (HIV-1) gag (strain CAM-1; 1526 bp) gene, and 3) the bovine growth hormone polyadenylation signal sequence, was cloned into the E1 deletion in Ad17 shuttle plasmid, pABSAd17-3, generating pABSAd17HCMVgagBGHpA. The ITR cassette contains sequences from the right (bp 34469 to 35098) and left (bp 4 to 414 and bp 3373 to 4580) end of the Ad17 genome separated by plasmid sequences containing a bacterial origin of replication and an Ampicillin resistance gene. The ITR cassette contains a deletion of E1 sequences from Ad17 (bp 415 to 3372) with a unique *Swa* I site located in the deletion. The gag expression cassette was obtained from a previously constructed shuttle plasmid by *Eco*RI digestion. Following the digestion the desired fragment was gel purified, treated with Klenow to obtain blunt ends and cloned into the *Swa*I site in pABSAd17-3. This cloning step resulted in the gag expression cassette being

cloned into the E1 deletion between bp 414 and 3373 in the E1 parallel orientation. The shuttle vector containing the gag transgene was digested to generate a DNA fragment consisting of the gag expression cassette flanked by Ad17 bp 4 to 414 and bp 3373 to 4580 and the fragment was purified after electrophoresis on an agarose gel. Cotransformation of BJ 5183 bacteria with the shuttle vector fragment and one of the Ad24 pre-Ad plasmids (pAd24ΔE1ΔE4Ad5Orf6, pAd24ΔE1ΔOrf6Ad5Orf6), linearized in the E1 region by digestion with *Swa* I, resulted in the generation of the corresponding Ad24 gag-containing pre-Adenovirus plasmids (pAd24ΔE1gagΔE4Ad5Orf6, pAd24ΔE1gagΔOrf6Ad5Orf6) by homologous recombination. Potential clones were screened by restriction analysis.

A similar strategy was used to generate Ad24 pre-Ad plasmids containing a SEAP expression cassette. In this case a SEAP expression cassette consisting of: 1) the immediate early gene promoter from the human cytomegalovirus, 2) the coding sequence of the human placental SEAP gene, and 3) the bovine growth hormone polyadenylation signal sequence was cloned into the E1 deletion in Ad17 shuttle plasmid, pABSAd17-3, generating pABSAd17HCMVSEAPBGH. The SEAP expression cassette was obtained from a previously constructed shuttle plasmid by *Eco*RI digestion. Following the digestion the desired fragment was gel purified, treated with Klenow to obtain blunt ends and cloned into the *Swa*I site in pABSAd17-3. The shuttle vector containing the SEAP transgene was digested to generate a DNA fragment consisting of the SEAP expression cassette flanked by Ad17 bp 4 to 414 and bp 3373 to 4580 and the fragment was purified after electrophoresis on an agarose gel. Cotransformation of BJ 5183 bacteria with the shuttle vector fragment and one of the Ad24 pre-Ad plasmids (pAd24ΔE1ΔE4Ad5Orf6, pAd24ΔE1ΔOrf6Ad5Orf6), linearized in the E1 region by digestion with *Swa* I, resulted in the generation of the corresponding Ad24 SEAP-containing pre-Adenovirus plasmids (pAd24ΔE1SEAPΔE4Ad5Orf6, pAd24ΔE1SEAPΔOrf6Ad5Orf6) by homologous recombination. Potential clones were screened by restriction analysis. All pre-Ad plasmids were rescued into virus and expanded to prepare CsCl purified stocks as described above.

EXAMPLE 13

In Vivo Immunogenicity

A. Immunization

Cohorts of 3-6 animals were given intramuscular injections at wk 0 and wk 4 of either of the following constructs: (1) 10^{11} vp MRKAd5-HIV1 gag; (2) 10^{10} vp MRKAd5-HIV1 gag; (3) 10^{11} vp of Ad24ΔE1gagΔOrf6Ad5Orf6; (4) 10^{10} vp of

Ad24ΔE1gagΔOrf6Ad5Orf6; or (5) 10^{10} vp of Ad24ΔE1gagΔE4Ad5Orf6. Rhesus macaques were between 3-10 kg in weight. In all cases, the total dose of each vaccine was suspended in 1 mL of buffer. The macaques were anesthetized (ketamine/xylazine) and the vaccines were delivered i.m. in 0.5-mL aliquots into both deltoid muscles using tuberculin syringes (Becton-Dickinson, Franklin Lakes, NJ). Peripheral blood mononuclear cells (PBMC) were prepared from blood samples collected at several time points (typically 4 wk intervals) during the immunization regimen. All animal care and treatment were in accordance with standards approved by the Institutional Animal Care and Use Committee according to the principles set forth in the *Guide for Care and Use of Laboratory Animals*, Institute of Laboratory Animal Resources, National Research Council.

B. ELISPOT Assay

The IFN- γ ELISPOT assays for rhesus macaques were conducted following a previously described protocol (Allen et al., 2001 *J. Virol.* 75(2):738-749; Casimiro et al., 2002 *J. Virol.* 76:185-94), with some modifications. For antigen-specific stimulation, a peptide pool was prepared from 20-aa peptides that encompass the entire HIV-1 gag sequence with 10-aa overlaps (Synpep Corp., Dublin, CA). To each well, 50 μ L of $2-4 \times 10^5$ peripheral blood mononuclear cells (PBMCs) were added; the cells were counted using Beckman Coulter Z2 particle analyzer with a lower size cut-off set at 80 femtoliters ("fL"). Either 50 μ L of media or the gag peptide pool at 8 μ g/mL concentration per peptide was added to the PBMC. The samples were incubated at 37°C, 5% CO₂ for 20-24 hrs. Spots were developed accordingly and the plates were processed using custom-built imager and automatic counting subroutine based on the ImagePro platform (Silver Spring, MD); the counts were normalized to 10^6 cell input.

C. Intracellular Cytokine Staining

To 1 ml of 2×10^6 PBMC/mL in complete RPMI media (in 17x100mm round bottom polypropylene tubes (Sarstedt, Newton, NC)), anti-hCD28 (clone L293, Becton-Dickinson) and anti-hCD49d (clone L25, Becton-Dickinson) monoclonal antibodies were added to a final concentration of 1 μ g/mL. For gag-specific stimulation, 10 μ L of the peptide pool (at 0.4 mg/mL per peptide) were added. The tubes were incubated at 37 °C for 1 hr., after which 20 μ L of 5 mg/mL of brefeldin A (Sigma) were added. The cells were incubated for 16 hr at 37 °C, 5% CO₂, 90% humidity. 4 mL cold PBS/2%FBS were added to each tube and the cells were pelleted for 10 min at 1200 rpm. The cells were re-suspended in PBS/2%FBS and stained (30 min, 4 °C) for surface markers using several fluorescent-tagged mAbs: 20 μ L per tube anti-hCD3-APC, clone FN-18 (Biosource); 20 μ L anti-hCD8-PerCP, clone SK1 (Becton Dickinson);

and 20 μ L anti-hCD4-PE, clone SK3 (Becton Dickinson). Sample handling from this stage was conducted in the dark. The cells were washed and incubated in 750 μ L 1xFACS Perm buffer (Becton Dickinson) for 10 min at room temperature. The cells were pelleted and re-suspended in PBS/2%FBS and 0.1 μ g of FITC-anti-hIFN- γ , clone MD-1 (Biosource) was added. After 30 min incubation, the cells were washed and re-suspended in PBS. Samples were analyzed using all four color channels of the Becton Dickinson FACSCalibur instrument. To analyze the data, the low side- and forward-scatter lymphocyte population was initially gated; a common fluorescence cut-off for cytokine-positive events was used for both CD4⁺ and CD8⁺ populations, and for both mock and gag-peptide reaction tubes of a sample.

D. Anti-p24 ELISA

A modified competitive anti-p24 assay was developed using reagents from the Coulter p24 Antigen Assay kit (Beckman Coulter, Fullerton, CA). Briefly, to a 250- μ L serum sample, 20 μ L of Lyse Buffer and 15 μ L of p24 antigen (9.375 pg) from the Coulter kit were added. After mixing, 200 μ L of each sample were added to wells coated with a mouse anti-p24 mAb from the Coulter kit and incubated for 1.5 hr at 37°C. The wells were then washed and 200 μ L of Biotin Reagent (polyclonal anti-p24-biotin) from the Coulter kit was added to each well. After a 1 hr, 37°C incubation, detection was achieved using streptavidin-conjugated horseradish peroxidase and TMB substrate as described in the Coulter Kit. OD_{450nm} values were recorded. A 7-point standard curve was generated using a serial 2-fold dilution of serum from an HIV-seropositive individual. The lower cut-off for the assay is arbitrarily set at 10 milli Merck units/mL (mMU/mL) defined by a dilution of the seropositive human serum. This cutoff falls at approximately 65% of the maximum bound control signal which corresponds to that obtained with the diluent control only and with no positive analyte.

E. Results

PBMCs collected at regular 4-wk intervals were analyzed in an ELISPOT assay (Figure 17). Both Ad24 Δ E1gag Δ Orf6Ad5Orf6 and Ad24 Δ E1gag Δ E4Ad5Orf6 were able to induce significant levels of gag-specific T cells in non-human primates. At 10¹¹ vp dose level, the Ad24-induced responses were within 2-3-fold of those of MRKAd5-HIV1 gag. Both Ad24 vectors were also able to induce detectable levels of gag-specific T cells at 10¹⁰ vp but were lower than those observed using MRKad5gag at the same dose.

PBMCs collected at wk 12 from the vaccinees were analyzed for intracellular IFN- γ staining after the priming immunizations. The assay results provided information on the relative amounts of CD4⁺ and CD8⁺ gag-specific T cells in the peripheral blood (Figure 18). The

results indicated that the prime-boost immunization approach was able to elicit in rhesus macaques both HIV-specific CD4⁺ and CD8⁺ T cells.

F. Humoral Immune Responses

The Ad24-based vaccine vector was able to generate detectable levels of circulating anti-gag antibodies at the reasonably high dose level (Figure 19). No detectable titers were observed at equal to or lower than 10¹⁰ vp, suggesting the existence of a dose-dependent response.

EXAMPLE 14

In Vivo Transgene Expression

A. Immunization

Cohorts of 5 C3H/HeN mice were given single intramuscular injections of one of the following vectors: (1) 10¹⁰ vp Ad24ΔE1SEAPΔE4Ad5Orf6; (2) 10¹⁰ vp Ad24ΔE1SEAPΔOrf6Ad5Orf6; (3) 10¹⁰ vp MRKAd5SEAP; and (4) 10⁹ vp MRKAd5SEAP. Female mice were between 4-10 weeks old. The total dose of each vaccine was suspended in 0.1 mL of buffer. The vectors were given to both quadriceps of each of the animals with a volume of 50 uL per quad and using 0.3-mL 28G1/2 insulin syringes (Becton-Dickinson, Franklin Lakes, NJ). For the primates, the total dose of each vaccine was suspended in 1 mL of buffer. The monkeys were anesthetized (ketamine/xylazine mixture) and the vaccines were delivered i.m. in 0.5-mL aliquots into two muscle sites using tuberculin syringes (Becton-Dickinson, Franklin Lakes, NJ). Serum samples were collected at defined intervals and stored frozen until the assay date. All animal care and treatment were in accordance with standards approved by the Institutional Animal Care and Use Committee according to the principles set forth in the Guide for Care and Use of Laboratory Animals, Institute of Laboratory Animal Resources, National Research Council.

B. SEAP Assay

Serum samples were analyzed for circulating SEAP levels using TROPIX phospho-light chemiluminescent kit (Applied Biosystems Inc). Duplicate 5 uL aliquots of each serum were mixed with 45 uL of kit-supplied dilution buffer in a 96-well white DYNEX plate. Serially diluted solutions of a human placental alkaline phosphatase (Catalog no. M5905, Sigma, St. Louis, MO) in 10% naïve monkey serum served to provide the standard curve. Endogenous SEAP activity in the samples was inactivated by heating the wells for 30 minutes at 65 °C.

Enzymatic SEAP activities in the samples were determined following the procedures described in the kit. Chemiluminescence readings (in relative light units) were recorded using DYNEX luminometer. RLU readings are converted to ng/mL SEAP using a log-log regression analyses.

5 C. Rodent Results

Serum samples prior to and after the injection were analyzed for circulating SEAP activities and the results are shown in Figure 20. Results indicate that (1) both Ad24 constructs are all capable of expressing the SEAP transgene in vivo to comparable levels; and that (2) the level of expression achieved using the Ad24 vectors are comparable to that of Ad5 at 10-fold lower dose. The levels of SEAP in the serum dropped dramatically after day 2 and were at background levels by day 12.

15 D. Primate Results

Cohorts of 3 rhesus macaques were given single intramuscular injections of one of the following vectors: (1) 10^{11} vp MRKAd5-SEAP; (2) 10^9 vp MRKAd5-SEAP; (3) 10^{11} vp Ad24 Δ E1SEAP Δ Orf6Ad5Orf6; or (4) 10^{11} vp Ad24 Δ E1SEAP Δ E4Ad5Orf6. Serum samples prior to and after the injection were analyzed for circulating SEAP activities and the results are shown in Figure 21.

Results indicate that the peak levels of SEAP product produced by adenovirus serotype 24 were lower than but were within 3-fold of that of MRKAd5 at the same high dose level of 10^{11} vp (Figure 21). The levels observed with adenovirus serotype 24 are generally 50-fold higher than those observed using 10^9 vp of MRKAd5. The levels of SEAP in the serum dropped dramatically after day 10 and were close to background as early as day 15. These observations strongly indicate that adenovirus serotype 24 is very efficient in expressing a transgene following intramuscular administration in a primate.

25 EXAMPLE 15

Construction of pMRKAd24 Δ E1 Δ E4Ad5Orf6

To construct pMRKAd24 Δ E1 Δ E4Ad5Orf6 (An Ad24 pre-Ad plasmid, composed entirely of Ad24 sequence and containing an E1 deletion and an E4 deletion substituted with Ad5 Orf6), an Ad24 ITR cassette was constructed containing sequences from the right (bp 31978 to 32264 and bp 34713 to 35164) and left (bp 4 to 450 and bp 3364 to 3799) end of the Ad24 genome separated by plasmid sequences containing a bacterial origin of replication and an ampicillin resistance gene. These four segments were generated by PCR and cloned sequentially into pNEB193, generating pNEBAd24-4. Next the Ad5 Orf6 open reading frame (Ad5 bp 31192 to bp 34078) was generated by PCR and cloned between Ad24 bp 32264 and 34713 generating

pNEBAd24E-Ad5Orf6 (the ITR cassette). PNEB193 is a commonly used commercially available cloning plasmid (New England Biolabs cat# N3051S) containing a bacterial origin of replication, ampicillin resistance gene and a multiple cloning site into which the PCR products were introduced. The ITR cassette contains a deletion of E1 sequences from Ad24 bp 451 to 3363 with a unique Sma I restriction site located in the deletion and an E4 deletion from Ad24 bp 32265 to 34712 into which Ad5 Orf6 was introduced in an E4 parallel orientation. In this construct Ad5 Orf6 expression is driven by the Ad24 E4 promoter. The Ad24 sequences (bp 31978 to 32264 and bp 3464 to 3799) in the ITR cassette provide regions of homology with the purified Ad24 viral DNA in which bacterial recombination can occur following cotransformation into BJ 5183 bacteria (Figure 22). The ITR cassette was also designed to contain unique restriction enzyme sites (PmeI) located at the end of the viral ITR's so that digestion will release the recombinant Ad24 genome from plasmid sequences. Potential clones will be screened by restriction analysis and one clone was selected as pMRKAd24ΔE1ΔE4Ad5Orf6. Pre-Adenovirus plasmid pMRKAd24ΔE1ΔE4Ad5Orf6 should contain Ad24 sequences from bp 4 to 450; bp 3364 to bp 32264 and bp 34713 to bp 35164 with Ad5Orf6 cloned between bp 32264 and bp 34713. The bp numbering in the above description refers to the wt sequence for both Ad24 and Ad5.

EXAMPLE 16

20 Insertion of HIV-1 gag and SEAP transgenes into pAd24ΔE1ΔE4Ad5Orf6

In order to introduce a gag or SEAP expression cassettes into the E1 region of pMRKAd24ΔE1ΔE4Ad5Orf6, bacterial recombination will be used. An HIV-1 gag expression cassette will consist of the following: 1) the immediate early gene promoter from the human cytomegalovirus, 2) the coding sequence of the human immunodeficiency virus type 1 (HIV-1) gag (strain CAM-1; 1526 bp) gene, and 3) the bovine growth hormone polyadenylation signal sequence, in the E1 deletion of an Ad24 shuttle plasmid, pNEBAd24-2 (a precursor to the Ad24 ITR cassette described above), generating pNEBAd24CMVgagBGHpa. PNEBAd24-2 contains Ad24 sequences from the left end of the genome (bp 4 to 450 and bp 3364 to 3799) that define the E1 deletion. The gag expression cassette will be obtained from a previously constructed plasmid and cloned into the E1 deletion between bp 450 and 3364 in the E1 parallel orientation. The shuttle vector containing the gag transgene will be digested to generate a DNA fragment consisting of the gag expression cassette flanked by Ad24 bp 4 to 450 and bp 3364 to 3799 and the fragment will be purified after electrophoresis on an agarose gel. Cotransformation of BJ 5183 bacteria with the shuttle vector fragment and pMRKAd24ΔE1ΔE4Ad5Orf6 which was linearized in the E1 region by digestion with SmaI, should result in the generation of Ad24 gag-

containing pre-Adenovirus plasmids pMRKAd24ΔE1gagΔE4Ad5Orf6 by homologous recombination. Potential clones will be screened by restriction analysis.

A similar strategy will be used to generate Ad24 pre-Ad plasmids containing a SEAP expression cassette. In this case, a SEAP expression cassette will consist of: 1) the immediate early gene promoter from the human cytomegalovirus, 2) the coding sequence of the human placental SEAP gene, and 3) the bovine growth hormone polyadenylation signal sequence cloned into the E1 deletion of an Ad24 shuttle plasmid, pNEBAd24-2, generating pNEBAd24CMVSEAPBGHPA. The transgene will then be recombined into pMRKAd24ΔE1ΔE4Ad5Orf6 as described above for the gag transgene.

EXAMPLE 17

In Vivo Immunogenicity

A. Immunization

Rhesus macaques were between 3-10 kg in weight. In all cases, the total dose of each vaccine was suspended in 1 mL of buffer. The macaques were anesthetized (ketamine/xylazine) and the vaccines were delivered i.m. in 0.5-mL aliquots into both deltoid muscles using tuberculin syringes (Becton-Dickinson, Franklin Lakes, NJ). Peripheral blood mononuclear cells (PBMC) were prepared from blood samples collected at several time points during the immunization regimen. All animal care and treatment were in accordance with standards approved by the Institutional Animal Care and Use Committee according to the principles set forth in the *Guide for Care and Use of Laboratory Animals*, Institute of Laboratory Animal Resources, National Research Council.

B. T Cell Responses

Ad24 Vaccine Vector as a Heterologous Booster: Cohort of 4 rhesus macaques was immunized initially with 3 doses (wk 0, 4, 26) of either 10^7 or 10^9 vp of MRKAd5-gag (see, PCT/US01/28861, published March 21, 2002) or MRKAd6-gag. At wk 56, the animals received a booster vaccine of 10^{11} vp Ad24ΔE1gagΔOrf6Ad5Orf6. A separate cohort of naïve animals received a single dose of the booster vaccine. The results of the IFN-γ ELISPOT analyses of PBMC collected during the course of the studies are shown in Figure 23. It is apparent that the Ad24 HIV vectors can be utilized to amplify the existing pools of HIV-specific T cells. The increases in the levels of gag-specific T cells from the pre-boost levels to those measured at 4 wks post boost were consistently larger than the levels induced by the same booster vaccine in naïve animals. PBMCs from the vaccinees of the heterologous MRKAd5/MRKAd6-Ad24 boost

regimen were analyzed for intracellular IFN- γ staining after the priming immunizations (wk 60). The assay results provided information on the relative amounts of CD4⁺ and CD8⁺ gag-specific T cells in the peripheral blood (Figure 24). The results indicated that heterologous prime-boost immunization approach was able to elicit in rhesus macaques both HIV-specific CD4⁺ and CD8⁺ T cells.

Ad24 Vaccine Vector as a Heterologous Primer: In a separate study, a cohort of 3 rhesus macaques was immunized initially with 2 doses (wk 0, 4) of 10^{11} vp Ad24 Δ E1 gag Δ Orf6Ad5Orf6 and boosted at wk 24 with 10^7 vp of MRKAd5-gag. The low dose of MRKAd5-gag is selected to mimic the effect of pre-existing neutralizing immunity to the vector in a subject. A separate cohort of naïve animals was given a single dose of 10^7 vp MRKAd5-gag. The results of the IFN- γ ELISPOT analyses of PBMC collected during the course of the studies are shown in Figure 25.

The Ad24-based vaccine was able to prime effectively for HIV-specific T cell responses in macaques. Boosting with a low dose MRKAd5-gag resulted in a significant increase in the levels of gag-specific T cells. The increases in 2 out of 3 animals exceed the levels typically observed after treatment of naïve animals with the same low dose of MRKAd5-gag.

EXAMPLE 18

Construction of pAd34 Δ E1 Δ E4Ad5Orf6

To generate an E1- Ad34 based vector that can propagate in existing group C/Ad5 E1 complementing cell lines (293, PER.C6), Ad5 Orf6 was inserted in place of the native E4 region. Since at the time, the complete sequence of Ad34 (see Figures 28A-1 to 28A-9; subject of copending application serial no. 60/458,825, filed March 28, 2003) was unknown, advantage was taken of the sequence homology between Ad34 and Ad35 in order to construct the Ad34 pre-Adenovirus plasmid. Cotransformation of BJ 5183 bacteria with purified wild-type Ad34 viral DNA and the appropriately constructed Ad35 ITR cassette resulted in the circularization of the viral genome by homologous recombination. The construction of the pre-Ad plasmid based on Ad34, is outlined below:

To construct pAd34 Δ E1 Δ E4Ad5Orf6 (An Ad34 pre-Ad plasmid containing an E1 deletion and an E4 deletion substituted with Ad5 Orf6), we utilized an Ad35 ITR cassette. We anticipated that sequence homology between Ad34 and Ad35 would allow homologous recombination to occur. The Ad35 ITR cassette was constructed containing sequences from the right (bp 31599 to 31913 and bp 34419 to 34793) and left (bp 4 to 456 and bp 3403 to 3886) end of the Ad35 genome (see Figures 2A-1 to 2A-10) separated by plasmid sequences containing a

bacterial origin of replication and an ampicillin resistance gene. The four segments were generated by PCR and cloned sequentially into pNEB193, generating pNEBAd35-4. Next the Ad5 Orf6 open reading frame was generated by PCR and cloned between Ad35 bp 31913 and 34419 generating pNEBAd35-4Ad5Orf6 (the ITR cassette). PNEB193 is a commonly used commercially available cloning plasmid (New England Biolabs cat# N3051S) containing a bacterial origin of replication, ampicillin resistance gene and a multiple cloning site into which the PCR products were introduced. The ITR cassette contains a deletion of E1 sequences from Ad35 bp 457 to 3402 with a unique *Swa* I restriction site located in the deletion and an E4 deletion from Ad35 bp 31914 to 34418 into which Ad5 Orf6 was introduced in an E4 parallel orientation. In this construct Ad5Orf6 expression is driven by the Ad35 E4 promoter. The Ad35 sequences (bp 31599 to 31913 and bp 3403 to 3886) in the ITR cassette provided regions of homology with the purified Ad34 viral DNA in which bacterial recombination could occur following cotransformation into BJ 5183 bacteria (Figure 26). The ITR cassette was also designed to contain unique restriction enzyme sites (*Pme*I) located at the end of the viral ITR's so that digestion would release the recombinant Ad34 genome from the plasmid sequences. Potential clones were screened by restriction analysis and one clone was selected as pAd34ΔE1ΔE4Ad5Orf6.

EXAMPLE 19

Rescue of pAd34ΔE1ΔE4Ad5Orf6 into Virus

In order to determine if pre-adenovirus plasmid pAd34ΔE1ΔE4Ad5Orf6, could be rescued into virus and propagated in a group C E1 complementing cell line, the plasmid was digested with *Pme* I and transfected into T-25 flasks of PER.C6 cells using the calcium phosphate co-precipitation technique (Cell Pfect Transfection Kit, Amersham Pharmacia Biotech Inc). *Pme*I digestion releases the viral genome from plasmid sequences allowing viral replication to occur after cell entry. Viral cytopathic effect (CPE), indicating that virus replication and amplification was occurring was observed following transfection. When CPE was complete, approximately 7-10 days post transfection, the infected cells and media were harvested, freeze/thawed three times and the cell debris pelleted by centrifugation. Approximately 1 ml of the cell lysate was used to infect a T-225 flask of PER.C6 cells at 80-90% confluence. Once CPE was reached, infected cells and media were harvested, freeze/thawed three times and the cell debris pelleted by centrifugation. Clarified cell lysates were then used to infect 2-layer NUNC cell factories of PER.C6 cells. Following complete CPE, the virus was purified by ultracentrifugation on CsCl density gradients. In order to verify the genetic structure of the rescued viruses, viral DNA was extracted using pronase treatment

followed by phenol chloroform extraction and ethanol precipitation. Viral DNA was then digested with *HindIII* and treated with Klenow fragment to end-label the restriction fragments with P33-dATP. The end-labeled restriction fragments were then size-fractionated by gel electrophoresis and visualized by autoradiography. The digestion products were compared with the digestion products of the corresponding pre-Adenovirus plasmid (that had been digested with *PmeI/HindIII* prior to labeling) from which they were derived. The expected sizes were observed, indicating that the viruses had been successfully rescued.

EXAMPLE 20

10 Insertion of an Expression Cassette into pAd34ΔE1ΔE4Ad5Orf6

In order to introduce a gag or SEAP expression cassette (see Figures 6 and 7, respectively) into the E1 region of pAd34ΔE1ΔE4Ad5Orf6, bacterial recombination was again used. A gag expression cassette consisting of the following: 1) the immediate early gene promoter from human cytomegalovirus, 2) the coding sequence of the human immunodeficiency virus type 1 (HIV-1) gag (strain CAM-1; 1526 bp) gene, and 3) the bovine growth hormone polyadenylation signal sequence, was cloned into the E1 deletion in Ad35 shuttle plasmid, pNEBAd35-2 (a precursor to the Ad35 ITR cassettes described above), generating pNEBAd35CMVgagBGHPA. pNEBAd35-2 contains Ad35 sequences from the left end of the genome (bp 4 to 456 and bp 3403 to 3886) with a unique *SwaI* site between bp 456 and 3403 at the position of the deletion. The gag expression cassette was obtained from a previously constructed shuttle plasmid by *EcoRI* digestion. Following the digestion the desired fragment was gel purified, treated with Klenow to obtain blunt ends and cloned into the *SwaI* site in pNEBAd35-2. This cloning step resulted in the gag expression cassette being inserted into the E1 deletion between bp 456 and 3403 in the E1 parallel orientation. The shuttle vector containing the gag transgene was digested to generate a DNA fragment consisting of the gag expression cassette flanked by Ad35 bp 4 to 456 and bp 3403 to 3886 and the fragment was purified after electrophoresis on an agarose gel. Cotransformation of BJ-5183 bacteria with the shuttle vector fragment and pAd34ΔE1ΔE4Ad5Orf6, linearized in the E1 region by digestion with *SwaI*, resulted in the generation of the Ad34 gag-containing pre-Adenovirus plasmid pAd34ΔE1gagΔE4Ad5Orf6 by homologous recombination. Potential clones were screened by restriction analysis.

A similar strategy was used to generate Ad34 pre-Ad plasmids containing a SEAP expression cassette. In this case a SEAP expression cassette consisting of: 1) the immediate early gene promoter from human cytomegalovirus, 2) the coding sequence of the human placental SEAP gene, and 3) the bovine growth hormone polyadenylation signal sequence was

cloned into the E1 deletion in Ad35 shuttle plasmid, pNEBAd35-2, generating pNEBAd35CMVSEAPBGHPA. The SEAP expression cassette was obtained from a previously constructed shuttle plasmid by *EcoRI* digestion. Following the digestion the desired fragment was gel purified, treated with Klenow to obtain blunt ends and cloned into the *SwaI* site in pNEBAd35-2. The transgene was then recombined into the pAd34 Δ E1 Δ E4Ad5Orf6, generating pAd34 Δ E1SEAP Δ E4Ad5Orf6 as described above for the gag transgene.

All pre-Ad plasmids were rescued into virus and expanded to prepare CsCl purified stocks as described above.

10 EXAMPLE 21

Construction of pMRKAd34 Δ E1 Δ E4Ad5Orf6

To construct an Ad34 pre-Ad plasmid that was composed entirely of Ad34 sequences, an Ad34 ITR cassette was generated. The Ad34 ITR cassette was constructed containing sequences from the right (bp 31584 to 31895 and bp 34409 to 34772) and left (bp 4 to 456 and bp 3402 to 3885) end of the Ad34 genome (see Figures 28A-1 to 28A-9) separated by plasmid sequences containing a bacterial origin of replication and an ampicillin resistance gene. These four segments were generated by PCR and cloned sequentially into pNEB193, generating pNEBAd34-4. Next the Ad5 Orf6 open reading frame was generated by PCR and cloned between Ad34 bp 31895 and 34409 generating pNEBAd34-4Ad5Orf6 (the ITR cassette).

20 pNEB193 is a commonly used commercially available cloning plasmid (New England Biolabs cat# N3051S) containing a bacterial origin of replication, ampicillin resistance gene and a multiple cloning site into which the PCR products were introduced. The ITR cassette contains a deletion of E1 sequences from Ad34 bp 457 to 3401 with a unique *Swa I* restriction site located in the deletion and an E4 deletion from Ad34 bp 31896 to 34408 into which Ad5 Orf6 was introduced in an E4 parallel orientation. In this construct Ad5Orf6 expression is driven by the Ad34 E4 promoter. The Ad34 sequences (bp 31584 to 31895 and bp 3402 to 3885) in the ITR cassette provided regions of homology with the purified Ad34 viral DNA in which bacterial recombination could occur following cotransformation into BJ 5183 bacteria (Figure 27). The ITR cassette was also designed to contain unique restriction enzyme sites (*PmeI*) located at the end of the viral ITR's so that digestion would release the recombinant Ad34 genome from the plasmid sequences. Potential clones were screened by restriction analysis and one clone was selected as pMRKAd34 Δ E1 Δ E4Ad5Orf6.

EXAMPLE 22

In Vivo StudiesA. Immunization

5 Cohorts of 3 rhesus macaques were given single intramuscular injections of one of the two vectors: (1) 10^{11} vp MRKAd5-SEAP (in MRKAd vector backbone disclosed in PCT/US01/28861, published March 21, 2002); and (2) 10^{11} vp Ad34 Δ E1SEAP Δ E4Ad5Orf6. Rhesus macaques were between 3-10 kg in weight. In all cases, the total dose of each vaccine was suspended in 1 mL of buffer. The macaques were anesthetized (ketamine/xylazine) and the
10 vaccines were delivered i.m. in 0.5-mL aliquots into both deltoid muscles using tuberculin syringes (Becton-Dickinson, Franklin Lakes, NJ). Peripheral blood mononuclear cells (PBMC) were prepared from blood samples collected at several time points during the immunization regimen. All animal care and treatment were in accordance with standards approved by the Institutional Animal Care and Use Committee according to the principles set forth in the *Guide*
15 *for Care and Use of Laboratory Animals*, Institute of Laboratory Animal Resources, National Research Council.

B. SEAP Assay

Serum samples were analyzed for circulating human secreted alkaline
20 phosphatase (SEAP) levels using TROPIX phospho-light chemiluminescent kit (Applied Biosystems Inc). Duplicate 5 μ L aliquots of each serum were mixed with 45 μ L of kit-supplied dilution buffer in a 96-well white DYNEX plate. Serially diluted solutions of a human placental alkaline phosphatase (Catalog no. M5905, Sigma, St. Louis, MO) in 10% naïve monkey serum served to provide the standard curve. Endogenous SEAP activity in the samples was inactivated
25 by heating the well for 30 minutes at 65 °C. Enzymatic SEAP activities in the samples were determined following the procedures described in the kit. Chemiluminescence readings (in relative light units) were recorded using DYNEX luminometer. RLU readings were converted to ng/mL SEAP using a log-log regression analyses.

C. ELISPOT Assay

The IFN- γ ELISPOT assays for rhesus macaques were conducted following a previously described protocol (Allen *et al.*, 2001 *J. Virol.* 75(2):738-749), with some modifications. For antigen-specific stimulation, a peptide pool was prepared from 20-aa peptides that encompass the entire HIV-1 gag sequence with 10-aa overlaps (Synpep Corp.,
35 Dublin, CA). To each well, 50 μ L of $2-4 \times 10^5$ peripheral blood mononuclear cells (PBMCs) were added; the cells were counted using Beckman Coulter Z2 particle analyzer with a lower

size cut-off set at 80 femtoliters ("fL"). Either 50 μ L of media or the gag peptide pool at 8 μ g/mL concentration per peptide was added to the PBMC. The samples were incubated at 37°C, 5% CO₂ for 20-24 hrs. Spots were developed accordingly and the plates were processed using custom-built imager and automatic counting subroutine based on the ImagePro platform (Silver Spring, MD); the counts were normalized to 10⁶ cell input.

D. Intracellular Cytokine Staining (ICS)

To 1 ml of 2 x 10⁶ PBMC/mL in complete RPMI media (in 17x100mm round bottom polypropylene tubes (Sarstedt, Newton, NC)), anti-hCD28 (clone L293, Becton-Dickinson) and anti-hCD49d (clone L25, Becton-Dickinson) monoclonal antibodies were added to a final concentration of 1 μ g/mL. For gag-specific stimulation, 10 μ L of the peptide pool (at 0.4 mg/mL per peptide) were added. The tubes were incubated at 37 °C for 1 hr., after which 20 μ L of 5 mg/mL of brefeldin A (Sigma) were added. The cells were incubated for 16 hr at 37 °C, 5% CO₂, 90% humidity. 4 mL cold PBS/2%FBS were added to each tube and the cells were pelleted for 10 min at 1200 rpm. The cells were re-suspended in PBS/2%FBS and stained (30 min, 4 °C) for surface markers using several fluorescent-tagged mAbs: 20 μ L per tube anti-hCD3-APC, clone FN-18 (Biosource); 20 μ L anti-hCD8-PerCP, clone SK1 (Becton Dickinson); and 20 μ L anti-hCD4-PE, clone SK3 (Becton Dickinson). Sample handling from this stage was conducted in the dark. The cells were washed and incubated in 750 μ L 1xFACS Perm buffer (Becton Dickinson) for 10 min at room temperature. The cells were pelleted and re-suspended in PBS/2%FBS and 0.1 μ g of FITC-anti-hIFN- γ , clone MD-1 (Biosource) was added. After 30 min incubation, the cells were washed and re-suspended in PBS. Samples were analyzed using all four color channels of the Becton Dickinson FACSCalibur instrument. To analyze the data, the low side- and forward-scatter lymphocyte population was initially gated; a common fluorescence cut-off for cytokine-positive events was used for both CD4⁺ and CD8⁺ populations, and for both mock and gag-peptide reaction tubes of a sample.

E. Results

Expression: Serum samples prior to and after the injection were analyzed for circulating SEAP activities and the results are shown in Figure 29. Results indicate that the peak levels of SEAP protein produced by the alternative adenovirus serotype were lower than but were within 3-fold of that of MRKAd5 at the same high dose level of 10¹¹ vp (Figure 29). The levels of SEAP in the serum dropped dramatically after day 10 and were close to background as early as day 15. These observations strongly indicate that the Ad34-based vector is efficient in expressing a transgene following intramuscular administration in a primate.

Immunogenicity: Vaccine-induced T cell responses against HIV-1 gag were quantified using IFN- γ ELISPOT assay against a pool of 20-aa peptides that encompassed the entire protein sequence. The results are shown in Figure 30; they are expressed as the number of spot-forming cells (SFC) per million peripheral blood mononuclear cells (PBMCs) that responded to the peptide pool or the mock (no peptide) control.

Immunization with gag-expressing Ad34 vector induced detectable levels of circulating gag-specific T cells immediately after a single dose of the vector. The responses improved following a second dose given at wk 4. Overall, the responses to the Ad34-based vector were slightly lower than those induced by the same dose of MRKAd5-gag. The results strongly indicate the Ad34-based vector can prime effectively for HIV-specific T cell responses.

IFN- γ ICS analyses of the PBMC from the Ad34-immunized animals revealed that the vector can induce detectable levels of both CD4⁺ and CD8⁺ HIV-specific T cells (Figure 31).

EXAMPLE 23

Heterologous Immunization

Cohorts of 3 monkeys were immunized (at wks 0, 4) with 10^{11} vp Ad34 Δ E1gag Δ E4Ad5Orf6 followed by a booster at week 24 with 10^{10} vp Ad35 Δ E1gag Δ E4Ad5Orf6. Vaccine-induced T cell responses against HIV-1 gag were quantified using IFN- γ ELISPOT assay against a pool of 20-aa peptides that encompassed the entire protein sequence. The results are shown in Figure 32; they are expressed as the number of spot-forming cells (SFC) per million peripheral blood mononuclear cells (PBMCs) that responded to the peptide pool or the mock (no peptide) control.

Immunization with gag-expressing Ad34 vector induced detectable levels of circulating gag-specific T cells that decreased to between 94-139 SFC/ 10^6 PBMC at the time of the boost. Heterologous immunization with an Ad35-based HIV vector resulted in as much as a 3-fold increase in T cell responses.

IFN- γ ICS analyses of the PBMCs from the Ad34 primed/Ad35 boosted animals at week 28 revealed that the vector can induce detectable levels of both CD4⁺ and CD8⁺ HIV-specific T cells (Figure 33).

WHAT IS CLAIMED IS:

1. A means for propagating replication-defective adenovirus in an adenoviral E1-complementing cell line expressing E1 gene product(s) which are non-native to the adenovirus, which comprises:
 - (a) inserting all or a portion of a heterologous adenoviral E4 region comprising nucleic acid sequence encoding open reading frame 6 (ORF6) into a replication-defective adenovirus; wherein the E4 region or portion thereof inserted into the adenovirus is native to a virus of the same adenovirus serotype as the E1 gene product(s) expressed by the complementing cell line;
 - (b) introducing the replication-defective adenovirus into the adenoviral E1-complementing cell line;
 - (c) allowing the replication-defective adenovirus to propagate in the adenoviral E1-complementing cell line; and
 - (d) rescuing the propagated adenovirus.
2. A means in accordance with claim 1 wherein the heterologous adenoviral E4 region or portion thereof comprises the complete adenoviral E4-encoding region.
3. A means in accordance with claim 2 wherein the heterologous adenoviral E4 region or portion thereof comprises the complete adenoviral E4-encoding region and native E4 promoter.
4. A means in accordance with claim 1 wherein the heterologous adenoviral E4 region or portion thereof is inserted into the replication-defective virus in place of nucleic acid sequence encoding open reading frame 6 (ORF6).

5. A means in accordance with claim 1 wherein the heterologous adenoviral E4 region or portion thereof is inserted into the replication-defective virus in place of nucleic acid sequence encoding the complete adenoviral E4-encoding region.

6. A means in accordance with claim 1 wherein the heterologous adenoviral E4 region or portion thereof is derived from a subgroup C adenovirus.

7. A means in accordance with claim 1 wherein the subgroup C adenovirus is adenovirus of serotype 5.

8. A means in accordance with claim 7 wherein the replication-defective adenovirus is an adenovirus of subgroup B.

9. A means in accordance with claim 7 wherein the replication-defective adenovirus is an adenovirus of serotype 35.

10. A means in accordance with claim 1 wherein the heterologous adenoviral E4 region or portion thereof is operatively linked to a heterologous promoter.

11. A means in accordance with claim 1 wherein the adenoviral E1-complementing cell line is a PER.C6® cell line.

12. A replication-defective adenovirus comprising all or a portion of a heterologous E4 region comprising a heterologous adenoviral open reading frame 6 (ORF6).

13. A replication-defective adenovirus in accordance with claim 12 wherein the adenovirus comprises a heterologous gene of interest.

14. A replication-defective adenovirus in accordance with claim 13 wherein the heterologous gene of interest is a gene encoding an HIV-1 antigen.

15. A replication-defective adenovirus in accordance with claim 14 wherein the HIV-1 antigen is selected from the group consisting of HIV-1 gag, pol, nef and env.

16. A replication-defective adenovirus comprising all or a portion of a heterologous E4 region comprising a heterologous adenoviral open reading frame 6 (ORF6) and a gene encoding HIV-1 gag.

17. A replication-defective adenovirus comprising all or a portion of a heterologous E4 region comprising a heterologous adenoviral open reading frame 6 (ORF6) in place of a native E4 region or portion thereof comprising ORF6.

18. A replication-defective adenovirus comprising all or a portion of a heterologous E4 region comprising a complete heterologous E4 region in place of a complete native E4 region.

19. A replication-defective adenovirus comprising a heterologous E4 region or portion thereof comprising a complete heterologous E4 region including E4 promoter in place of a complete native E4 region.

20. Adenovirus propagated in accordance with the means of claim 1.

21. A means in accordance with claim 1 wherein the replication-defective adenovirus comprises a heterologous gene of interest.

22. A means in accordance with claim 21 wherein the heterologous gene of interest is a gene encoding an HIV-1 antigen.

23. A means in accordance with claim 22 wherein the HIV-1 antigen is selected from the group consisting of: HIV-1 gag, pol, nef and env.

24. A replication-defective adenovirus of serotype 35 comprising all or a portion of an adenovirus serotype 5 E4 region comprising open reading frame 6 (ORF6) and a heterologous gene of interest.

25. A replication-defective adenovirus in accordance with claim 24 wherein the heterologous gene of interest is a gene encoding an HIV-1 antigen.

26. A replication-defective adenovirus in accordance with claim 25 wherein the HIV-1 antigen is selected from the group consisting of: HIV-1 gag, pol, nef and env.

27. A replication-defective adenovirus of serotype 35 comprising all or a portion of an adenovirus serotype 5 E4 region comprising open reading frame 6 (ORF6) and a
5 gene encoding HIV-1 gag.

28. A recombinant adenoviral vector of serotype 24 which comprises an E4 gene or a segment of an E4 gene comprising open reading frame 6 ("ORF6") of an alternative serotype.

29. A population of cells comprising the recombinant adenoviral vector of
10 claim 28.

30. A method for producing recombinant, replication-defective adenovirus particles comprising:

(a) introducing a recombinant adenoviral vector of claim 28 into a population of cells expressing adenovirus E1; and

15 (b) harvesting the resultant recombinant, replication-defective adenovirus.

31. Purified recombinant, replication-defective adenovirus particles harvested in accordance with the method of claim 30.

32. A composition comprising purified recombinant adenovirus particles in accordance with claim 31.

20 33. A composition in accordance with claim 32 which comprises a physiologically acceptable carrier.

34. A recombinant adenoviral vector in accordance with claim 28 which is at least partially deleted in E1 and devoid of E1 activity and comprises a heterologous nucleic acid.

35. A composition comprising purified recombinant adenoviral particles in accordance with claim 31 which are at least partially deleted in E1 and devoid of E1 activity and comprise a heterologous nucleic acid.

36. A method for effecting the delivery and expression of heterologous nucleic acid comprising administering the composition of claim 35 prior or subsequent to administration of the heterologous nucleic acid with the same or different vector.

37. A method in accordance with claim 36 wherein the composition is preceded or followed by administration of heterologous nucleic acid with an adenovirus of a different serotype.

38. A composition in accordance with claim 35 wherein the heterologous nucleic acid encodes an HIV antigen.

39. A method for generating a cellular-mediated immune response against HIV in an individual comprising administering to the individual a composition of claim 38.

40. A composition in accordance with claim 39 wherein the HIV antigen is HIV-1 gag or immunologically relevant modification thereof.

41. A composition in accordance with claim 39 wherein the HIV antigen is HIV-1 nef or immunologically relevant modification thereof.

42. A composition in accordance with claim 39 wherein the HIV antigen is HIV-1 pol or immunologically relevant modification thereof.

43. A recombinant adenoviral vector of serotype 24 which is at least partially deleted in E1 and devoid of E1 activity; wherein said vector comprises an E4 gene or a segment of an E4 gene from adenovirus serotype 5 comprising open reading frame 6 ("ORF6"), and a heterologous nucleic acid.

44. A population of cells comprising the recombinant adenoviral vector of claim 43.

45. A method for producing recombinant, replication-defective adenovirus particles comprising:

5 (a) introducing a recombinant adenoviral vector of claim 43 into a population of cells expressing adenovirus serotype 5 E1; and

(b) harvesting the resultant recombinant, replication-defective adenovirus.

46. Purified recombinant, replication-defective adenovirus particles harvested in accordance with the method of claim 45.

10 47. A composition comprising purified recombinant adenovirus particles in accordance with claim 46.

48. A composition in accordance with claim 47 which comprises a physiologically acceptable carrier.

15 49. A method for effecting the delivery and expression of the heterologous nucleic acid comprising administering the composition of claim 48 prior or subsequent to administration of the heterologous nucleic acid with the same or different vector.

50. A method in accordance with claim 49 above wherein the composition is preceded or followed by administration of the heterologous nucleic acid with an adenovirus of a different serotype.

20 51. A composition in accordance with claim 48 wherein the heterologous nucleic acid encodes an HIV antigen.

52. A method for generating a cellular-mediated immune response against HIV in an individual comprising administering to the individual a composition of claim 51.

53. A composition in accordance with claim 51 wherein the HIV antigen is HIV-1 gag or immunologically relevant modification thereof.

54. A composition in accordance with claim 51 wherein the HIV antigen is HIV-1 nef or immunologically relevant modification thereof.

55. A composition in accordance with claim 51 wherein the HIV antigen is HIV-1 pol or immunologically relevant modification thereof.

56. A recombinant adenoviral vector of serotype 34 which comprises an E4 gene or a segment of an E4 gene comprising open reading frame 6 ("ORF6") of an alternative serotype.

57. A population of cells comprising the recombinant adenoviral vector of claim 56.

58. A method for producing recombinant, replication-defective adenovirus particles comprising:

(a) introducing a recombinant adenoviral vector of claim 56 into a population of cells expressing adenovirus E1; and

(b) harvesting the resultant recombinant, replication-defective adenovirus.

59. Purified recombinant, replication-defective adenovirus particles harvested in accordance with the method of claim 58.

60. A composition comprising purified recombinant adenovirus particles in accordance with claim 59.

61. A composition in accordance with claim 60 which comprises a physiologically acceptable carrier.

62. A recombinant adenoviral vector in accordance with claim 56 which is at least partially deleted in E1 and devoid of E1 activity and comprises a heterologous nucleic acid.

63. A composition comprising purified recombinant adenoviral particles in accordance with claim 59 which are at least partially deleted in E1 and devoid of E1 activity and comprise a heterologous nucleic acid.

5 64. A method for effecting the delivery and expression of heterologous nucleic acid comprising administering the composition of claim 63 prior or subsequent to administration of the heterologous nucleic acid with the same or different vector.

65. A method in accordance with claim 64 wherein the composition is preceded or followed by administration of heterologous nucleic acid with an adenovirus of a different serotype.

10 66. A composition in accordance with claim 63 wherein the heterologous nucleic acid encodes an HIV antigen.

67. A method for generating a cellular-mediated immune response against HIV in an individual comprising administering to the individual a composition of claim 66.

15 68. A composition in accordance with claim 67 wherein the HIV antigen is HIV-1 gag or immunologically relevant modification thereof.

69. A composition in accordance with claim 67 wherein the HIV antigen is HIV-1 nef or immunologically relevant modification thereof.

70. A composition in accordance with claim 67 wherein the HIV antigen is HIV-1 pol or immunologically relevant modification thereof.

20 71. A recombinant adenoviral vector of serotype 34 which is at least partially deleted in E1 and devoid of E1 activity; wherein said vector comprises an E4 gene or a segment of an E4 gene from adenovirus serotype 5 comprising open reading frame 6 ("ORF6"), and a heterologous nucleic acid.

72. A population of cells comprising the recombinant adenoviral vector of claim 71.

73. A method for producing recombinant, replication-defective adenovirus particles comprising:

5 (a) introducing a recombinant adenoviral vector of claim 71 into a population of cells expressing adenovirus serotype 5 E1; and

(b) harvesting the resultant recombinant, replication-defective adenovirus.

74. Purified recombinant, replication-defective adenovirus particles harvested in accordance with the method of claim 73.

10 75. A composition comprising purified recombinant adenovirus particles in accordance with claim 74.

76. A composition in accordance with claim 75 which comprises a physiologically acceptable carrier.

15 77. A method for effecting the delivery and expression of the heterologous nucleic acid comprising administering the composition of claim 76 prior or subsequent to administration of the heterologous nucleic acid with the same or different vector.

78. A method in accordance with claim 77 above wherein the composition is preceded or followed by administration of the heterologous nucleic acid with an adenovirus of a different serotype.

20 79. A composition in accordance with claim 76 wherein the heterologous nucleic acid encodes an HIV antigen.

80. A method for generating a cellular-mediated immune response against HIV in an individual comprising administering to the individual a composition of claim 79.

81. A composition in accordance with claim 79 wherein the HIV antigen is HIV-1 gag or immunologically relevant modification thereof.

82. A composition in accordance with claim 79 wherein the HIV antigen is HIV-1 nef or immunologically relevant modification thereof.

5 83. A composition in accordance with claim 79 wherein the HIV antigen is HIV-1 pol or immunologically relevant modification thereof.

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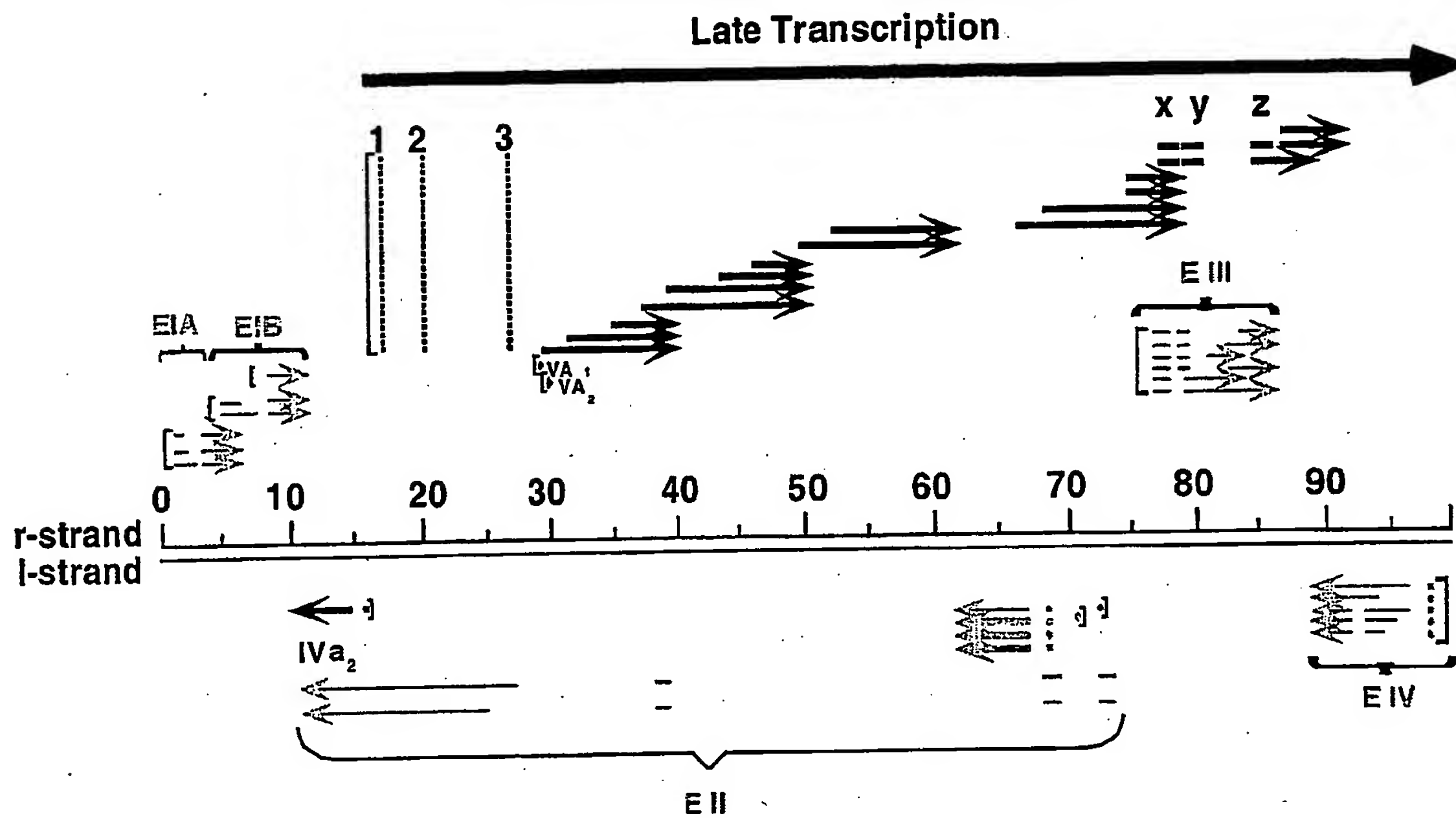


FIG. 1

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FIG. 2A-1

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6421 agttcatcgg gagggctctg atccatggta aagattcccc gaagtaaata cttatcaaaa
6481 tagctgatgg gagtggggct atctaaggcc atttgccatt ctcgagctgc cagtgccgcg
6541 tcatatgggt taaggggact gcccagggc atgggatggg tgagagcaga ggcatacatg
6601 ccacagatgt catagacgta gatgggatcc tcaaagatgc ctatgtaggt catgtgatgg
6661 cgcccccttc tgatacttgc tgcacatag tcatatagtt catgtgatgg cgctagcagc
6721 cccggaccca agttggtgcg attgggtttt tctgttctgt agacgatctg gcgaaagatg
6781 gcgtgagaat tgggaagagat ggtgggtctt tgaaaaatgt tgaaatgggc atgaggtaga
6841 cctacagagt ctctgacaaa gtgggcataa gattcttgaa gcttggttac cagttcgggc
6901 gtgacaagta cgtctagggc gcagtagtca agtgtttctt gaatgatgtc ataacctggt
6961 tgggttttct tttcccacag ttcgcggttg agaaggtatt cttcgcgac cttccagtac
7021 tcttctagcg gaaacccgtc tttgtctgca cggttaagat ctagcatgta gaactgatta
7081 actgccttgt aagggcagca gcccttctct acgggtagag agtatgcttg agcagctttt
7141 cgtagcgaag cgtgagtaag ggcaaagggt tctctgacca tgactttgag aaattggtat
7201 ttgaagtcca tgtcgtcaca ggctccctgt tcccagagtt ggaagtctac ccgtttcttg
7261 taggcggggg tgggcaaagc gaaagtaaca tcattgaaga gaatcttacc ggctctgggc
7321 ataaaattgc gagtgatgcg gaaaggctgt ggtacttccg ctcgattgtt gatcacctgg
7381 gcagctagga cgatttctgc gaaaccgttg atgttgtgtc ctacgatgta taattctatg
7441 aaacgcggcg tgcctctgac gtgaggtagc ttactgagct catcaaaggt taggtctgtg
7501 gggtcagata aggcgtagtg ttcgagagcc cattcgtgca ggtgaggatt tgcattgtagg

FIG. 2A-2

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7561 aatgatgacc aaagatctac cgccagtgtt gtttgttaact ggtcccgata ctgacgaaaa
7621 tgccggccaa ttgccatttt ttctggagtg acacagtaga aggttctggg gtcttgttgc
7681 catcgatccc acttgagttt aatggctaga tctggtggcca tgttgacgag acgtctttct
7741 cctgagagtt tcatgaccag catgaaagga actagtgtgt tgcgaagga tcccatccag
7801 gtgtaagttt ccacatcgta ggtcaggaag agtctttctg tgcgaggatg agagccgac
7861 gggaagaact ggatttcctg ccaccagtgt gaggattggc tgttgatgtg atggaagtag
7921 aagtttctgc ggcgcgccga gcattcgtgt ttgtgcttgt acagacggcc gcagtgtcgt
7981 cagcgttgca cgggttgtat ctcgtgaatg agctgtacct ggcttccctt gacgagaaat
8041 ttcagtggga agccgaggcc tggcgattgt atctcgtgct cttctatatt cgctgtatcg
8101 gcctgttcat cttctgtttc gatgggtgtc atgctgacga gccccgcgg gaggcaagtc
8161 cagacctcgg cgcgggaggg gcggagctga aggacgagag cgcgaggct ggagctgtcc
8221 agagtcctga gacgctgcgg actcaggtta gtaggtaggg acagaagatt aacttgcag
8281 atcttttcca ggcgtgcgg gaggttcaga tggacttga tttccacagg ttcgtttgta
8341 gagacgtcaa tggcttgca ggttccgtgt cctttggcg ccactaccgt acctttgtt
8401 tttcttttga tcggtggtgg ctctcttgct tcttgcatgc tcagaagcgg tgacggggac
8461 gcgcgcggg cggcagcggg tgttccggac ccgggggcat ggctggtagt ggcacgtcgg
8521 cgccgcgcac ggcaggttc tggatttgcg ctctgagaag acttgctgc gccaccacgc
8581 gtcgattgac gtcttgtatc tgacgtctct ggggtgaaag taccggcccc gtgagcttga
8641 acctgaaaga gagttcaaca gaatcaattt cggatcgtt aacggcagct tgtctcagta
8701 tttcttgtac gtcaccagag ttgtcctggg aggcgatctc cgccatgaac tgctcgattt
8761 ctctctcctg aagatctccg cgaccgcctc tttcgacggt ggcgcgagg tcattggaga
8821 tacggcccat gagttgggag aatgcattca tgccgcctc gttccagacg cggctgtaaa
8881 ccacggcccc ctcgaggtct cttgcgcgca tcaccacctg agcgaggtta agctccacgt
8941 gtctggtgaa gaccgcatag ttgcataggc gctgaaaaag gtatgtgagt gtggtggcaa
9001 tgtgttcggc gacgaagaaa tacatgatcc atcgtctcag cggcatttcg ctaacatcgc
9061 ccagagcttc caagcgctcc atggcctcgt agaagtccac ggcaaaatta aaaaactggg
9121 agtttcgcgc ggacacggtc aattcctcct cgagaagacg gatgagttcg gctatggtgg
9181 cccgtacttc gcgttcgaag gctcccggga tctcttcttc ctcttctatc tcttcttcca
9241 ctaacatctc ttcttcgtct tcaggcgggg gcggaggggg cagcggcgca cgtcgacggc
9301 gcacgggcaa acggtcgatg aatcgttcaa tgacctctcc gcggcggcgg cgcattgttt
9361 cagtgcggc gcggccgttc tcgcgcggtc gcagagtaaa aacaccgccg cgcattctct
9421 taaagtgggt actgggaggt tctccgtttg ggagggagag ggcgtgatt atacatttta
9481 ttaattggcc cgtagggact gcgcgcagag atctgatcgt gtcaagatcc acgggatctg
9541 aaaacctttc gacgaaagcg tctaaccagt cacagtcaca aggtaggctg agtacggctt
9601 cttgtggggc ggggtggtta tgtgttcggg ctgggtcttc tgtttcttct tcattctcggg
9661 aaggtgagac gatgctgctg gtgatgaaat taaagtaggc agttctaaga cggcggtagg
9721 tggcgaggag caccaggtct ttgggtccgg cttgctggat acgcaggcga ttggccattc
9781 cccaagcatt atcctgacat ctagcaagat ctttgtagta gtcttgcatg agccgttcta
9841 cgggcacttc ttcctcacc cgttctgcat gcatacgtgt gatgctcaat ccgcgcattg
9901 gttgtaccag tgccaagtca gctacgactc tttcggcgag gatggcttgc tgtacttggg
9961 taagggtggc ttgaaagtca tcaaaatcca caaagcgggt gtaagcccc gtattaatgg
10021 tgtaagcaca gttggccatg actgaccagt taactgtctg gtgaccaggg cgcacgagct
10081 cgggtgtattt aaggcgcgaa taggcgcggg tgtcaaatag gtaatcgttg caggtgcgca
10141 ccagatactg gtaccctata agaaaatgcg gcggtggttg gcggtagaga ggccatcgtt
10201 ctgtagctgg agcgcaggg gcgaggtctt ccaacataag gcggtgatag ccgtagatgt
10261 acctggacat ccaggtgatt cctgcggcgg tagtagaagc ccgaggaaac tgcgtacgc
10321 ggttccaaat gttgcgtagc ggcattgaag agttcattgt aggcacgggt tgaccagtga
10381 ggcgcgcgca gtcattgatg ctctatagac acggagaaaa tgaagcgtt cagcgactcg
10441 actccgtagc ctggaggaac gtgaacgggt tgggtcgcgg tgtaccccg ttcgagactt
10501 gtactcgagc cggccggagc cgcggttaac gtggtattgg cactcccgtc tcgaccacgc
10561 ctacaaaaat ccaggatacg gaatcgagtc gttttgctgg tttccgaatg gcagggaggt
10621 gagtcctatt tttttttttt ttttgccgct cagatgcac ccgtgctgcg acagatgcgc
10681 cccaacaac agccccctc gcagcagcag cagcagcagc aaccacaaaa ggctgtccct
10741 gcaactactg caactgccgc cgtgagcggg gcgggacagc ccgcctatga tctggacttg
10801 gaagagggcg aaggactggc acgtctaggt gcgccttcgc ccgagcggca tccgcgagtt
10861 caactgaaaa aagattctcg cgaggcgtat gtgccccaac agaacctatt tagagacaga
10921 agcggcgagg agccggagga gatgcgagct tcccgttta acgcgggtcg tgagctgcgt
10981 cacggttttg accgaagac agtgttgcca gacgaggatt tcgaagttga tgaagtgaca
11041 gggatcagtc ctgccagggc acacgtggct gcagccaacc ttgtatcggc ttacgagcag
11101 acagtaaagg aagagcgtaa cttccaaaag tcttttaata atcatgtgcg aaccctgatt
11161 gcccgcgaag aagttacctt tggtttgatg catttggtgg atttgatgga agctatcatt
11221 cagaacccta ctagcaaac tctgaccgcc cagctgttcc tgggtggtgca acacagcaga
11281 gacaatgagg ctttcagaga ggcgtgctg aacatcaccg aacccgaggg gagatggttg

FIG. 2A-3

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11341	tatgatctta	tcaacattct	acagagtatc	atagtgcagg	agcggagcct	gggcctggcc
11401	gagaaggtag	ctgccatcaa	ttactcgggt	ttgagcttgg	gaaaatatta	cgctcgcaaa
11461	atctacaaga	ctccatacgt	tcccatagac	aaggaggtga	agatagatgg	gttctacatg
11521	cgcatgacgc	tcaaggtctt	gaccctgagc	gatgatcttg	gggtgtatcg	caatgacaga
11581	atgcatcgcg	cggtttagcgc	cagcaggagg	cgcgagttaa	gcgacaggga	actgatgcac
11641	agtttgcaaa	gagctctgac	tggagctgga	accgaggggtg	agaattactt	cgacatggga
11701	gctgacttgc	agtggcagcc	tagtcgcagg	gctctgagcg	ccgcgacggc	aggatgtgag
11761	cttccttaca	tagaagaggg	ggatgaaggg	gaggaggaag	agggcgagta	cttgggaagac
11821	tgatggcaca	accctgtgtt	tttgctagat	ggaacagcaa	gcaccggatc	ccgcaatgcg
11881	ggcggcgctg	cagagccagc	cgtccggcat	taactcctcg	gacgattgga	cccaggccat
11941	gcaacgtatc	atggcggtga	cgactcgcaa	ccccgaagcc	tttagacagc	aacccaggcc
12001	caaccgtcta	tccggccatca	tgggaagctgt	agtgccttcc	cgatctaatc	ccactcatga
12061	gaaggctcctg	gccatcgtga	acgcgttgggt	ggagaacaaa	gctattcgtc	cagatgaggg
12121	cggactggta	tacaacgctc	tcttagaacg	cgtggctcgc	tacaacagta	gcaatgtgca
12181	aaccaatttg	gaccgtatga	taacagatgt	acgcgaagcc	gtgtctcagc	gcgaaagggt
12241	ccagcgtgat	gccaaacctgg	gttcgctgggt	ggcggttaaat	gctttcttga	gtactcagcc
12301	tgctaattgtg	ccgcgtgggtc	aacaggatta	tactaacttt	ttaagtgtct	tgagactgat
12361	ggtatcagaa	gtacctcaga	gcgaagtgtg	tcagtccgggt	cctgattact	tctttcagac
12421	tagcagacag	ggcttgcaga	cggtaaatct	gagccaagct	tttaaaaacc	ttaaagggtt
12481	gtggggagtg	catgccccgg	taggagaaag	agcaaccgtg	tctagcttgt	taactccgaa
12541	ctcccgctg	ttattactgt	tggtagctcc	tttcaccgac	agcggtagca	tcgaccgtaa
12601	ttcctatttg	ggttacctac	taaacctgta	tcgcgaagcc	atagggcaaa	gtcagggtgga
12661	cgagcagacc	tatcaagaaa	ttacccaagt	cagtcgcgct	ttgggacagg	aagacactgg
12721	cagtttgga	gccactctga	acttcttgct	taccaatcgg	tctcaaaaaga	tccctcctca
12781	atatgctctt	actgcggagg	aggagaggat	ccttagatat	gtgcagcaga	gcgtgggatt
12841	gtttctgatg	caagaggggg	caactccgac	tgcagcactg	gacatgacag	cgcgaaatat
12901	ggagcccagc	atgtatgcca	gtaaccgacc	tttcattaac	aaactgctgg	actacttgca
12961	cagagctgcc	gctatgaact	ctgattatgt	caccaatgcc	atcttaaacc	cgactgggt
13021	gccccacct	ggtttctaca	cgggcgaata	tgacatgccc	gaccctaattg	acggatttct
13081	gtgggacgac	gtggacagcg	atgttttttc	acctctttct	gatcatcgca	cgtggaaaaa
13141	ggaaggcgggt	gatagaatgc	attcttctgc	atcgctgtcc	ggggtcatgg	gtgctaccgc
13201	ggctgagccc	gagtcctgcaa	gtccttttcc	tagtctaccc	ttttctctac	acagtgtacg
13261	tagcagcgaa	gtgggtagaa	taagtcgccc	gagtttaattg	ggcgaagagg	agtacctaaa
13321	cgattccttg	ctcagaccgg	caagagaaaa	aaatttccca	aacaatggaa	tagaaagttt
13381	ggtggataaa	atgagtagat	ggaagactta	tgctcaggat	cacagagacg	agcctgggat
13441	catggggact	acaagtagag	cgagccgtag	acgccagcgc	catgacagac	agaggggtct
13501	tgtgtgggac	gatgaggatt	cggccgatga	tagcagcgtg	ttggacttgg	gtgggagagg
13561	aaggggcaac	ccgtttgctc	atgtgcgccc	tcgcttgggt	ggtatgttgt	gaaaaaaaat
13621	aaaaaagaaa	aactcaccaa	ggccatggcg	acgagcgtac	gttcgttctt	ctttattatc
13681	tgtgtctagt	ataatgaggg	gagtcgtgct	aggcggagcg	gtgggtgtatc	cggaggggtcc
13741	tcctccttcg	tacgagagcg	tgatgcagca	gcagcagggc	acggcgggtga	tgcaatcccc
13801	actggaggct	ccctttgtgc	ctccgcgata	cctggcacct	acggagggca	gaaacagcat
13861	tcgttactcg	gaactggcac	ctcagtacga	taccaccagg	ttgtatctgg	tggacaacaa
13921	gtcggcgggac	attgcttctc	tgaactatca	gaatgaccac	agcaacttct	tgaccacgggt
13981	ggtgcagaac	aatgacttta	cccctacgga	agccagcacc	cagaccatta	actttgatga
14041	acgatcgcg	tggggcggtc	agctaaagac	catcatgcat	actaacatgc	caaacgtgaa
14101	cgagtatatg	tttagtaaca	agttcaaagc	gcgtgtgatg	gtgtccagaa	aacctcccga
14161	cgggtgctgca	gttggggata	cttatgatca	caagcaggat	attttggaat	atgagtgggt
14221	cgagtttact	ttgccagaag	gcaacttttc	agttactatg	actattgatt	tgatgaacaa
14281	tgccatcata	gataattact	tgaaagtggg	tagacagaat	ggagtgcctg	aaagtgcacat
14341	tggtgttaag	ttcgacacca	ggaacttcaa	gctgggatgg	gatcccgaag	ccaagtgtgat
14401	catgcctgga	gtgtatacgt	atgaagcctt	ccatcctgac	attgtcttac	tgccctggctg
14461	cggagtggat	tttaccgaga	gtcgtttgag	caaccttctt	ggtatcagaa	aaaaacagcc
14521	atttcaagag	ggttttaaga	ttttgtatga	agatttagaa	ggtggtaata	ttccggccct
14581	cttggtatga	gatgcctatg	agaacagtaa	gaaagaacaa	aaagccaaaa	tagaagctgc
14641	tacagctgct	gcagaagcta	aggcaaacat	agttgccagc	gactctacaa	gggttgctaa
14701	cgctggagag	gtcagaggag	acaattttgc	gccaacacct	gttccgactg	cagaatcatt
14761	attggccgat	gtgtctgatg	gaacggacgt	gaaactcact	attcaacctg	tagaaaaaga
14821	tagtaagaat	agaagctata	atgtgttggg	agacaaaatc	aacacagcct	atcgagcttg
14881	gtatctttcg	tacaattatg	gcgatcccga	aaaaggagtg	cgttcctgga	cattgctcac
14941	cacctcagat	gtcacctgcg	gagcagagca	ggtttactgg	tcgcttccag	acatgatgaa
15001	ggatcctgtc	actttccgct	ccactagaca	agtcagtaac	taccctgtgg	tgggtgcaga
15061	gcttatgccc	gtctttctcaa	agagcttcta	caacgaacaa	gctgtgtaet	cccagcagct

FIG. 2A-4

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15121 ccgccagttcc acctcgctta cgcacgtctt caaccgcttt cctgagaacc agattttaat
15181 ccgtccgccg gcgccacca ttaccaccgt cagtgaanaac gttcctgctc tcacagatca
15241 cgggaccctg ccgttgccga gcagtatccg gggagtccaa cgtgtgaccg ttactgacgc
15301 cagacgccgc acctgtccct acgtgtacaa ggcactgggc atagtcgcac cgcgcgtcct
15361 ttcaagccgc actttctaaa aaaaaaatgt ccattcttat ctgccccagt aataacaccg
15421 gttgggggtct gcgcgtccca agcaagatgt acggaggcgc acgcaaacgt tctaccaaac
15481 atcccgtgcg tggttcgcgga cattttcgcg ctccatgggg tgccctcaag ggccgcactc
15541 gcgttcgaac caccgtcgat gatgtaatcg atcagggtgt tgccgacgcc cgtaattata
15601 ctccactatgc gcctacatct actgtggatg cagttattga cagtgtagt gctgacgctc
15661 gcaactatgc tcgacgtaag agccggcgaa ggcgcattgc cagacgccac cgagctacca
15721 ctgccatgcy agccgcaaga gctctgctac gaagagctag acgcgtgggg cgaagagcca
15781 tgcttagggc ggccagacgt gcagcttcg ggcgcagcgc cggcagggtc cgcaggcaag
15841 cagccgctgt cgcagcggcg actattgccg acatggccca atcgcgaaga ggcaatgtat
15901 actgggtgcy tgacgctgcc accggtcaac gtgtaccctg gcgcaccctg cccctcgcga
15961 cttagaagat actgagcagt ctccgatgt gtgtcccagc ggcgaggatg tccaagcgca
16021 aatacaagga agaaatgctg caggttatcg cacctgaagt ctacggccaa ccgttgaagg
16081 atgaaaaaaa accccgcaaa atcaagcggg ttaaaaagga caaaaaagaa gaggaagatg
16141 gcgatgatgg gctggcggag tttgtgcgcy agtttgcccc acggcgacgc gtgcaatggc
16201 gtgggcycaa agttcgacat gtgttgagac ctggaacttc ggtggtcttt acaccggcg
16261 agcgttcaag cgtactttt aagcgttct atgatgaggt gtacggggat gatgatattc
16321 ttgagcaggc ggctgaccga ttaggcgagt ttgcttatgg caagcgtagt agaataactt
16381 ccaaggatga gacagtgtca atacccttg atcatggaaa tcccaccctt agtcttaaac
16441 cggtcacttt gcagcaagtg ttaccctgaa ctccgcgaac aggtgttaaa cgcgaagggtg
16501 aagatttgta tcccactatg caactgatgg tacccaaacg ccagaagtgt gaggacgttt
16561 tggagaaagt aaaagtggat ccagatattc aacctgaggt taaagtgaga cccattaagc
16621 aggtagcgc ccgtctgggg gtacaaactg tagacattaa gattccact gaaagtatgg
16681 aagtgcacac tgaaccgcga aagcctactg ccacctccac tgaagtgcac acggatccat
16741 ggatgccccat gcctattaca actgacgcgc ccggtccac tcgaagatcc cgacgaaagt
16801 acggtccagc aagtctgttg atgcccatt atgttgatca cccatctatt attcctactc
16861 ctggttaccg aggcactcgc tactatcgca gccgaacag tacttccgc cgtcgcgcga
16921 agacacctgc aaatcgagc cgtcgcgcga gacgcacaag caaacgcact cccggcgccc
16981 tgggtgcggca agtgtaccgc aatggtagtg cggaaacctt gacactgccg cgtgcgcgtt
17041 accatccgag tatcatcact taatcaatgt tgccgctgcy tcttgaga tatggccctc
17101 acttgctgc ccgtcggttcc catcactggg taccgaggaa gaaactcgc ccgtagaaga
17161 gggatgtttg gacgcggaat gcgacgctac aggcgacggc gtgctatccg caagcaattg
17221 cggggtggtt ttttaccagc cttaattcca attatcgct ctgcaattgg cgcgatacca
17281 ggcatagctt ccgtggcggt tcaggcctcg caacgacatt gacattggaa aaaaaacgta
17341 taaataaaaa aaaatacaat ggactctgac actcctgggt ctgtgactat gttttcttag
17401 agatggaaga catcaatttt tcatecttgg ctccgcgaca cggcacgaag ccgtacatgg
17461 gcacctggag cgacatcggc acgagccaac tgaacggggg cgccttcaat tggagcagta
17521 tctggagcgg gcttaaaaaat tttggctcaa ccataaaaa acacgacttc caacaaaaag
17581 acagcagtac aggcagggcg cttagaaata aacttaaaga ccagaacttc caggctgtgc
17641 tagtcgatgg gatagcttcc ggcataatg gagtggtaga tttggctaac atgcaagtgg
17701 agaaaaagat aaacagtcgt ttggaccgc cgcagcaac cccagggtga gatttggaag
17761 aggaagaaat tcctccgcca gaaaaacgag ggcacaagcg tccgcgtccc aagcttgga
17821 agacgctggg gacgcgcgta gatgaaccgc cttcttatga ggaagcaacg aagcttgga
17881 tgcccaccac tagaccgata gcccacatgg ccaccgggt gatgaaacct tctcagttgc
17941 atcgaccctg caccttggat ttgccccct cccctgctgc tactgctgta cccgcttcta
18001 agcctgtcgc tgccccgaaa ccagtcgccc tagccagggt acgtcccggg ggcgctcctc
18061 gtccaaatgc gactggcaa aatactctga acagcatcgt ggggtctagg gtgcaaagtg
18121 taaaacgccc tcgctgcttt taattaaata tggagttagc cttaacttgc ctatctgtgt
18181 atatgtgtca ttacacgcgc tcacagcagc agaggaaaaa aggaagaggt cgtgcgtcga
18241 cgctgagtta ctttcaagat ggccacccca tcgatgctgc cccaatgggc ataatgacac
18301 atcgccggac aggatgtctt ggagtagctg agtccgggtc tgggtgagtt cgcgcgcgc
18361 acagacacct acttcaatct gggaaataag tttagaaatc ccaccgtagc gccgaccac
18421 gatgtgacca ccgaccgtag ccagcggctc atgttgctgt tegtgcctgt tgaccgggag
18481 gacaatacat actcttaca agtgcggtac accctggccg tgggcgacaa cagagtgtgt
18541 gatattggcca gcacgttctt tgacattagg ggcgtgttgg acagaggtcc cagtttcaaa
18601 ccctattctg gtacggctta caactctctg gctcctaaag gcgctccaaa tgcattctca
18661 tggattgcaa aaggcgtacc aactgcagca gccgcaggca atggtgaaga agaactgaa
18721 acagaggaga aaactgttac ttacactttt gccaatgtc ctgtaaaagc cgaggctcaa
18781 attacaaaag agggcttacc aataggtttg gagatttcag ctgaaaacga atctaaaccc
18841 atctatgcag ataaacttta tcagccagaa cctcaagtgg gagatgaaac ttggactgac

FIG. 2A-5

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18901	ctagacggaa	aaaccgaaga	gtatggaggc	agggctctaa	agcctactac	taacatgaaa
18961	ccctgttacg	ggtcctatgc	gaagcctact	aatttaaaag	gtggtcaggc	aaaaccgaaa
19021	aactcggaac	cgtcgagtg	aaaaattgaa	tatgatattg	acatggaatt	ttttgataac
19081	tcatcgcaaa	gaacaaactt	cagtcctaaa	attgtcatgt	atgcagaaaa	tgtaggtttg
19141	gaaacgccag	acactcatgt	agtgtacaaa	cctggaacag	aagacacaag	ttccgaagct
19201	aatttgggac	aacagtctat	gcccacacag	cccaactaca	ttggcttcag	agataacttt
19261	attggactca	tgtactataa	cagtactggt	aacatggggg	tgctggctgg	tcaagcgtct
19321	cagttaaagt	cagtgggtga	cttgcaggac	agaaacacag	aactttctta	ccaactcttg
19381	cttgactctc	tgggagacag	aaccagatac	tttagcatgt	ggaatcaggc	tgtggacagt
19441	tatgatcctg	atgtacgtgt	tattgaaaat	catgggtgtg	aagatgaact	tcccaactat
19501	tgttttccac	tggacggcat	aggtgttcca	acaaccagtt	acaaatcaat	agttccaaat
19561	ggagaagata	ataataattg	gaaagaacct	gaagtaaagt	gaacaagtga	gatcggacag
19621	ggtaatttgt	ttgccatgga	aattaacctt	caagccaatc	tatggcgaag	tttcctttat
19681	tccaatgtgg	ctctgtatct	cccagactcg	tacaaataca	ccccgtccaa	tgtcactctt
19741	ccagaaaaca	aaaacaccta	cgactacatg	aacgggaggc	tggtgcccgc	atctctagta
19801	gacacctatg	tgaacattgg	tgccagggtg	tctctggatg	ccatggacaa	tgtcaaccca
19861	ttcaaccacc	accgtaacgc	tggtctgcgt	taccgatcta	tgcttctggg	taacggacgt
19921	tatgtgcctt	tccacataca	agtgcctcaa	aaattcttcg	ctgttaaaaa	cctgctgctt
19981	ctcccaggct	cctacactta	tgagtggaa	tttaggaagg	atgtgaacat	ggttctacag
20041	agttccctcg	gtaacgacct	gcgggtagat	ggcgccagca	tcagtttcac	gagcatcaac
20101	ctctatgcta	cttttttccc	catggctcac	aacaccgctt	ccacccttga	agccatgctg
20161	cggaatgaca	ccaatgatca	gtcattcaac	gactacctat	ctgcagctaa	catgctctac
20221	cccattcctg	ccaatgcaac	caatattccc	atttccattc	cttctcgcaa	ctgggaggct
20281	ttcagaggct	ggtcatttac	cagactgaaa	accaaagaaa	ctccctcttt	gggtcttgga
20341	tttgacccct	actttgtcta	ttctggttct	attccctacc	tggtgggtac	cttctacctg
20401	aaccacactt	ttaagaaggt	ttccatcatg	tttgactctt	cagtgaagct	gcctggaaat
20461	gacagggttac	tatctcctaa	cgaatttgaa	ataaagcgca	ctgtggatgg	cgaaggctac
20521	aacgtagccc	aatgcaacat	gaccaaagac	tggttcttgg	tacagatgct	cgccaactac
20581	aacatcggct	atcagggtct	ctacattcca	gaaggatata	aagatcgcat	gtattcattt
20641	ttcagaaact	tccagcccat	gagcaggcag	gtgggttgatg	aggtcaatta	caaagacttc
20701	aaggccgctg	ccatacccta	ccaacacaa	aactctggct	ttgtgggtta	catggctccg
20761	accatgcgcc	aagggtcaacc	ctatcccgtc	aactatccct	atccactcat	tggaacaact
20821	gccgtaaata	gtgttacgca	gaaaaagtct	ttgtgtgaca	gaaccatgtg	gcgcataacc
20881	ttctcgagca	acttcatgtc	tatggggggc	cttacagact	tgggacagaa	tatgctctat
20941	gccaaactcag	ctcatgctct	ggacatgacc	tttgagggtg	atcccatgga	tgagcccacc
21001	ctgctttatc	ttctcttcga	agttttcgac	gtggtcagag	tgcatcagcc	acaccgcggc
21061	atcatcgagg	cagtctacct	gcgtacaccg	ttctcgcccg	gtaacgctac	cacgtaagaa
21121	gcttcttgct	tcttgcaaat	agcagctgca	accatggcct	gcggatccca	aaacggctcc
21181	agcgagcaag	agctcagagc	cattgtccaa	gacctgggtt	gcggacccta	ttttttggga
21241	acctacgata	agcgcttccc	gggtttcatg	gcccccgata	agctcgccctg	tgccattgta
21301	aatacggccg	gacgtgagac	gggggggagag	cactgggttg	ctttcggttg	gaaccacagt
21361	tctaacacct	gctacctttt	tgatcctttt	ggattctcgg	atgatcgtct	caaacagatt
21421	taccagtttg	aatatgaggg	tctcctgcgc	cgcagcgtct	ttgctaccaa	ggaccgctgt
21481	attacgctgg	aaaaatctac	ccagaccgtg	cagggccccc	gttctgcccgc	ctgcggactt
21541	ttctgctgca	tgttccttca	cgcctttgtg	cactggcctg	accgtcccat	ggacggaaac
21601	cccaccatga	aattgctaac	tggagtggca	aacaacatgc	ttcattctcc	taaagtccag
21661	cccaccctgt	gtgacaatca	aaaagcactc	taccattttc	ttaataccca	ttcgccctat
21721	tttcgctctc	atcgtaacaa	catcgaaagg	gccactgcgt	tcgaccgtat	ggatgttcaa
21781	taatgactca	tgtaaacaac	gtgttcaata	aacatcactt	tattttttta	catgtatcaa
21841	ggctctggat	tacttattta	tttacaagtc	gaatgggttc	tgacgagaat	cagaatgacc
21901	cgcaggcagt	gatacgttgc	ggaactgata	cttgggttgc	cacttgaatt	cggaatcac
21961	caacttggga	accggtatat	cgggcaggat	gtcactccac	agctttcttg	tcagctgcaa
22021	agctccaagc	aggtcaggag	ccgaaatctt	gaaatcacaa	ttaggaccag	tgctctgagc
22081	gcgagagtgg	cggtacaccg	gattgcagca	ctgaaacacc	atcagcgacg	gatgtctcac
22141	gcttgccagc	acggtgggat	ctgcaatcat	gcccacatcc	agatcttcag	cattggcaat
22201	gctgaacggg	gtcatcttgc	aggtctgcct	acccatggcg	ggcaccatcc	taggcttggg
22261	gttgcaatcg	cagtgcaggg	ggatcagtat	catcttggcc	tgatcctgtc	tgattcctgg
22321	atacacggct	ctcatgaaag	catcatattg	cttgaaagcc	tgctgggctt	tactaccctc
22381	ggtataaaac	atcccgcagg	acctgctcga	aaactgggta	gctgcacagc	cgccatcatt
22441	cacacagcag	cgggagtcac	tggtggctat	ttgcaccaca	cttctgcccc	agcggttttg
22501	ggtgattttg	gttcgctcgg	gattctcctt	taaggctcgt	tgccgcttct	cgctggccac
22561	atccatctcg	ataatctgct	ccttctgaat	cataatattg	ccatgcaggc	acttcagctt
22621	gccctcataa	tcattgcagc	catgaggcca	caacgcacag	cctgtacatt	cccaattatg

FIG. 2A-6

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22681 gtgggcgatc tgagaaaaag aatgtatcat tccctgcaga aatcttccca tcatcgtgct
22741 cagtgtcttg tgactagtga aagttaactg gatgcctcgg tgctcttcgt ttacgtactg
22801 gtgacagatg cgcttggtatt gttcgtgttg ctcaggcatt agtttaaaac aggttctaag
22861 ttcgttatcc agcctgtact tctccatcag cagacacatc acttccatgc ctttctccca
22921 agcagacacc aggggcaagc taatcggatt cttaacagtg caggcagcag ctcctttagc
22981 cagaggggtca tcttttagcga tcttctcaat gcttcttttg ccatccttct caacgatgcg
23041 cacgggcggg tagctgaaac ccactgctac aagttgcgcc tcttctcttt cttcttcgct
23101 gtcttgactg atgtcttgca tggggatatg tttggtcttc cttgggttct ttttgggggg
23161 tatcggagga ggaggactgt cgctccgttc cggagacagg gaggattgtg acgtttcgct
23221 caccattacc aactgactgt cggtagaaga acctgacccc acacggcgac aggtgttttt
23281 cttcgggggc agaggtggag gcgattgcga agggctgcgg tccgacctgg aaggcggatg
23341 actggcagaa ccccttccgc gttcgggggt gtgctccctg tggcggtcgc ttaactgatt
23401 tccttcgogg ctggccattg tgttctccta ggcagagaaa caacagacat ggaaactcag
23461 ccattgctgt caacatcgcc acgagtgcga tcacatctcg tctcagcga cgaggaaaag
23521 gagcagagct taagcattcc accgcccagt cctgccacca cctctaccct agaagataag
23581 gaggtcgcag catctcatga catgcagaat aaaaaagcga aagagtctga gacagacatc
23641 gagcaagacc cgggctatgt gacaccggtg gaacacgagg aagagtgaac acgctttcta
23701 gagagagagg atgaaaactg cccaaaacag cgagcagata actatcacca agatgctgga
23761 aatagggatc agaacaccga ctacctcata gggcttgacg ggggaagacgc gctccttaa
23821 catctagcaa gacagtgcgt catagtcaag gatgcattat tggacagaaac tgaagtgcgc
23881 atcagtgtgg aagagctcag ctgcgcctac gagcttaacc ttttttcacc tcgtactccc
23941 cccaaacgtc agccaaacgg cacctgcgag ccaaatcctc gcttaaactt ttatccagct
24001 tttgctgtgc cagaagtact ggctacctat cacatctttt ttaaaaatca aaaaattcca
24061 gtctcctgcc gcgctaactg caccgcgcgc gatgccttac tcaatctggg acctgggtca
24121 cgcttacctg atatagcttc cttggaagag gttccaaaga tcttcgaggg tctgggcaat
24181 aatgagactc gggccgcaaa tgctctgcaa aaggagaaaa atggcatgga tgagcatcac
24241 agcgttcttg tgggaattgga aggcgataat gccagactcg cagtactcaa gcgaagcgtc
24301 gaggtcacac acttcgcata tcccgctgtc aacctgcccc cttaaagtcac gacggcggtc
24361 atggaccagt tactcattaa gcgcgcaagt cccctttcag aagacatgca tgaccagat
24421 gcctgtgatg agggtaaacc agtggtcagt gatgagcagc taaccgatg gctgggcacc
24481 gactctcccc gggatttggga agagcgtcgc aagcttatga tggccgtggt gctggttacc
24541 gtagaactag agtgtctccg acgtttcttt accgattcag aaaccttgcg caaactcgaa
24601 gagaatctgc actacacttt tagacacggc tttgtgcggc aggcattgcaa gatattctaac
24661 gtggaactca ccaacctggt ttcctacatg ggtattctgc atgagaatcg cctaggacaa
24721 agcgtgctgc acagcaccct taagggggaa gcccgccgtg attacatccg cgatttgttc
24781 tatctctacc tgtgccacac gtggcaaac ggcatgggtg tatggcagca atgtttagaa
24841 gaacagaact tgaaagagct tgacaagctc ttacagaaat ctcttaaggt tctgtggaca
24901 ggggttcgacg agcgcaccgt cgcttccgac ctggcagacc tcatcttccc agagcgtctc
24961 agggttactt tgcgaaacgg attgcctgac tttatgagcc agagcatgct taacaatttt
25021 cgctctttca tcctggaacg ctccggtatc ctgcccgcga cctgctgcgc actgccctcc
25081 gactttgtgc ctctcaccta ccgcgagtgc ccccgccgc tatggagtca ctgctacctg
25141 ttccgtcttg ccaactatct ctctaccac tcggatgtga tccgaggatg gagcggagac
25201 ggcttgctgg agtgccactg ccgctgcaat ctgtgcacgc cccaccggtc cctagcttgc
25261 aacccccagt tgatgagcga aaccagata ataggcacct ttgaattgca agggcccagc
25321 agccaaggcg atgggtcttc tcctgggcaa agtttaaaac tgacccccgg actgtggacc
25381 tccgcctact tgcgcaagtt tgctccggaa gattaccacc cctatgaaat caagttctat
25441 gaggaccaat cacagcctcc aaaggccgaa ctttcggctt gcgtcatcac ccagggggca
25501 attctggccc aattgcaagc catccaaaaa tcccgccaag aatttctact gaaaaaggtt
25561 aagggggtct accttgacc ccagaccggc gaggaactca acacaaggtt cctcaggat
25621 gtcccaacga cgagaaaaca agaagtgaac ggtgcagccg ccgccccag aagatatgga
25681 ggaagattgg gacagtcagg cagaggaggc ggaggaggac agtctggagg acagtctgga
25741 ggaagacagt ttggaggagg aaaacgagga ggcagaggag gtggaagaag taaccgccga
25801 caaacagtta tcctcggctg cggagacaag caacagcgtc accatctccg ctccgagtgc
25861 aggaacccgg cggcgtccca gcagtagatg ggacgagacc ggacgcttcc cgaaccaaac
25921 cagcgttccc aagaccggtg agaaggatcg gcagggatac aagtcctggc gggggcataa
25981 gaatgccatc atctcctgct tgcattgagt cgggggcaac atatccttca cgcggcgcta
26041 cttgctattc caccatgggg tgaactttcc gcgcaatgtt ttgcattact accgtcacct
26101 ccacagcccc tactatagcc agcaaattccc gacagtctcg acagataaag acagcggcgg
26161 cgacctccaa cagaaaacca gcagcggcag ttagaaaaata cacaacaagt gcagcaacag
26221 gaggattaaa gattacagcc aacgagccag cgcaaaccgg agagttaaga aatcggatct
26281 ttccaaccct gtatgccatc ttccagcaga gtcgggggtc agagcaggaa ctgaaaataa
26341 aaaaccgatc tctgcgttcg ctcaccagaa gttgtttgta tcacaagagc gaagatcaac
26401 ttcagcgcac tctcaggagc gccgaggctc tcttcaacaa gtactgcgcg ctgactctta

FIG. 2A-7

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26461 aagagtaggc agcgaccgcg cttattcaaa aaaggcggga attacatcat cctcgacatg
26521 agtaaagaaa ttcccacgcc ttacatgtgg agttatcaac cccaaatggg attggcagca
26581 ggcgcctccc aggactactc caccgcgatg aattggctca gcgcggggcc ttctatgatt
26641 tctcgagtta atgatatacg cgcctaccga aaccaatac ttttggaaca gtcagctctt
26701 accaccacgc cccgccaaca ccttaatccc agaaattggc ccgcccctt agtgtaccag
26761 gaaagtcccg ctcccaccac tgtattactt cctcgagacg cccaggccga agtccaaatg
26821 actaatgcag gtgcgaggtt agctggcggc tccaccctat gtcgtcacag gcctcggcat
26881 aatataaaac gcctgatgat cagaggccga ggtatccagc tcaacgacga gtcggtgagc
26941 tctccgcttg gtctacgacc agacggaatc tttcagattg ccggctgcgg gagatcttcc
27001 ttcacccctc gtcaggctgt tctgactttg gaaagtccgt cttcgcaacc ccgctcgggc
27061 ggaatcggga ccgttcaatt tgtagaggag tttactccct ctgtctactt caacccttc
27121 tccggatctc ctgggcaacta cccggacgag ttcataccga acttcgacgc gattagcgag
27181 tcagtggacg gctacgattg atgtctggtg acgcggtga gctatctcgg ctgcgacatc
27241 tagaccactg ccgcgcgttt cgctgctttg cccgggaact tattgagttc atctacttcg
27301 aactccccaa ggatcaccct caaggccggg cccacggagt gcggattact atcgaaggca
27361 aaatagactc tcgcctgcaa cgaattttct cccagcggcc cgtgctgac gagcgagacc
27421 agggaaacac cacggtttcc atctactgca tttgtaata ccccgattg catgaaagcc
27481 tttgctgtct tatgtgtact gagtttaata aaaactgaat taagactctc ctacggactg
27541 ccgcttcttc aaccggatt ttacaaccag aagaacaaaa cttttcctgt cgtccaggac
27601 tctgttaact tcacctttcc tactcaciaa ctagaagctc aacgactaca ccgcttttcc
27661 agaagcattt tccctactaa tactactttc aaaaccggag gtgagctcca cggctctcct
27721 acagaaaacc cttgggtgga agcgggcctt gtagtactag gaattcttgc ggttgggctt
27781 gtgattatcc tttgctacct atacacacct tgcttcaact tcctagtggg gttgtggtat
27841 tgggttataa aatggggccc atactagtct tgcttggttt actttcgctt ttggaaccgg
27901 gttctgcaa ttacgatcca tgtctagact ttgaccaga aaactgcaca cttacttttg
27961 caccgcacac aagccgcac ttgtggagtt ttattaagt cggtatggaa tgcaggcccg
28021 ttgaaattac acacaataac aaaacctgga acaatacctt atccaccaca tgggagccag
28081 gagttcccga gtggtacact gtctctgtcc gaggtcctga cggttccatc cgcattagta
28141 acaacacttt cattttttct gaaatgtgcy atctggccat gttcatgagc aaacagtatt
28201 ctctatggcc tcctagcaag gacaacatcg taacgttctc cattgcttat tgcttggcgc
28261 cttgccttct tactgcttta ctgtgcgtat gcatacacct gcttgaacc actcgcatca
28321 aaaacgcaa taacaaagaa aaaatgcctt aacctcttcc tgtttacaga catggcttct
28381 cttacatctc tcatatttgt cagcattgtc actgccgtc acggacaaac agtctgtctc
28441 atcccactag gacataatta cactctcata ggaccccaa tcacttcaga ggtcatctgg
28501 accaaactgg gaagcgttga ttactttgat ataactgtga acaaaacaaa accaataata
28561 gtaacttgca acatacaaaa tcttacattg attaattgta gcaaagtta cagcggttac
28621 tattatgggt atgacagata cagtagtcaa tatagaaatt acttgggttcg tgttaccag
28681 ttgaaaacca cgaaaatgcc aaatatggca aagattcgat ccgatgacaa ttctctagaa
28741 acttttacat ctcccaccac acccgacgaa aaaaacatcc cagattcaat gattgcaatt
28801 gttgcagcgg tggcagtggt gatggcacta ataataatat gcatgctttt atatgcttgt
28861 cgctacaaaa agtttcatcc taaaaaacia gatctcctac taaggcttaa catttaattt
28921 ctttttatac agccatgggt tccactacca cattccttat gcttactagt ctcgcaactc
28981 tgacttctgc tcgctcacac ctactgttaa ctataggctc aaactgcaca ctaaaaggac
29041 ctcaaggtgg tcatgtcttt tgggtggaga tatatgacaa tggatgggtt acaaaaccat
29101 gtgaccaacc tggtagattt ttctgcaacg gcagagacct aaccattatc aacgtgacag
29161 caaatgacaa aggccttctat tatggaaccg actataaaag tagtttagat tataacatta
29221 ttgtactgcc atctaccact ccagcaccct gcacaactac tttctctagc agcagtgtcg
29281 ctaacaatac aatttccaat ccaacctttg ccgcgctttt aaaacgcact gtgaataatt
29341 ctacaacttc acatacaaca atttccactt caacaatcag catcatcgct gcagtacaa
29401 ttggaatata tattcttgtt tttaccataa cctactacgc ctgctgctat agaaaagaca
29461 aacataaagg tgatccatta cttagatttg atatttaatt tgttcttttt ttttatttac
29521 agtatgggtg acaccaatca tggtagctag aaatttcttc ttcaccatac tcatctgtgc
29581 ttttaatggt tgcgctactt tcacagcagt agccacagca accccagact gtataggagc
29641 atttgcttcc tatgcacttt ttgcttttgt tacttgcatc tgcgtatgta gcatagtctg
29701 cctggttatt aattttttcc aacttctaga ctggatcctt gtgcgaattg cctacctgcg
29761 ccaccatccc gaataccgca accaaaatat cgcggcactt cttagactca tctaaaacca
29821 tgcaggctat actaccaata tttttgcttc tattgcttcc ctacgctgtc tcaacccag
29881 ctgcctatag tactccacca gaacacctta gaaaatgcaa attccaacaa ccgtgggtcat
29941 ttcttgcttg ctatcgagaa aatcagaaa tcccccaaa tttataatg attgctggaa
30001 taattaatat aatctgttgc accataattt catttttgat ataccctcta tttgattttg
30061 gctggaatgc tccaatgca catgatcatc cacaagaccc agaggaacac attccccac
30121 aaaacatgca acatccaata gcgctaatag attacgaaag tgaaccacaa cccccactac
30181 tccctgctat tagttacttc aacctaacgg gcggagatga ctgaaacact caccacctcc

FIG. 2A-8

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30241 aattccgccg aggatctgct cgatatggac ggccgcgtct cagaacaacg acttgcccaa
30301 ctacgcatcc gccagcagca ggaacgcgtg gccaaagagc tcagagatgt catccaaatt
30361 caccaatgca aaaaaggcat attctgtttg gtaaaacaag ccaagatatc ctacgagatc
30421 accgctactg accatcgcct ctcttacgaa cttggccccc aacgacaaaa atttacctgc
30481 atggtgggaa tcaaccccat agttatcacc caacaaagtg gagatactaa gggttgcatt
30541 cactgctcct gcgattccat cgagtgcacc tacaccctgc tgaagaccct atgcggccta
30601 agagacctgc taccaatgaa ttaaaaaaaa atgattaata aaaaatcact tacttgaaat
30661 cagcaataag gtctctgttg aaattttctc ccagcagcac ctacttccc tcttcccaac
30721 tctggtattc taaaccccggt tcagcggcat actttctcca tactttaaa gggatgtcaa
30781 attttagctc ctctcctgta cccacaatct tcatgtcttt cttcccagat gaccaagaga
30841 gtccggctca gtgactcctt caaccctgtc taccctatg aagatgaaag cacctcccaa
30901 cacccttta taaacccagg gtttatttcc ccaaatggct tcacacaaa cccagacgga
30961 gttcttactt taaaatgttt aacccacta acaaccacag gcggatctct acagctaaaa
31021 gtgggagggg gacttacagt ggatgacact gatggtacct tacaagaaaa catactgtct
31081 acagcaccca ttactaaaaa taatcactct gtagaactat ccattggaaa tggattagaa
31141 actcaaaaaca ataaactatg tgccaaattg ggaaatgggt taaaatttaa caacggtgac
31201 atttgtataa aggatagtat taacacctta tggactggaa taaacctctc acctaaactgt
31261 caaattgttg aaaacactaa tacaatgat ggcaactta ctttagtatt agtaaaaaat
31321 ggagggcttg ttaatggcta cgtgtctcta gttggtgtat cagacactgt gaaccaaatg
31381 ttcacacaaa agacagcaaa catccaatta agattatatt ttgactcttc tggaaatcta
31441 ttaactgagg aatcagactt aaaaattcca cttaaaaata aatcttctac agcgaccagt
31501 gaaactgtag ccagcagcaa agcctttatg ccaagtacta cagcttatcc cttcaacacc
31561 actactaggg atagtgaaaa ctacattcat ggaatatgtt actacatgac tagttatgat
31621 agaagtctat ttcccttgaa catttctata atgctaaaca gccgtatgat ttcttccaat
31681 gttgcctatg ccatacaatt tgaatggaat ctaaattgcaa gtgaatctcc agaaagcaac
31741 atagctacgc tgaccacatc ccccttttct ttttcttaca ttacagaaga cgacaactaa
31801 aataaagttt aagtgttttt atttaaaatc acaaaattcg agtagttatt ttgcctccac
31861 cttcccattt gacagaatac accaatctct ccccacgcac agctttaaac atttggtac
31921 cattagagat agacattgtt ttagattcca cattccaaac agtttcagag cgagccaatc
31981 tggggtcagt gatagataaa aatccatcgc gatagtcttt taaagcgctt tcacagtcca
32041 actgctgcgg atgcgactcc ggagtttgga tcacggtcat ctggaagaag aacgatggga
32101 atcataatcc gaaaacggta tcggacgatt gtgtctcatc aaaccacaa gcagccgctg
32161 tctgcgtcgc tccgtgcgac tgctgtttat gggatcaggg tccacagttt cctgaagcat
32221 gattttaata gcccttaaca tcaactttct ggtgcgatgc gcgcagcaac gcattctgat
32281 ttcactcaaa tctttgcagt aggtacaaca cattattaca atattgttta ataaaccata
32341 attaaaagcg ctccagccaa aactcatatc tgatataatc gccctgcat gaccatcata
32401 ccaaagttta atataaatta aatgacgttc cctcaaaaac acactacca catacatgat
32461 ctcttttggc atgtgcatat taacaatctg tctgtaccat ggacaacggt ggttaatcat
32521 gcaaccaat ataaccttc ggaaccacac tgccaacacc gctccccag ccatgcattg
32581 aagtgaacc tgctgattac aatgacaatg aagaacccaa ttctctcgac cgtgaatcac
32641 ttgagaatga aaaatatcta tagtggcaca acatagacat aaatgcatgc atcttctcat
32701 aatttttaac tcttcaggat ttagaaacat atcccaggga ataggaagct cttgcagaac
32761 agtaaagctg gcagaacaag gaagaccacg aacacaactt acactatgca tagtcatagt
32821 atcacatct ggcaacagcg ggtggtcttc agtcatagaa gctcgggttt cattttctc
32881 acaacgtggt aactgggctc tgggtgtaagg gtgatgtctg gcgcatgatg tcgagcgtgc
32941 gcgcaacctt gtcataatgg agttgcttcc tgacattctc gtattttgta tagcaaaacg
33001 cggccctggc agaacacact cttcttcgcc ttctatcctg ccgcttagcg tgttccgtgt
33061 gatagttcaa gtacagccac actcttaagt tgggtcaaaag aatgctggct tcagttgtaa
33121 tcaaaactcc atcgcatcta attgttctga ggaaatcatc cacggtagca tatgcaaatc
33181 ccaaccaagc aatgcaactg gattgcgttt caagcaggag aggagaggga agagacggaa
33241 gaaccatgtt aatttttatt ccaaacgatc tcgcagtact tcaaattgta gatcgcgag
33301 atggcatctc tcgccccac tgtgttggtg aaaaagcaca gctaaatcaa aagaaatgcg
33361 attttcaagg tgctcaacgg tggcttccaa caaagcctcc acgcgacat ccaagaacaa
33421 aagaatacca aaagaaggag cattttctaa ctctcaatc atcatattac attcctgcac
33481 cattcccaga taattttcag ctttccagcc ttgaattatt cgtgtcagtt cttgtggtaa
33541 atccaatcca cacattacaa acaggtcccg gagggcgccc tccaccacca ttcttaaaaa
33601 caccctcata atgacaaaat atcttgctcc tgtgtcacct gtagcgaatt gagaatggca
33661 acatcaattg acatgccctt ggctctaagt tcttctttaa gttctagtgt taaaaactct
33721 ctcatattat caccaaactg cttagccaga agcccccgga gaacaagagc aggggacgct
33781 acagtgcagt acaagcgcag acctcccaa ttggctccag caaaaacaag attggaataa
33841 gcatattggg aaccaccagt aatatcatcg aagttgctgg aaatataatc aggcagagtt
33901 tcttgtagaa attgaataaa agaaaaattt gccaaaaaaa cattcaaac ctctgggatg
33961 caaatgcaat aggttaccgc gctgcgctcc aacattgtta gttttgaatt agtctgcaa

FIG. 2A-9

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```
34021 aataaaaaaa aaacaagcgt catatcatag tagcctgacg aacaggtgga taaatcagtc
34081 ttccatcac aagacaagcc acaggggtctc cagctcgacc ctcgtaaaac ctgtcatcgt
34141 gattaaacaa cagcaccgaa agttcctcgc ggtgaccagc atgaataagt cttgatgaag
34201 catacaatcc agacatgtta gcatcagtta aggagaaaaa acagccaaca tagcctttgg
34261 gtataattat gcttaatcgt aagtatagca aagccacccc tcgoggatac aaagtaaaag
34321 gcacaggaga ataaaaaata taattatttc tctgctgctg tttaggcaac gtcgcccccg
34381 gtccctctaa atacacatac aaagcctcat cagccatggc ttaccagaga aagtacagcg
34441 ggcacacaaa ccacaagctc taaagtcact ctccaacctc tccacaatat atatacacia
34501 gccctaaact gacgtaatgg gactaaagtg taaaaaatcc cgccaaaccc aacacacacc
34561 ccgaaactgc gtcaccaggg aaaagtacag tttcacttcc gcaatcccaa caagcgtcac
34621 ttcctctttc tcacggtacg tcacatccca ttaacttaca acgtcatttt cccacggccg
34681 cgccgccccct ttttaaccgtt aaccccacag ccaatcacca cacggcccac acttttttaa
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SEQ ID NO: 1
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FIG. 2A-10

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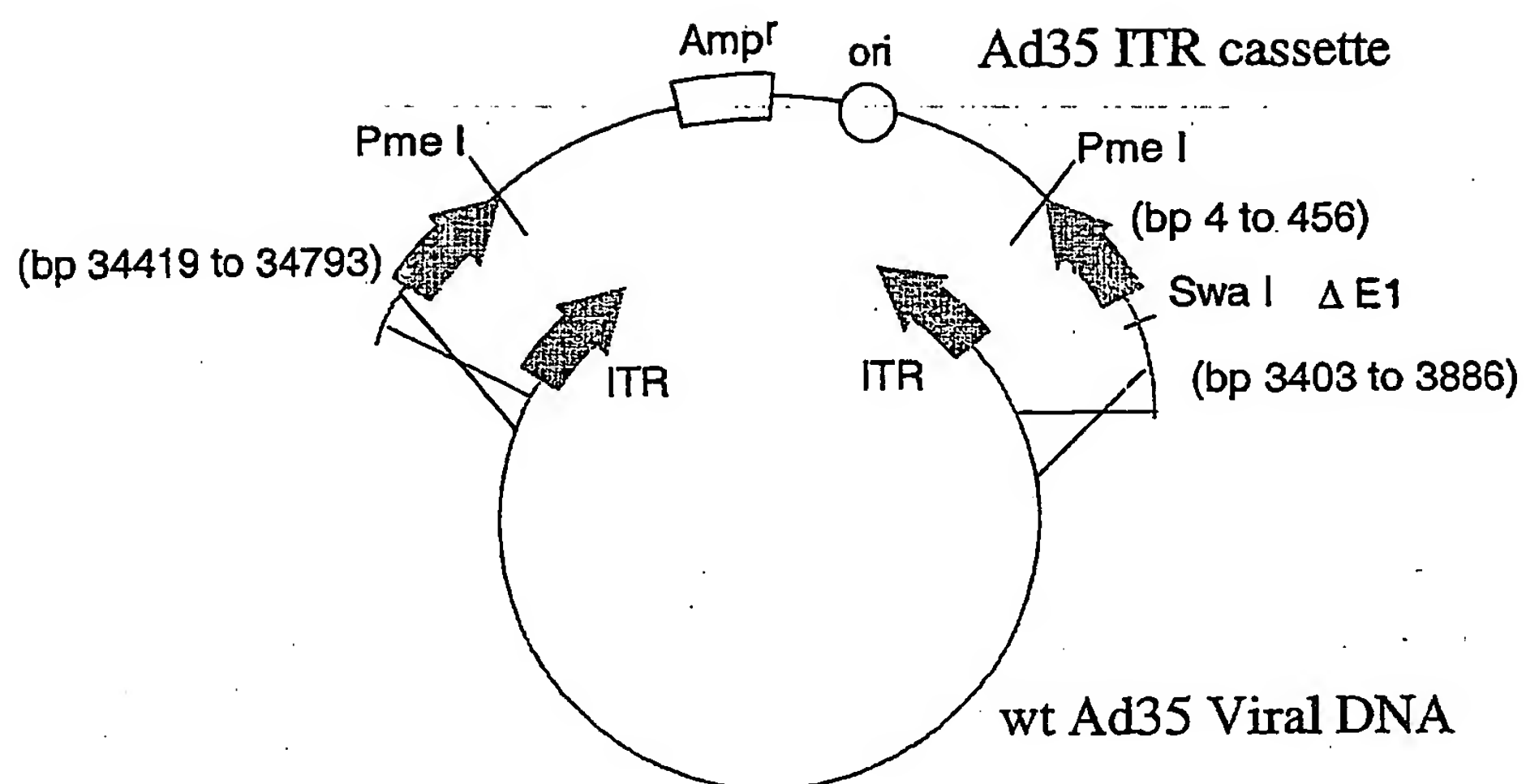


FIG. 3

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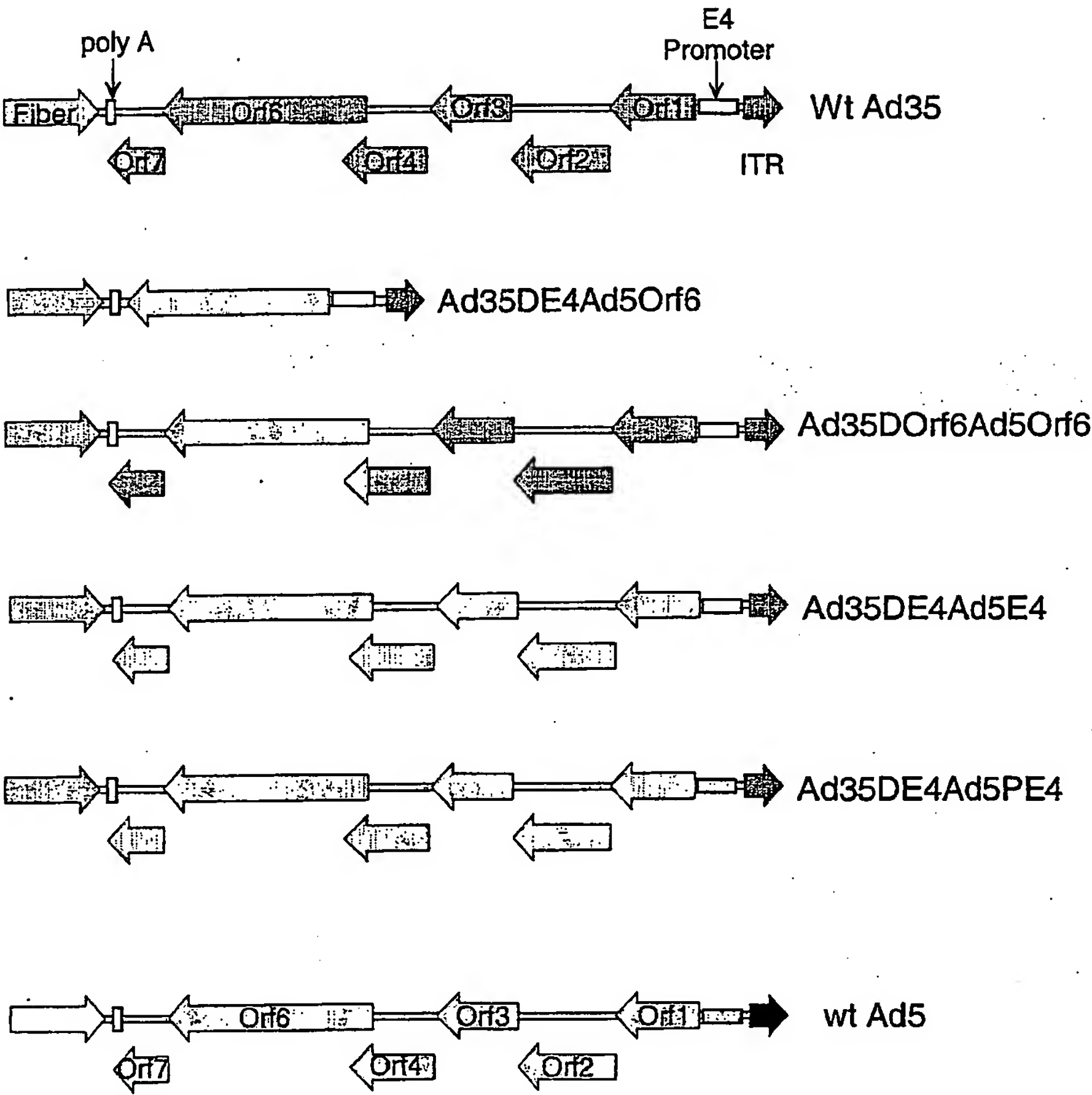


FIG. 4

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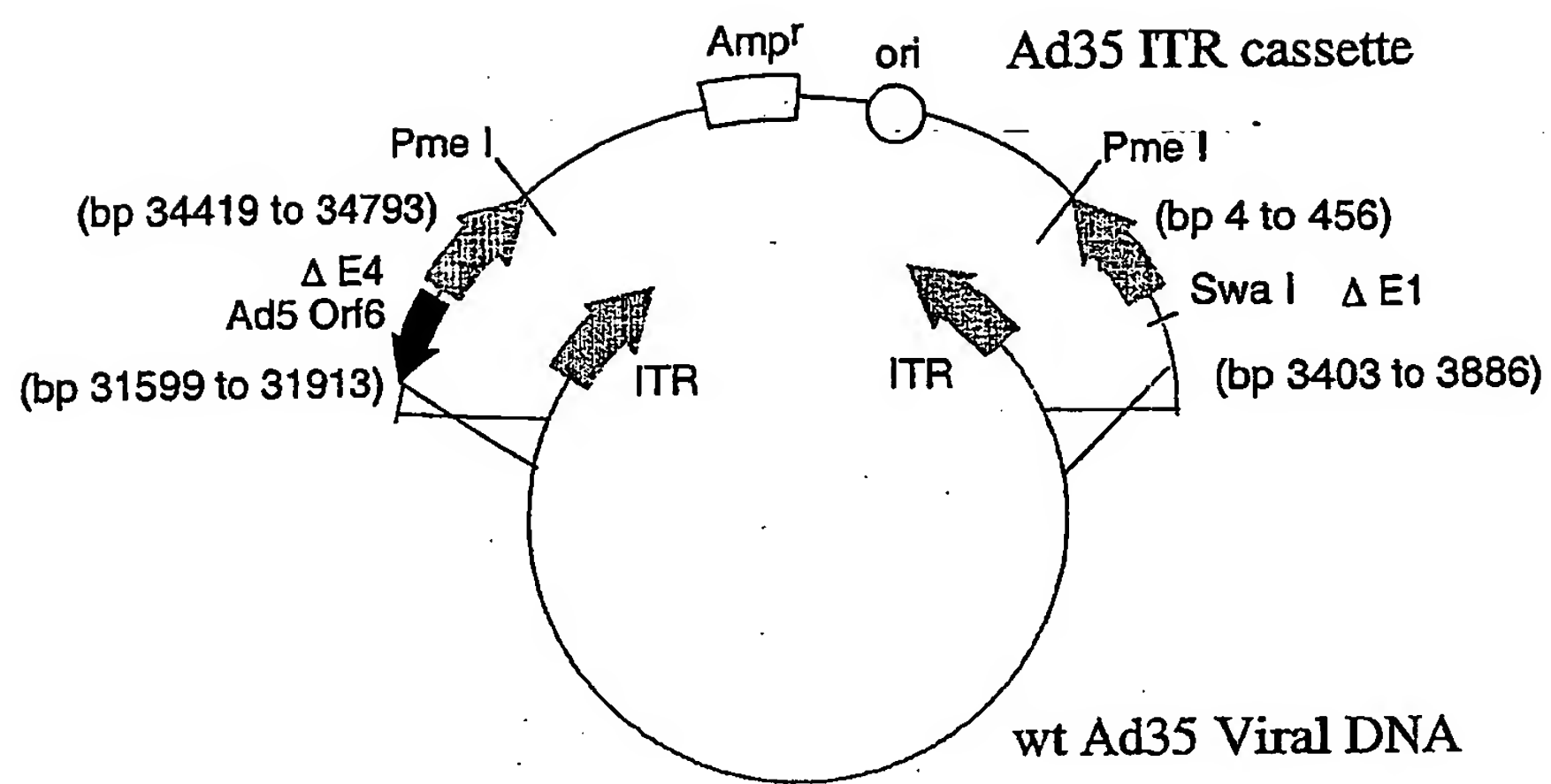


FIG. 5

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1 ccattgcata cgttgtatcc atatcataat atgtacattt atattggctc atgtccaaca
61 ttaccgccat gttgacattg attattgact agttattaat agtaatcaat tacgggggtca
121 ttagttcata gcccatatat ggagttccgc gttacataac ttacggtaaa tggcccgccct
181 ggctgaccgc ccaacgaccc cggcccatg acgtcaataa tgacgtatgt tcccatagta
241 acgccaatag ggactttcca ttgacgtcaa tgggtggagt atttacggta aactgcccac
301 ttggcagtac atcaagtgt tcatatgcc agtacgccc ctattgacgt caatgacggt
361 aaatggcccg cctggcatta tgcccagtac atgaccttat gggactttcc tacttggcag
421 tacatctacg tattagtcat cgctattacc atggtgatgc ggttttggca gtacatcaat
481 gggcgtggat agcggttga ctacgggga tttccaagtc tccaccccat tgacgtcaat
541 gggagtttgt tttggcacca aaatcaacgg gactttccaa aatgtcgtaa caactccgcc
601 ccattgacgc aaatgggagg taggcgtgta cggtgggagg tctatataag cagagctcgt
661 ttagtgaacc gtcagatcgc ctggagacgc catccacgct gttttgacct ccatagaaga
721 caccgggacc gatccagcct ccgcgccgg gaacgggtgca ttggaacgcg gattccccgt
781 gccaagagtg agatctacca TGGGTGCTAG GGCTTCTGTG CTGTCTGGTG GTGAGCTGGA
841 CAAGTGGGAG AAGATCAGGC TGAGGCCTGG TGGCAAGAAG AAGTACAAGC TAAAGCACAT
901 TGTGTGGGCC TCCAGGGAGC TGGAGAGGTT TGCTGTGAAC CCTGGCCTGC TGGAGACCTC
961 TGAGGGGTGC AGGCAGATCC TGGGCCAGCT CCAGCCCTCC CTGCAAACAG GCTCTGAGGA
1021 GCTGAGGTCC CTGTACAACA CAGTGGCTAC CCTGTACTGT GTGCACCAGA AGATTGATGT
1081 GAAGGACACC AAGGAGGCC TGGAGAAGAT TGAGGAGGAG CAGAACAAGT CCAAGAAGAA
1141 GGGCCAGCAG GCTGCTGCTG GCACAGGCCA CTCCAGCCAG GTGTCCCAGA ACTACCCCAT
1201 TGTGCAGAAC CTCCAGGGCC AGATGGTGCA CCAGGCCATC TCCCCCGGA CCCTGAATGC
1261 CTGGGTGAAG GTGGTGGAGG AGAAGGCCTT CTCCCCTGAG GTGATCCCCA TGTCTCTGC
1321 CCTGTCTGAG GGTGCCACCC CCCAGGACCT GAACACCATG CTGAACACAG TGGGGGGCCA
1381 TCAGGCTGCC ATGCAGATGC TGAAGGAGAC CATCAATGAG GAGGCTGCTG AGTGGGACAG
1441 GCTGCATCCT GTGCACGCTG GCCCCATTGC CCCCAGCCAG ATGAGGGAGC CCAGGGGCTC
1501 TGACATTGCT GGCACCACCT CCACCCTCCA GGAGCAGATT GGCTGGATGA CCAACAACCC
1561 CCCCATCCCT GTGGGGGAAA TCTACAAGAG GTGGATCATC CTGGGCCTGA ACAAGATTGT
1621 GAGGATGTAC TCCCCACCT CCATCCTGGA CATCAGGCAG GGCCCCAAGG AGCCCTTCAG
1681 GGAATATGTG GACAGGTTCT ACAAGACCCT GAGGGCTGAG CAGGCCTCCC AGGAGGTGAA
1741 GAAGTGGATG ACAGAGACCC TGCTGGTGCA GAATGCCAAC CCTGACTGCA AGACCATCCT
1801 GAAGGCCCTG GGCCCTGCTG CCACCCTGGA GGAGATGATG ACAGCCTGCC AGGGGGTGGG
1861 GGGCCCTGGT CACAAGGCCA GGGTGCTGGC TGAGGCCATG TCCCAGGTGA CCAACTCCGC
1921 CACCATCATG ATGCAGAGGG GCAACTTCAG GAACCAGAGG AAGACAGTGA AGTGCTTCAA
1981 CTGTGGCAAG GTGGGCCACA TTGCCAAGAA CTGTAGGGCC CCCAGGAAGA AGGGCTGCTG
2041 GAAGTGTGGC AAGGAGGGCC ACCAGATGAA GGAATGCAAT GAGAGGCAGG CCAACTTCCT
2101 GGGCAAAATC TGGCCCTCCC ACAAGGGCAG GCCTGGCAAC TTCTCCAGT CCAGGCCTGA
2161 GCCCACAGCC CCTCCCGAGG AGTCCTTCAG GTTTGGGGAG GAGAAGACCA CCCCAGCCA
2221 GAAGCAGGAG CCCATTGACA AGGAGCTGTA CCCCCTGGCC TCCCTGAGGT CCCTGTTTGG
2281 CAACGACCCC TCCTCCAGT AAaataaagc ccgggcagat ctgatctgct gtgccttcta
2341 gttgccagcc atctgttgtt tgcccctccc ccgtgccttc cttgaccctg gaaggtgcca
2401 ctcccactgt cttttcctaa taaaatgagg aaattgcatc gcattgtctg agtaggtgtc
2461 attctattct ggggggtggg gtggggcagc acagcaaggg ggaggattgg gaagacaata
2521 gcaggcatgc tggggatgcg gtgggctcta

SEQ ID NO: 2

FIG. 6

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1 ccattgcata cgttgtatcc atatcataat atgtacattt atattggctc atgtccaaca
61 ttaccgccat gttgacattg attattgact agttattaat agtaatcaat tacgggggtca
121 ttagttcata gcccatatat ggagttccgc gttacataac ttacggtaaa tggcccgcct
181 ggctgaccgc ccaacgaccc ccgcccattg acgtcaataa tgacgtatgt tcccatagta
241 acgccaatag ggactttcca ttgacgtcaa tgggtggagt atttacggta aactgcccac
301 ttggcagtac atcaagtgt tcatatgcc agtacgcccc ctattgacgt caatgacggg
361 aaatggcccg cctggcatta tgcccagtac atgaccttat gggactttcc tacttggcag
421 tacatctacg tattagtcac cgctattacc atgggtgatgc ggttttggca gtacatcaat
481 gggcgtggat agcggtttga ctcacgggga tttccaagtc tccaccccat tgacgtcaat
541 gggagtttgt tttggcacca aaatcaacgg gactttccaa aatgtcgtaa caactccgcc
601 ccattgacgc aaatgggccc taggcgtgta cgggtgggagg tctatataag cagagctcgt
661 ttagtgaacc gtcagatcgc ctggagacgc catccacgct gttttgacct ccatagaaga
721 caccgggacc gatccagcct ccgcgcccg gaacgggtgca ttggaacgcg gattccccgt
781 gccaaagagt agatcgatct aagtaagctt CCTGCATGCT GCTGCTGCTG CTGCTGCTGG
841 GCCTGAGGCT ACAGCTCTCC CTGGGCATCA TCCAGTTGA GGAGGAGAAC CCGGACTTCT
901 GGAACCGCGA GGCAGCCGAG GCCCTGGGTG CCGCAAGAA GCTGCAGCCT GCACAGACAG
961 CCGCAAGAA CCTCATCATC TTCCTGGGCG ATGGGATGGG GGTGTCTACG GTGACAGCTG
1021 CCAGGATCCT AAAAGGGCAG AAGAAGGACA AACTGGGGCC TGAGATACCC CTGGCCATGG
1081 ACCGCTTCCC ATATGTGGCT CTGTCCAAGA CATAAATGT AGACAAACAT GTGCCAGACA
1141 GTGGAGCCAC AGCCACGGCC TACCTGTGCG GGTCAAGGG CAACTTCCAG ACCATTGGCT
1201 TGAGTGCAGC CGCCCGCTTT AACCAGTGCA ACACGACACG CGGCAACGAG GTCATCTCCG
1261 TGATGAATCG GGCCAAGAAA GCAGGGAAGT CAGTGGGAGT GGTAACCACC ACACGAGTGC
1321 AGCACGCCTC GCCAGCCGGC ACCTACGCCC ACACGGTGAA CCGCAACTGG TACTCGGACG
1381 CCGACGTGCC TGCC'TCCGCC CGCCAGGAGG GGTGCCAGGA CATCGCTACG CAGCTCATCT
1441 CCAACATGGA CATTGACGTG ATCCTAGGTG GAGGCCGAAA GTACATGTTT CGCATGGGAA
1501 CCCCAGACCC TGAGTACCCA GATGACTACA GCCAAGGTGG GACCAGGCTG GACGGGAAGA
1561 ATCTGGTGCA GGAATGGCTG GCGAAGCGCC AGGGTGCCCG GTATGTGTGG AACC GCACTG
1621 AGCTCATGCA GGCTTCCCTG GACCCGTCTG TGACCCATCT CATGGGTCTC TTTGAGCCTG
1681 GAGACATGAA ATACGAGATC CACCGAGACT CCACACTGGA CCCCTCCCTG ATGGAGATGA
1741 CAGAGGCTGC CCTGCGCCTG CTGAGCAGGA ACCCCGCGG CTTCTTCCTC TTCGTGGAGG
1801 GTGGTCGCAT CGACCATGGT CATCATGAAA GCAGGGCTTA CCGGGCACTG ACTGAGACGA
1861 TCATGTTTCA CGACGCCATT GAGAGGGCGG GCCAGCTCAC CAGCGAGGAG GACACGCTGA
1921 GCCTCGTCAC TGCCGACCAC TCCCACGTCT TCTCCTTCGG AGGCTACCCC CTGCGAGGGA
1981 GCTCCATCTT CGGGCTGGCC CCTGGCAAGG CCCGGGACAG GAAGGCCCTAC ACGGTCCTCC
2041 TATACGGAAG CGGTCCAGGC TATGTGCTCA AGGACGGCGC CCGGCCGGAT GTTACCGAGA
2101 GCGAGAGCGG GAGCCCCGAG TATCGGCAGC AGTCAGCAGT GCCCTGGAC GAAGAGACCC
2161 ACGCAGGCGA GGACGTGGCG GTGTTGCGCG GCGGCCCGCA GGCGCACCTG GTTCACGGCG
2221 TGCAGGAGCA GACCTTCATA GCGCACGTCA TGGCCTTCGC CGCCTGCCTG GAGCCCTACA
2281 CCGCCTGCGA CCTGGCGCCC CCCGCCGGCA CCACCGACGC CGCGCACCCG GGTAAcccg
2341 tgggtccccgc gttgcttctt ctgctggccg ggacatcagg tggccccgc tgaattggaa
2401 tcgatcagaa ttgatctgat ctgctgtgcc ttctagttgc cagccatctg ttgtttgccc
2461 ctcccccggtg ccttccttga ccttgggaagg tgccactccc actgtccttt cctaataaaa
2521 tgaggaaatt gcatcgcat gtctgagtag gtgtcattct attctggggg gtgggggtggg
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2641 ctcta
SEQ ID NO: 3

FIG. 7

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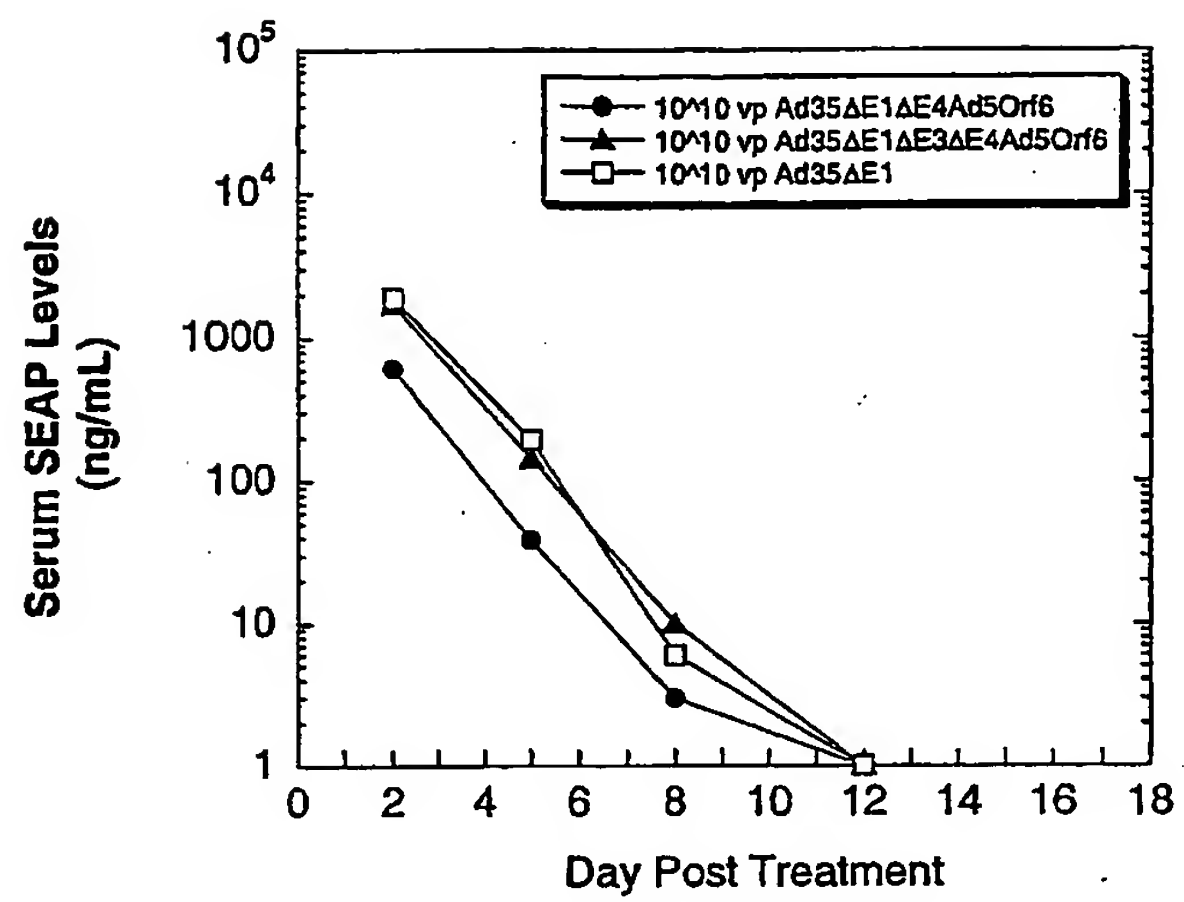


FIG. 8

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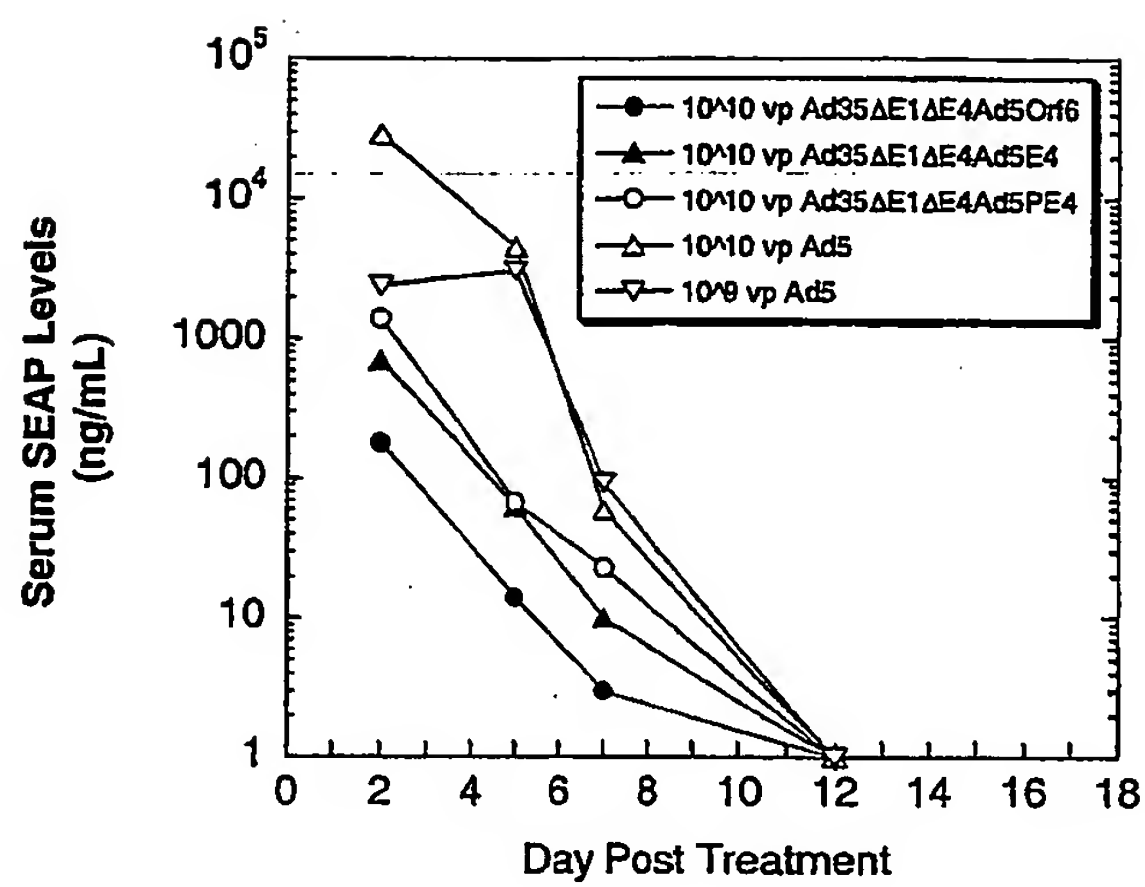


FIG. 9

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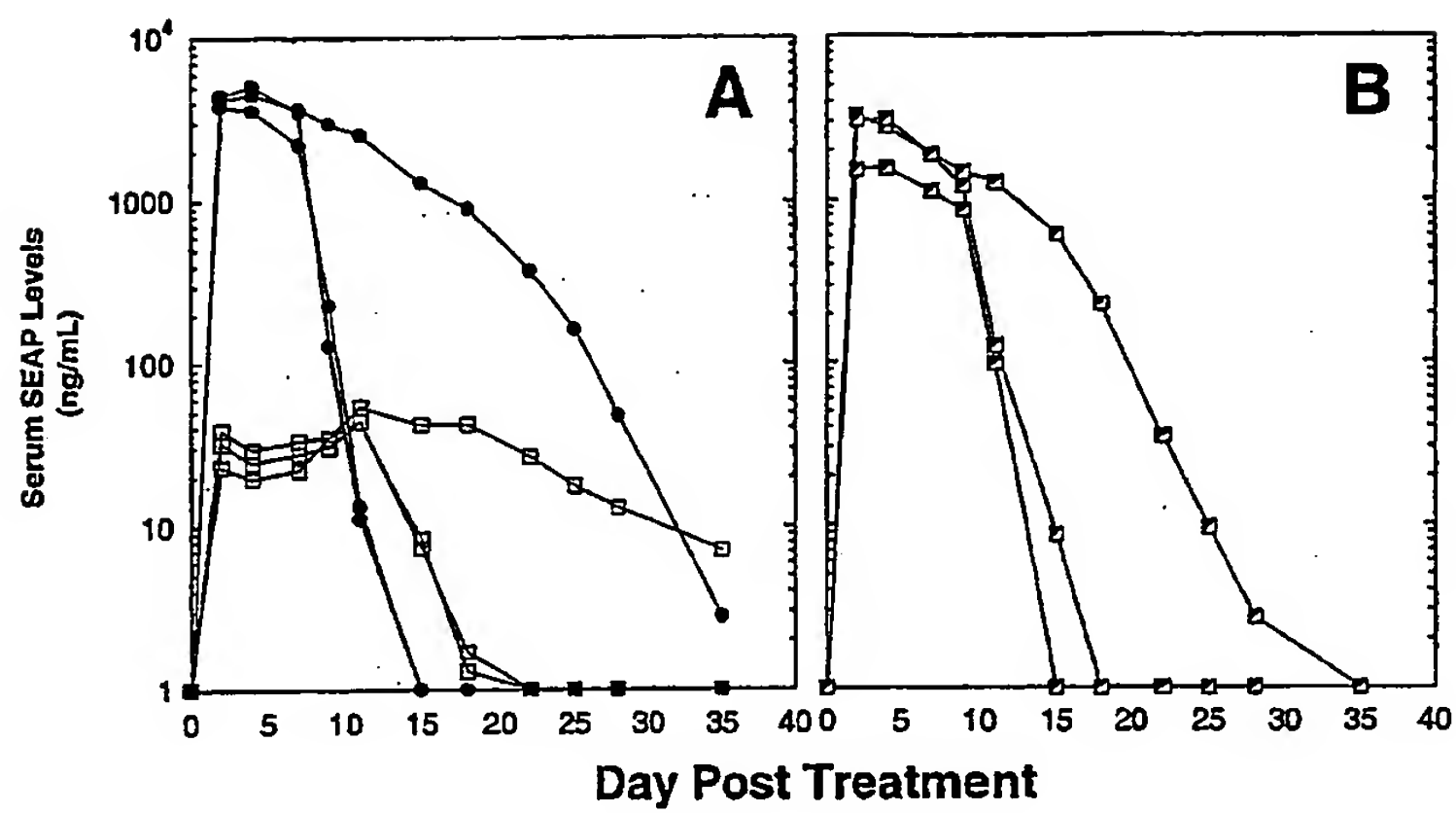


FIG. 10A-B

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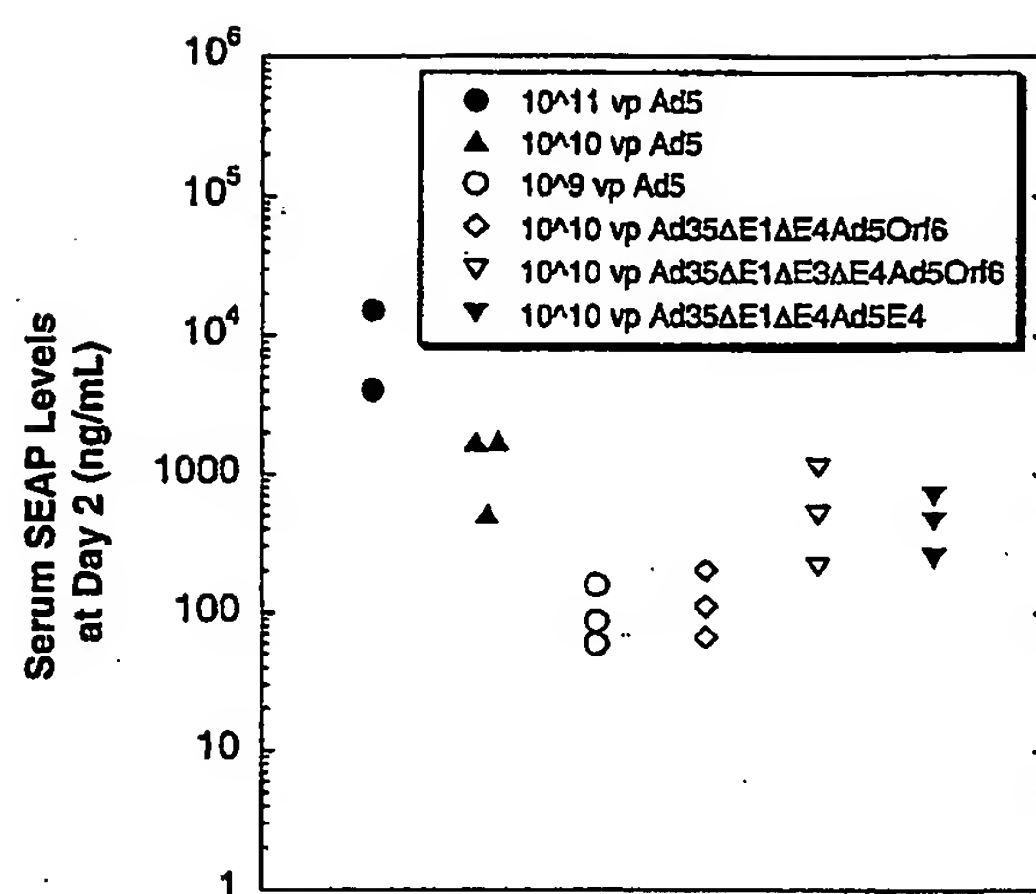


FIG. 11

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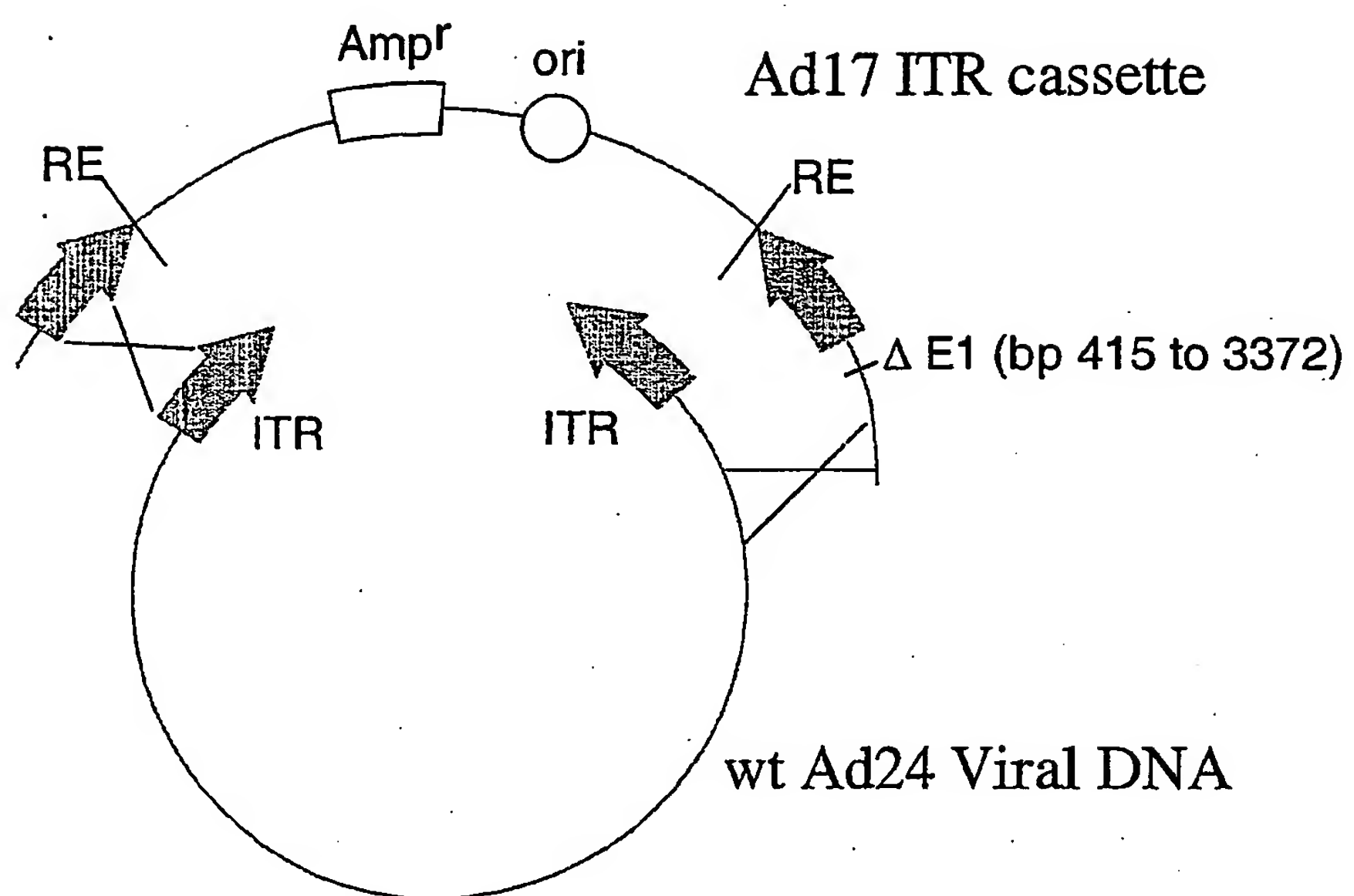


FIG. 12

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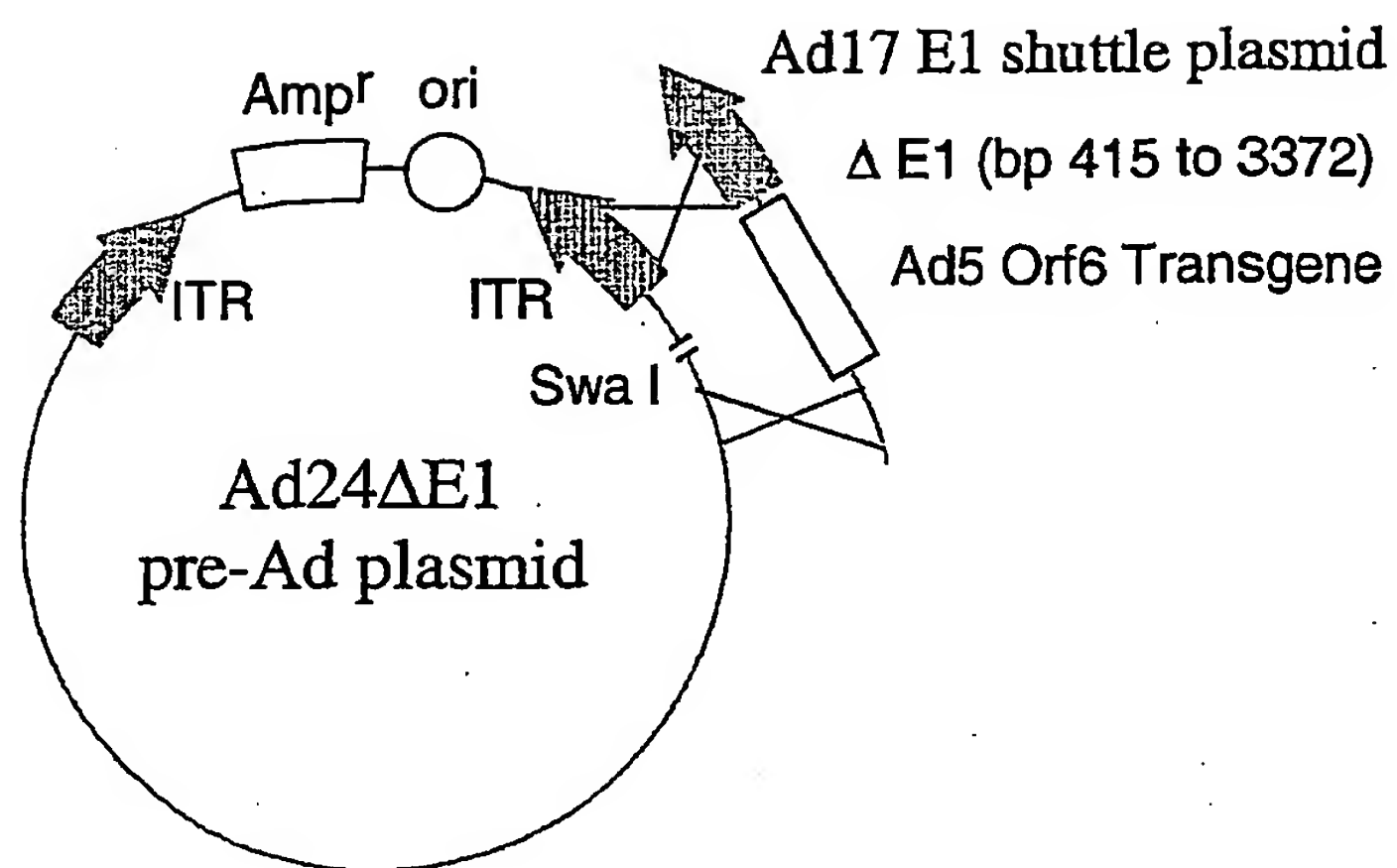


FIG. 13

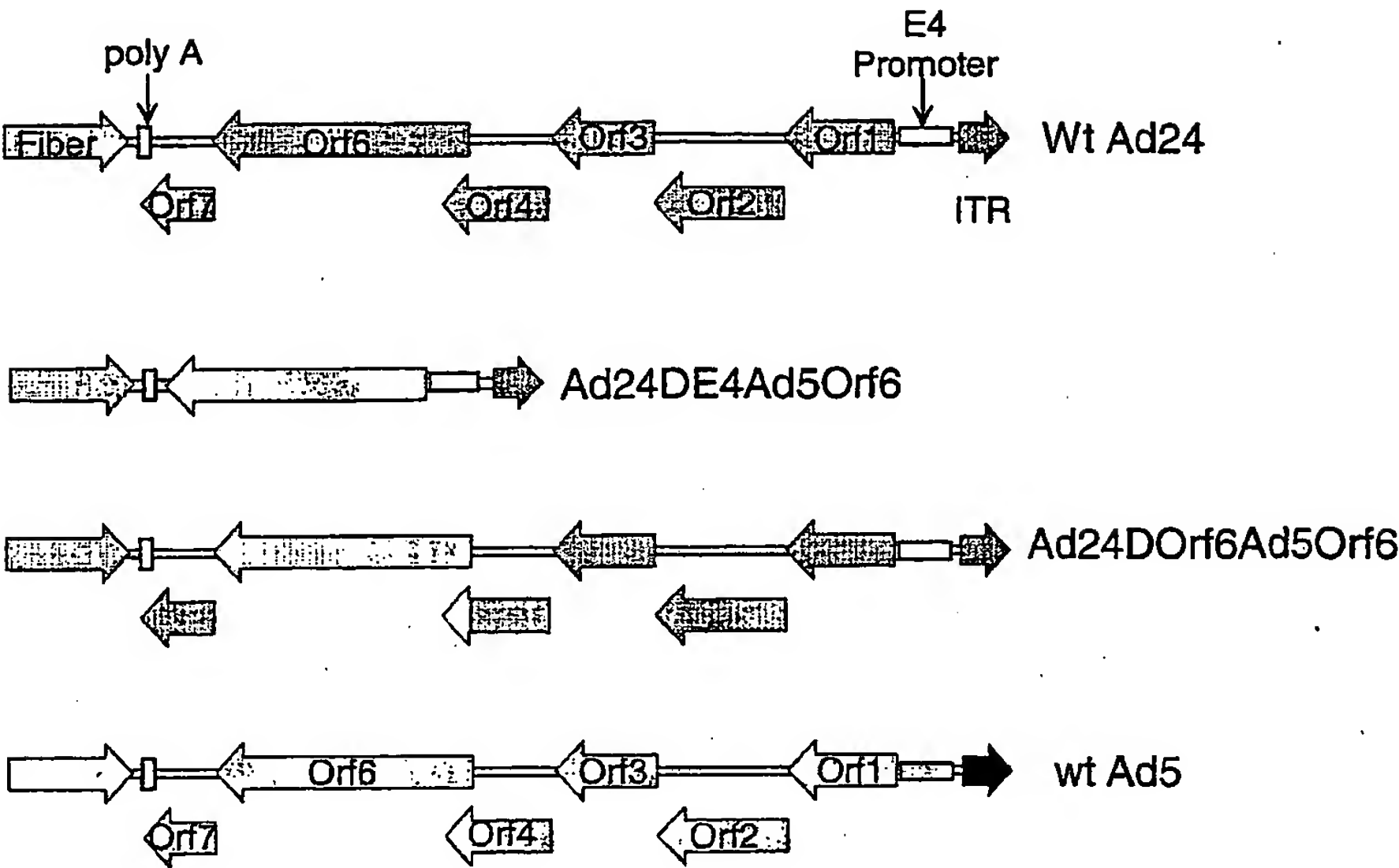


FIG. 14

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Growth Curve Comparison of Ad24 Based Vectors

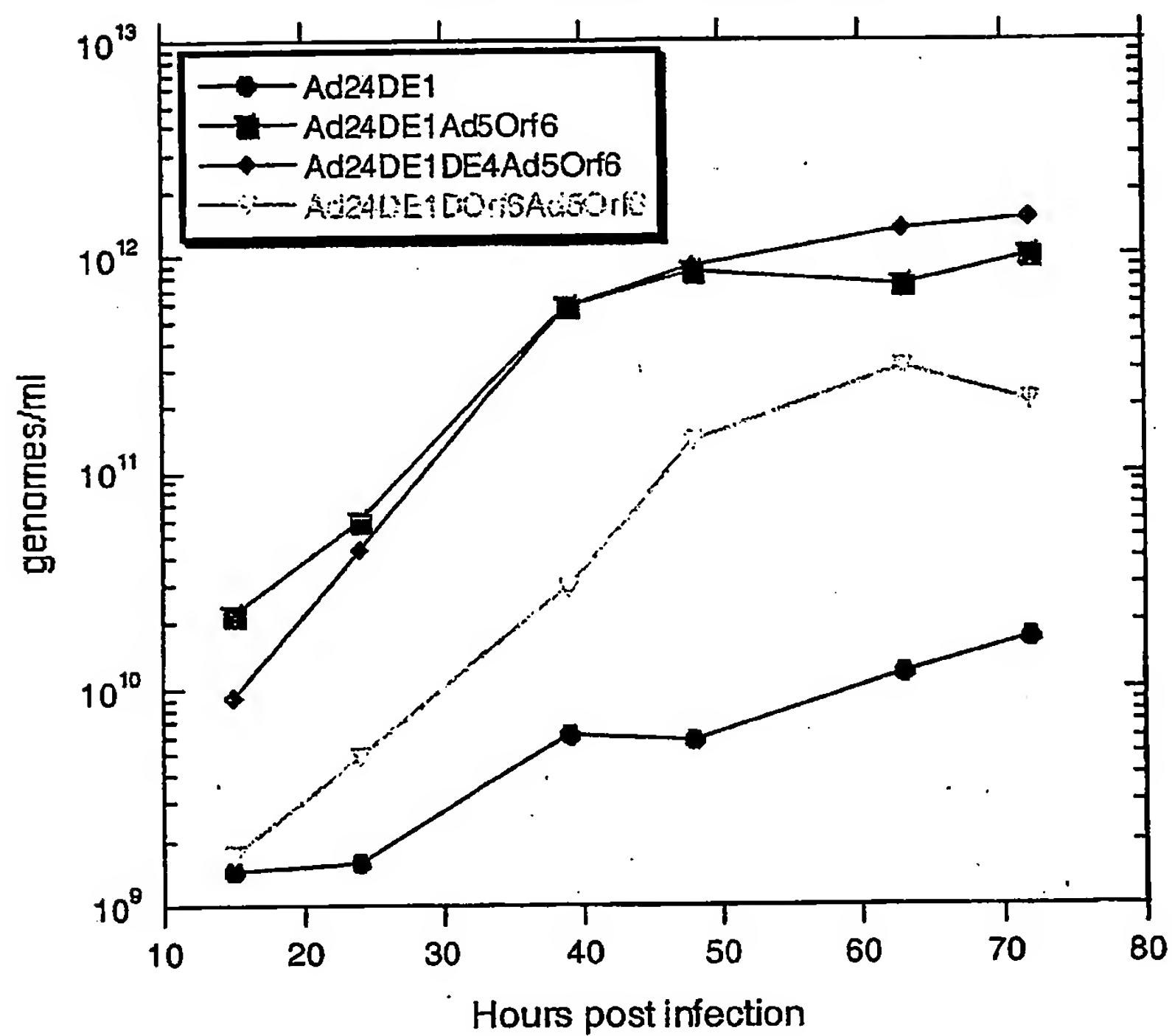


FIG. 15

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61 tttagggcgg ggccagcgct gattggacga gagaagatga tgcaaatagac gtcacgacgc
121 acggctaacg gtcgcccggg aggcgtggcc tagcccggaa gcaagtcgcg gggctgatga
181 cgtataaaaa agcggacttt agacccggaa acggccgatt ttcccggcggc cacgcccggg
241 tatgaggtaa ttctggggcg atgcaagtaa aattaggtca ttttggcgcg aaaactgaat
301 gaggaagtga aaagtgaaaa ataccgggtcc cgcccagggc ggaatattta ccgagggccg
361 agagactttg accgattacg tgggggtttc gattgcggtg ttttttcgcg aatttcgcg
421 tccgtgtcaa agtccggtgt ttatgtcaca gatcagctga tccacagggt atttaaacca
481 gtcgagcccg tcaagaggcc actcttgagt gccagcgagt agagatttct ctgagctccg
541 ctcccagagt ctgagaaaaa tgagacacct gcgcctcctt tcttcaactg tgcctattga
601 catggccgca ttattgctgg aggattatgt gagtacaata ttggaggacg aactgcatcc
661 atctccattt gagctgggac ctacacttca ggacctatat gatttggagg tagatgccca
721 tgatgacgac ccgaacgaag aggctgtgaa tttaatat ttccagaatctc tgattcttca
781 ggctgacata gccagcgaag ctgtacctac accacttcat acaccgactc tgtcaccat
841 acctgaattg gaagaggagg acgagctaga cctccgatgt tatgaggaag gttttcctcc
901 cagcgattca gaggacgaac aggggtgagca gagcatggct ctaatctcaa aatatgcttg
961 tgtggttgtg gaagagcatt ttgtgttgga caatcctgag gtgcccgggc aaggctgtag
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1381 cctttggacc tgagcttgaa acgccccagg aactaggctc agctgtgctt agtcatgtgt
1441 aaataaagtt gtacaataaa agtatatgtg acgcatgcaa ggtgtggttt atgactcatg
1501 ggcgtggctt agtcctatat aagtggcaac acctgggcac tggggcacag accttcaggg
1561 agttcctgat ggatgtgtgg actatccttg cagactttag caagacacgc cggctgttag
1621 aggatagttc agacgggtgc tccgggttct ggagacactg gtttggaaact cctctatctc
1681 gtctggtgta cacagttaag aaggattata acgaggaatt tgaaaatctt tttgctgatt
1741 gctctggcct gctagattct ctaaactctcg gccaccagtc cttttccag gaaagggtac
1801 tccacagcct tgatttttca agcccagggc gcactacagc cggggttgct tttgtggttt
1861 ttctggttga caaatggagc cagaacacc aactgagcag gggctacatt ctggacttcg
1921 cagccatgca cctgtggagg gcatgggtga ggcagcggg acagagaatc ttgaactact
1981 ggcttataca gccagcagct ccgggtcttc ttctctaca cagacaaaca tccatgttgg
2041 aggaagaaat gaggcaggcc atggacgaga acccgaggag cggcctggac cctccgtcgg
2101 aagaggagct ggattgaatc aggtatccag cctgtacca gagcttagca ggggtgctgac
2161 atccatggcc aggggagtg agagggagag gagcgatggg ggcaataccg ggatgatgac
2221 cgagctgacg gccagcctga tgaatcgcaa gcgtccagag cgcattacct ggcacgagct
2281 acagatggag tgtagggatg aggtgggcct gatgcaggat aaatatggcc tggagcagat
2341 aaaaaccac tggttgaacc cagatgagga ttgggaggag gccattaaga aatatgccaa
2401 gatagccctg cgcccagatt gcaagtacag ggtgaccaag acggtgaata tcagacatgc
2461 ctgctacatc tcggggaacg gggcagaggt ggtcatcgat accctggaca aggccgcctt
2521 caggtgttgc atgatgggaa tgagagccgg agtgatgaat atgaattcca tgattttcat
2581 gaacatgaag ttcaatggag agaagtttaa tggggtgatg ttcatggcca acagtcacat
2641 gaccctgcac ggctgcagtt tcttcggctt caacaatatg tgcgagagg tctggggcgc
2701 tgctaagatc aggggatgta agttttatgg ctgctggatg ggcgtggtcg gaagacccaa
2761 gagegagatg tctgtgaagc agtgtgtgtt tgagaaatgc tacctgggag tctctaccga
2821 gggcaatgct agagtgagac attgctcttc cctggagacg ggctgcttct gcctggtgaa
2881 gggcacagcc tctctgaagc ataatatggt gaagggtgc acggatgagc gcatgtacaa
2941 catgctgaca tgcgactcgg ggtctgcca tatcctgaag aacatccatg tgacctcca
3001 cccccggaag aagtggccag tgtttgagaa taacctactg atcaagtgcc acatgcacct
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3121 gctggagaac gatgccttct ccagggtgaa cctgaacggc atctttgaca tggatgtctc
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3241 gggcagacac accaggatgc aaccagtggc cctggatgtg accgaggagc tgaggcccg
3301 ccacctggtg atggcttgta ccgggaccga gttcagctcc agtggggagg acacagatta
3361 gaggtaggtt gagtattagt gggcgtggct aaggtgacta taaaggcggg tgtcttacga
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3481 ctttttagcc cttatttgac aaccgcctg ccgggatggg ccggagtctc tcagaatgtg
3541 atgggatcga cgggtggacg gcgtccagtg cttccagcaa attcctcgac catgacctac
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```

FIG. 16A-1

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3661 atgacagcga cgagactggc ttcgagctac atgcccagca gcagcagtag cccctctgtg
3721 cccagttcca tcatcgccga ggagaaactg ctggccctgc tggccgagct ggaagccctg
3781 agccgccagc tggccgccct gaccagcag gtgtccgagc tccgcgaaca gcagcagcag
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4201 tgcgggggga gatgatgtgg agtttggcct ggatcttgag gttggcgatg ttgccacca
4261 gatccgcct ggggttcattg ttgtgcagga ccaccagaac ggtgtagccc gtgcacttgg
4321 ggaacttgte atgcaacttg gaagggaatg cgtgaaagaa tttggagacg cccttgtgcc
4381 caccaggtt ttccatgcac tcatccatga tgatggcgat gggcccgtgg gctgcggctt
4441 tggcaaagac gtttctgggg tcagagacat cgtaattatg ctcctgggtg agatcatcat
4501 aagacatttt aatgaatttg gggcgagggg tgccagattg ggggacaatg gttccctcgg
4561 gccccggggc gaagttcccc tcacatattt gcatctcca ggctttcatc tcggaggggg
4621 ggatcatgtc cacctgcggg gcgatgaaaa aaacggttcc cggggcgggg gtgatgagct
4681 gcgaggagag caggtttctc aacagctggg acttgccgca cccggtcggg ccgtagatga
4741 ccccgatgac gggttgcagg tggtagttca aggacatgca gctgccgtcg tcccggagga
4801 gggggggcac ctcgttgagc atgtctctga cttggagggt tcccggacg agctcgccga
4861 ggaggcggtc cccgccagc gagagcagct cttgcaggga agcaaagtgt ttcaggggct
4921 tgagcccgtc ggccatgggc atcttggcga gggctctgca gaggagtctg aggcggtccc
4981 agagctcggg gacgtgctct acggcatctc gatccagcag acttctctgt ttcgggggtt
5041 gggacgactg cgactgtagg gcacgagacg atgggcgtcc agcgctgcca gcgtcatgtc
5101 cttccagggt ctcagtgtcc gcgtgagcgt ggtctccgtc acggtgaagg ggtgggcccc
5161 gggctgtgcg cttgcaagggt tgcgcttgag actcatcctg ctgggtgctga aacgggcacg
5221 gtcttcgccc tgcgctcgg cgagatagca gttgaccatg agctcgtagt tgagggcctc
5281 ggcggcggtg cccttggcgc ggagcttgcc cttggaagag cgcccgcagg cgggacagag
5341 gagggattgc agggcgtaga gcttgggtgc gagaagacg gactcggggg cgaaagcatc
5401 cgctccgcag tgggcgcaga cggctctgca ctcgaccagc caggtagact cgggctgctc
5461 ggggtcaaaa accagtttcc cccggttctt tttgatgcgc ttcttacctc gcgtctccat
5521 gagtctgtgt ccgcgctcgg tgacaaacag gctgtctgtg tcccgtaga cggacttgat
5581 gggcctgtcc tgcagggggc tcccgcgggt ctcctcgtag agaaactcgg accactctga
5641 gacgaaggcg cgcgtccacg ccaagacaaa ggaggccacg tgcgaggggt agcggtcgtt
5701 gtccaccagg ggggtccact tttccacggg atgcagacac atgtccccct cctccgcatc
5761 caagaagggtg attggcttgt aggtgtaggc cacgtgaccc ggggtccccg acgggggggt
5821 ataaaagggtg gcgggtctgt gctcgtctc actctcttc gcgtcgtgt ccacgagcgc
5881 cagctgttgg ggtaggtatt ccctttcgag agcgggcatg acctcggcac tcaggttgtc
5941 agtttctaga aacgaggagg atttgatgtt ggcttgccct gccgcaatgc tttttaggag
6001 actttcatcc atctggtcag aaaagactat ttttttattg tcaagcttgg tggcgaagga
6061 gccatagagg gcgttgagga gaagcttggc gatggatctc atggtctgat tttgtcacg
6121 gtcggctcgc tccttggccg cgatgttgag ctggacatac tcgcgcgcga cgcacttcca
6181 ttcggggaag acggtggtgc gctcgtcggg cagatcctg acgcgccagc cgcggttatg
6241 cagggtgacc agatccacgc tgggtggccac ctgcgcgcgc aggggctcgt tggtcagca
6301 gaggcgtccg cccttgcgcg agcagaacgg gggcagcaca tcaagcagat gctcgtcagg
6361 ggggtccgca tcgatggtga agatgcccg acagagttcc ttgtcaaaat aatcgatttt
6421 tgaggatgca tcatccaagg ccacttgcca ctcgcgggcg gccagcgtc gctcgtaggg
6481 gttgaggggc ggaccccagg gcatgggatg cgtcagggcg gaggcgtaca tgccgcagat
6541 gtcgtagaca tagatgggct ccgagaggat gccgatgtag gtgggataac agcggcccc
6601 gcggatgctg gcgcgcacgt agtcatacaa ctcgtgcgag ggggccaaga aggcggggcc
6661 gagattggtg cgctggggct gctcggcgcg gaagacgatc tggcgaaaga tggcatgca
6721 gttggaggag atggtgggccc gttggaagat gttaaagtgg gcatgaggca gacgaaccga
6781 gtcgcggatg aagtgcgcgt aggtgtctt cagcttggcg acgagctcgg cggtgacgag
6841 gacgtccatg gcgcagtagt ccagcgttcc gcggatgatg tcataaccgg cctctccttt
6901 cttctcccat agctcgcggt tgagggcgta ctcctcgtca tccttccagt actcccggag
6961 cgggaatcct cgatcgtccg cacggtaaga gccagcatg tagaaatggt tcacggcctt
7021 gtagggacag cagcccttct ccacggggag ggcgtaagct tgagcggcct tgccgagcga
7081 ggtgtgcgtc agggcgaagg tatccctgac catgactttc aagaactggt acttgaaatc
7141 cgagtcgtcg cagccgccgt gctcccagag ctcgaaatcg gtgcgcttct tcgagagggg
7201 gttaggcaga gcgaaagtga cgtcattgaa gagaatcttg cctgcccgcg gcatgaaat
7261 gcgggtgatg cggaaagggc cgggacgga ggctcgggtt ttgatgacct gggcggcgag

FIG. 16A-2

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7321 gacgatctcg tcgaagccgt tgatgttgtg cccgacgatg tagagttcca tgaatcgcg
7381 gcggccttta atgtgcggca gctttttgag ctctcgtag gtgaggtcct cggggcaatg
7441 cagtccgtgc tgctcgagcg cccactcctg gagatgtggg ttggcttgca tgaatgaagc
7501 ccagagctcg cgggccataa gggctctggag ctcgctcgca aagaggcgga actgctggcc
7561 cacggccatc ttttctgggg tgacgcagta gaaagtaagg gggctccgct cccagcgatc
7621 ccagcgtaag cgcacggcta gatcgcgagc gagggcgacc agctctgggt ccccgagaa
7681 tttcataacc agcataaagg ggacgagctg cttgccgaag gaccccatcc aggtgtaggt
7741 ttctacatcg taggtgacaa agagccgctc cgtgcgagga tgagagccga ttgggaagaa
7801 ctggatttcc tgccaccagt tggacgagtg gctgttgatg tgatgaaagt agaaatccc
7861 ccggcgaacc gagcactcgt gctgatgctt gtaaaagcgt ccgcagtact cgcagcgctg
7921 cacgggctgt acctcatcca cgagatacac agcgcgctcc ttgaggagga acttcaggag
7981 tggcgccctt ggctgggtgg tttcatgttc gcctgcgtgg gactcacctt ggggctcctc
8041 gaggacggag aggtgacga gccgcgcgg gagccaggtc cagatctcgg cgcggcgggg
8101 gcggagagcg aagacgaggg cgcgcgattg ggagctgtcc atggtgtcgc ggagatccag
8161 gtccgggggc agggttctga ggttgacctc gtagaggcgg gtgaggcggt gcttgagatg
8221 cagatggtac ttgatctcca cgggtgagtt ggtggctgtg tccacgcatt gcatgagccc
8281 gtagctgcgc ggggccacga ccgtgcgcgc gtgcgctttt agaagcgggt tgcggacgc
8341 gctcccgcg gcagcgcgcg ttccggcccc gcgggcaggg gcggcagagg cacgtcggcg
8401 tggcgctcgg gcaggtcccg gtgctgcgc ctgagagcgc tggcggtgcg gacgacgcgg
8461 cgggtgacat cctggatctg ccgcctctgc gtgaagacca ccggccccgt gactttgaac
8521 ctgaaagaca gttcaacaga atcaatctcg gcgtcattga cggcgccctg acgcaggatc
8581 tcttgacagt cgcccgagtt gtcctggtag gcgatctcgg acatgaactg ctcgatctcc
8641 tcctcctgga gatcgccgcg gccgcgcgc tccacgggtg cggcgaggtc attggagatg
8701 cgacccatga gctgcgagaa ggcgcccagg ccgctctcat tccagacgcg gctgtagacc
8761 acgtccccgt cggcgctcgc cgcgcgcatg accacctgcg cgaggttgag ctccacgtgc
8821 cgcgtaaga cggcgtagtt gcgcaggcgc tggaagaggt agtttagggg ggtggcgatg
8881 tgctcgggtga cgaagaagta catgatccag cggcgagggg gcatctcgct gatgtcgccg
8941 atggcctcca gcctttccat ggcctcgtag aaatccacag cgaagttgaa aaactgggcg
9001 ttgcgggccg agaccgtgag ctcgctcctc aggagcctga tgagttcggc gatggtggcg
9061 cgcacctcgc gctcgaaatc cccggggggc tctcctctt cctcttctt catgacgacc
9121 tcttcttcta tttcttctc tgggggcggt ggtggtggcg gggccccgac acgacggcga
9181 cgcaccggga gacggtcgac gaagcgctcg atcatctccc cgcgcgggcg acgcatggtt
9241 tcggtgacgg cgcgaccccg ttcgcgagga cgcagcgtga agacgcccgc ggtcatctcc
9301 cggtaatggg gcgggtcccc gttgggcagc gagagggcgc tgacgatgca tcttatcaat
9361 tgcggtgtag gggacgtgag cggtcgcgaga tcgaccggat cggagaatct ttcgaggaaa
9421 gcgtctagcc aatcgagtc gcaaggtaag ctcaaacacg tagcagccct gtggacgctg
9481 ttagaattgc ggttgctgat gatgtaattg aagtaggcgt ttttaaggcg gcggatggtg
9541 gcgaggagga ccaggctcct gggctccgct tgctggatgc gaagccgctc ggccatgccc
9601 caggcctggc cctgacaccg gctcaggttc ttgtagtagt catgcatgag cctctcaatg
9661 tcatcactgg cggaggcgga gtcttccatg cgggtgaccc cgacgcccct gagcggctgc
9721 acgagcgcca ggtcggcgac gacgcgctcg gcgaggatgg cctgttgacac gcgggtgagg
9781 gtgtcctgga agtcgtccat gtcgacgaag cgggtgtagg ccccggtgtt gatggtgtag
9841 gtgcagttgg ccatgagcga ccagttgacg gtctgcaggg cgggttgacac gacctctgag
9901 tacctgagcc gcgagaaggc gcgcgagtcg aagacatagt cgttgacagg gcgcacgagg
9961 tactggtatc caactaggaa gtgcggcggc ggctggcggt agagcgccca gcgctgggtg
10021 gccggcgcg gcggggccag gtcctcgagc atgaggcggt ggtagccgta gaggtagcgg
10081 gacatccagg tgatgccggc ggcggtggtg gaggcgcgcg ggaactcgcg gacgcggtt
10141 cagatgttgc gcagcggcag gaaatagtc atggtcggca cggctctggc ggtgagacgc
10201 gcgcagtcac tgacgctcta gaggcaaaaa cgaaagcggt tgagcgggct cttcctcgt
10261 agcctggcgg aacgcaaacg ggttaggccg cgtgtgtacc ccggttcgag tcccccgaa
10321 tcaggctgga gccgcgacta acgtggtatt ggcactcccg tctcgaccgg agcccgatag
10381 ccgccaggat acggcgagga gccctttttg ccgaccgagg ggagtgcgta gacttgaaag
10441 cggccgaaaa ccccgccggg tagtggtcgc cgcccgtagt ctggagaagc tttgccaggg
10501 ttgagtcgcg gcagaacccg gttcgcggac ggccgcggcg agcgggactt ggtcaccgcc
10561 ccgatttaaa gaccacagc cagccgactt ctccagttac gggagcgagc cccctttttt
10621 ctttttgcca gatgcatccc gtctgcgcgc aaatgcgtcc caccctccct ccggcgacca
10681 ccgcgaccgc ggccgtagca ggcccgggcg ctgtagcccc gccacagcag acagagatgg
10741 acttggaaga gggcgaaggg ctggcgagac tgggggcgcc gtccccggag cgacaccccc
10801 gcgtgcagct gcagaaggac gtgcgcccgg cgtacgtgcc tgccgagaa ctgttcaggg
10861 accgcagcgg ggaggagccc gaggagatgc gcgactgcg ttttcggggc ggcagggagc
10921 tgcgcgaggg cctggaccgc cagcgcgctg tgccgcagca ggatttcgag ccgaacgagc

FIG. 16A-3

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10981	agacggggat	cagccccgcg	cgcgcgacag	tggcggcgcc	caacctggtg	acggcctacg
11041	agcagacggt	gaagcaggag	cgcaacttcc	aaaagagttt	caacaaccat	gtgcgcacgc
11101	taatcgcgcg	cgaggagggt	gccctgggct	tgatgcacct	gtgggacctg	gcggaggcca
11161	tcgtgcagaa	cccggacagc	aagcctctga	cggcgcagct	gttcctggtg	gtgcagcaca
11221	gcagggacaa	cgaggcggtc	agggaggcgc	tgctaaacat	cgccgagccc	gagggccgct
11281	ggctgctgga	gctgatcaac	atcttgacga	gcacgtagt	gcaggagcgc	agcctgagcc
11341	tggccgagaa	ggtggcggtc	atcaactact	cggtgctgag	cctgggcaag	ttttacgcgc
11401	gcaagattta	caagacgccg	tacgtgcccc	tagacaagga	ggtgaagata	gacagctttt
11461	acatgcgcat	ggcgctcaag	gtgctgacgc	tgagcgacga	cctgggctgt	taccgcaacg
11521	accgcatcca	caaggccgtg	agcgcgagcc	ggcggcgcg	gctgagcgac	cgcgagctga
11581	tgctgagtct	gcgcggggcg	ctggtagggg	gcgcggcgcg	cggtgaggag	tcctacttcg
11641	acatgggggc	ggacctgcat	tggcagccga	gcccggcgcg	cttgagggcc	gcctacggtc
11701	cagaggactt	ggatgaggat	gaggaagagg	aggaggatgc	accgctgcg	gggtactgac
11761	gcctccgtga	tgtgttttta	gatgcagcaa	gccccggacc	ccgccataag	ggcggcgctg
11821	caaagccagc	cgtccgggtc	agcatcggac	gactgggagg	ccgcgatgca	acgcatcatg
11881	gccctgacga	cccgaacccc	cgagtccttt	agacaacagc	cgaggcccaa	cagactctcg
11941	gccattctgg	aggcgggtgt	cccctctcgg	accaaccccc	cgacagagaa	ggtgctggcg
12001	atcgtgaacg	cgctggcgga	gaacaaggcc	atccgtcccc	acgaggccgg	gctggtgtac
12061	aacgccctgc	tggagcgctg	gggcccgtac	aacagcacaa	acgtgcagtc	caacctggac
12121	cggctggtga	cggacgtgcg	cgaggccgtg	gcgcagcgcg	agcggttcaa	gaacgagggc
12181	ctgggctcgt	tgggtggcgct	gaacgccttc	ctggcgacgc	agccggcgaa	cgtgccgcgc
12241	gggcaggacg	attacaccaa	ctttatcagc	gcgctgcggc	tgatggtgac	cgagggtccc
12301	cagagcgagg	tgtaccagtc	gggcccagac	tactttttcc	agacgagccg	gcagggtctg
12361	cagacggtga	acctaagcca	ggctttcaag	aatctgcgcg	ggctgtgggg	cgtgcaggcg
12421	cccgtgggcg	accggtcgac	ggtgagcagc	ttgctaaccg	ccaactcgcg	gctgctgctg
12481	ctgctgatcg	cgcccttcac	cgacagcggc	agcgtgaacc	gcaactcgta	cctgggccac
12541	ctgctgacgc	tttaccgcga	ggccataggg	caggcgaggg	tggacgagca	gaccttcag
12601	gagatcacta	gcgtgagccg	cgcgctgggt	cagaacgaca	ccgacagtct	gagagccacc
12661	ctgaacttct	tgctgacaaa	tagacagcag	aagattccgg	cgagtagcgc	gctgtcggcc
12721	gaggaggagc	gcacccctgag	atatgtgcag	cagagcgtag	ggcttttccct	gatgcaggag
12781	ggggccaccc	ccagcgccgc	gctggacatg	accgcgcgca	acatggaacc	tagcatgtac
12841	gccgccaacc	ggcggttcat	caataagctg	atggactacc	tgcaccgcgc	ggctgccatg
12901	aactcggact	actttactaa	tgctatacta	aaccgcgact	ggctcccggc	gccgggggtc
12961	tacacgggcg	agtacgacat	gcccgaaccc	aacgatgggt	tcctgtggga	cgacgtggac
13021	agcgcggtgt	tctccccgac	cttgcaaaaag	cgccaggagg	cggtacgcac	gcccgcgagc
13081	gagggcgcg	tgggtcggag	cccctttcct	agcttaggga	gtttgcatag	cttgccgggc
13141	tcggtgaaca	gcggcagggt	gagccggccg	cgcttgctgg	gcgaggacga	gtacctgaac
13201	gactcgctgc	tgcagccgcc	gcgggtcaag	aacgccatgg	ccaataacgg	gatagagagt
13261	ctggtggaca	aactgaaccg	ctggaagacc	tacgctcagg	accataggga	tgcgcccgcg
13321	ccgcggcgac	agcgccacga	ccggcagcgg	ggcctggtgt	gggacgacga	ggactcggcc
13381	gacgatagca	gcgtgttgga	cttggggcg	agcgggtggg	ccaaccggtt	cgcgcatctg
13441	cagcccagac	tggggcgacg	gatgttttga	atgaaataaa	actcaccaag	gccatagcgt
13501	gcgttctctt	ccttggttaga	gatgaggcgc	gcgggtggtg	cttcctctcc	tcctccctcg
13561	tacgagagcg	tgatggcgca	ggcaaccctg	gaggttccgt	ttgtgcctcc	gcggtatatg
13621	gctcctacgg	agggcagaaa	cagcattcgt	tactcggaac	tggctccgca	gtacgacacc
13681	actcgcggtg	acttggtgga	caacaagtgc	gcggacatcg	cttccctgaa	ctacaaaaac
13741	gaccacagca	acttcctgac	cacggtggtg	cagaacaacg	atttcacccc	cgccgaggcc
13801	agcacgcaga	cgataaattt	tgacgagcgg	tcgcgggtgg	gcggtgattt	gaagaccatt
13861	ctgcacacca	acatgcccac	tgtgaacgag	tacatgttca	ccagcaagtt	taaggcgcg
13921	gtgatggtgg	ctaggaaggt	ggtagatcag	aatgatagga	gcaaggatga	gttaaaatat
13981	gagtgggttg	agtttaccct	gcccgagggc	aacttttccg	agaccatgac	catagacctg
14041	atgaacaacg	ccatcttgga	aaactacttg	caagtggggc	ggcaaaaatg	cgtgctggag
14101	agcgatatcg	gagtcaagtt	tgacagcagg	aatttcaagc	tgggctggga	cccggtaacc
14161	aagctggtga	tgccctgggt	ctacacctac	gaggccttcc	acccggacgt	tgtgctgctg
14221	ccgggctgcg	gggtggactt	caccgagagc	cgccctgagca	acctcctggg	cattcgcaag
14281	aagcaacctt	tccaagaggg	cttcaggatc	atgtatgagg	atctcgaggg	tggtaacatc
14341	cccggccctc	tggatgtcaa	gcaatatatt	gatagtaaaa	agaagcttga	ggaggcaaca
14401	cagaatgcaa	ccagggctgc	tggagatatc	agaggagaca	gtcatattcc	aagagctgtg
14461	gaacaagcgg	ctgaaaagga	tctgggtcatt	gtaccagtaa	cacaagatga	aagtaagaga
14521	agctataatg	tcatagatgg	cacccatgac	accctctacc	gaagttggta	cctgtcctat
14581	acctacgggg	accccagaaa	gggggtgcag	tcgtggacgc	tgctcaccac	cccggacgtc

FIG. 16A-4

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14641 acctgcggcg cggagcaagt ctactggtcg ctgccggacc tcatgcaaga ccccgtcacc
14701 ttccgctcta cccagcaagt cagcaactac cccgtggttg gcgccgagct catgcccttc
14761 cgcgccaaaga gcttttacaa cgacctcgcc gtctactccc agctcatccg cagctacacc
14821 tccctcacc acgtcttcaa ccgcttcccc gacaaccaga tcctctgccg tccgcccgcg
14881 cccaccatca ccacggtcag tgaaaacgtg cctgctctca cagatcacgg gacgctaccg
14941 ctgcgcagca gtatccgcgg agtccagcga gtgaccgtca ctgacgcccg tcgccgcacc
15001 tgtccctacg tctacaaggc cctgggcata gtgcgcgcgc gcgtgctttc cagtgcgacc
15061 ttctaaaaaa tgtctattct catctcgccc agcaataaca cgggctgggg tcttactagg
15121 cccagcacca tgtacggagg agccaagaag cgctcccage agcaccccg cccggtccgc
15181 ggccacttcc gcgctccctg gggcgcttac aagcgcgggc ggacttctac cgcgcgctg
15241 cgcaccaccg tcgacgacgt catcgactcg gtggtcgccg acgcgcgcaa ctataccccc
15301 gccccctcca ccgtggacgc ggtcatcgac agcgtggtgg ccgacgcgcg cgactatgcc
15361 agacgcaaga gccggcgggc acggatcgcc agggccacc ggagtacgcc cgccatgcgc
15421 gccgcccggg ctctgctgcg ccgcgccaga cgcacgggccc gccgggcat gatgcgagcc
15481 gcgcgccgcg ccgcactgc acccccgcga ggcaggactc gcagacgagc ggccgcccgc
15541 gctgccgcg ccatttctag catgaccaga cccaggcgcg gaaacgtgta ctgggtgcgc
15601 gactccgtca cgggctgcg cgtgcccgtg cgcacccgtc ctccctcgcc ctgatctaat
15661 gcttgtgtcc tccccgcga gcgacgatgt caaagcgcaa aatcaaggag gagatgctcc
15721 aggtcgctgc cccggagatt tacggaccac cccaggcgga ccagaaacc cgcaaatca
15781 agcgggttaa aaaaaaggat gaggtggacg agggggcagt agagtttgtg cgcgagttcg
15841 ctccgcggcg gcgcgtaaat tgggaagggc gcagggtgca gcgcgtgttg cggcccggca
15901 cggcggtggt gtttacgccc ggcgagcggt cctcggtcag gagcaagcgt agctatgacg
15961 aggtgtacgg cgacgacgac atcctggacc agggcgcgga gcgggcgggc gagttcgct
16021 acgggaagcg gtcgcgcgaa gaggagctga tctcggtgcc gctggacgag agcaaccaca
16081 cgcctagcct gaagcccgtg accctgcagc aggtgctgcc ccaagcagtg ctgctgccga
16141 gccgcggggt caagcgcgag ggcgagaata tgtaccgcac catgcagatc atggtgcca
16201 agcgcggcg cgtggaagaa gtgctggaca ccgtgaaat ggatgtggag cccgaggtca
16261 aggtgcgccc catcaagcag gtggcgccgg gcctggcggt gcagaccgtg gacattcaga
16321 tccccaccga catggatggt gacaaaaaac cctcgaccag catcgaggtg cagaccgacc
16381 cctggctccc agcctccacc gctgcccgtc ccacttctac cgccgccacg gctaccgagc
16441 ctcccagaag gcgaagatgg ggccctgcca accggctgat gcccactac gtattgcac
16501 cttccattat cccgacgcgc ggctatcgcg gcacccggtg ctacgccagc cgcaggcgcc
16561 cagccagcaa acgcccgcgc cgcaccgcca cccgcgcgcg tctggcccc gcccgcgtgc
16621 gccgcgtaac cacgcgcggg ggccgctcgc tcgttctgce caccgtgcgc taccaccca
16681 gcatccttta atccgtgtgc tgtgatactg ttgcagagag atggctctca cttgccgct
16741 gcgcaccccc gtcccgaatt accgaggaag atcccgccgc aggagaggca tggcaggcag
16801 cggcctcaac cgcgcgcggc ggcgggcat gcgcaggcgc ctgagtggcg gctttctgcc
16861 cgcgctcatc ccataatcg cggcgcccat cggcacgac ccgggcatag ctccgctg
16921 gctgcaggcg tcgcagcgcc gttgatgtgc gaataaagcc tcttttagact ctgacacacc
16981 tggctcctgta tatttttaga atggaagaca tcaattttgc gtccctggct ccgcggcacg
17041 gcacgcggcc gttcatgggc acctggaacg agatcggcac cagccagctg aacggggcg
17101 ccttcaattg gagcagtgct tggagcgggc ttaaaaattt cggctcgacg ctccggacct
17161 atgggaacaa ggccctggaat agtagcacgg ggcagttggt aagggaagag ctcaaagacc
17221 agaacttcca gcagaaggat gtggacggcc tagcctcggg cattaacggg gtggtggaca
17281 tagcaaacca ggccgtgcag cgcgagataa acagccgctt ggacccgcgg ccgcccacgg
17341 tgggtggagat ggaagatgca actcctccgc cgccaaggg cgagaagcgg ccgcggcccc
17401 acgcggagga gacgatectg caggtggacg agccgcctc gtacgaggag gccgtcaagg
17461 ccggcatgcc caccacgcgt atcatcgcg cactggccac tgggtgtaat aaaccgcca
17521 cccttgacct gcctccgcca cccacgccc gtgcgcgcgc tcccgcgcg ggcagctccg gttgtgcagc
17581 cccctcctgt ggcgaccgcc gtgcgcgcgc tcccgcgcg ccgccaggcc cagaactggc
17641 agagcacgct gcacagtatc gtgggctggt gagtgaaaag tctgaagcgc cgccgatgct
17701 attgagagag aggaaagagg aactaaagg gagagcttaa cttgtatgtg ccttaccgcc
17761 agagaacgcg cgaagatggc taccctctcg atgatgccgc agtgggcgta catgcacatc
17821 gccgggcagg acgcctcgga gtacctgagc cggggtctgg tgcagtttgc ccgcgccacc
17881 gacacgtact tcagcctggg caacaagttt aggaaccca cgggtggctc caccacgat
17941 gtgaccacgg accggtccca gcgtctgacg ctgcgcttg tggccgtgga tcgcgaggac
18001 accacgtact cgtacaaggc gcgcttctac ctggccgtgg gcgacaaccg ggtgctagac
18061 atggccagca cttactttga catccgcggc gtcctggacc gcggtcccag cttcaaacc
18121 tactcgggca cggcttacia cagcctggcc cccaaaggcg ccccaactc tagtcagtgg
18181 gaacaagcta aagctaccaa tgccgggtcaa aaggaaactc acacatttgg agtagccgct
18241 atgggcggag aagacattac agtgaaagg cttcaaattg gaactgatga aactaaggaa

FIG. 16A-5

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18301 gatggagagg atgaaatttt tgcagatcaa acattccagc cagaacctca agtgggagaa
18361 cagaactggc aagaaacggt tgttttctat ggaggcagag ctcttaagaa agaaaccaa
18421 atgaagccat gttatggctc ttatgcgaga cccacaaatg aaaagggagg acaggctaaa
18481 ttacacttg atgaaaaagg tcagccaacc aaaattcctg atattacaat ggatttcttt
18541 gatagtccac aagatgatac atcaggtgta actaataagc cagatattgt catgtatgca
18601 gaaaatgtaa atttagaagc tcctgacaca catgtagttt acaaaccagg caaagatgat
18661 tctagttctt ccgctaacct cacacaacag gccatgccta acagaccgaa ctacatcggg
18721 ttcagagaca actttgtggg tcttatgtac tacaatagta ctggcaacat ggggtgtgctg
18781 gctggtcagg cctctcagtt gaatgctgtg gtcgacttgc aagacagaaa caccgagctg
18841 tcttaccagc tattgctaga ttctctgggt gacagaacca gatactttag catgtggaat
18901 tctgcagtgg acagctatga ccccgatgtc aggatcattg agaatcacgg tgtggaagat
18961 gaacttccaa actattgctt cccactgaat ggcagtgggt ctaacagcac atacaaaggt
19021 gttaaagctg gaactggaaa caattgggat gacgatgaaa atgttgcaag acaaaatcag
19081 attggcactg gcaacctggt cgccatggag atcaacctcc aggccaacct atggaagagt
19141 tttctgtact cgaacgtggc cctgtacctg cccgactcct acaagtacac gccggccaac
19201 gtcacgctgc ccaccaacac caacacctac gactacatga acggccgctg ggtagcccc
19261 tgcgtgggtg acgcctacat caacattggc gcccgctggt cgctggaccc catggacaat
19321 gtcaatccct tcaaccacca ccgcaacgcg ggcctgctgt accgctccat gctcctgggc
19381 aacggccgct acgtgccctt ccacatccaa gtgccccaaa agttctttgc catcaagaac
19441 ctgcttctgc tcccgggtt ctacacctac gagtggaaact tccgcaagga cgtcaacatg
19501 atcctgcaga gttccctcgg caacgacctg cgcgtcgacg gcgctcctg ccgcttcgac
19561 agcgtcaacc tctacgccac cttcttcccc atggcgcaac acaccgctc caccctggaa
19621 gccatgctgc gcaacgacac caacgaccag tccttcaacg actacctctc ggccgccaac
19681 atgctctacc ccattccggc caaggccacc aacgtgccca tctccatccc ctgcgcgaac
19741 tgggcccgtc tccgcggtg gaggttcacc cggctcaaga ccaaggaaac tccctcccctc
19801 ggctcgggtt tgcacccta ctttgtctac tggggtcca tcccctacct cgacgggacc
19861 ttctacctca accacacctt caagaaggte tccatcatgt tgcactctc ggtcagctgg
19921 cccggcaacg accggtgct cagccgaac gaggtcgaga tcaagcgag cgttgacggg
19981 gagggctaca acgtggccca atgcaacatg accaaggact ggttctctgt ccagatgctc
20041 tcccactaca acatcggtta ccagggttc cacgtgcccg agggctacaa ggaccgcatg
20101 tactccttct tccgcaactt ccagcccatg agcaggcagg tggtcgatga gatcaactac
20161 aaggactaca aggcggtcac cctacccttc cagcacaaca actcgggctt caccggctac
20221 cttgcgccca ccattgcgca ggggcagccc taccgcca acttccccta cccgctcatc
20281 ggctccaccg cagttccctc cgtcaccag aaaaagttcc tctgcgacag ggtcatgtgg
20341 cgcaccccat tctccagcaa ctttatgtcc atgggcgccc tcaccgacct gggtcagaac
20401 atgctctatg ccaactcggc ccacgcgctc gacatgacct ttgaggtgga ccccatggat
20461 gagcccaccc tctctatct tctcttcgaa gttttcgcag tggtcagagt gcaccagccg
20521 caccgcggcg tcatcgaggc cgtctacctg cgcacgcctt tctccgcccg caacgctacc
20581 acttaagcat gagcggctcc agcgaacaag agctcgcggc catcgtgcgc gacctgggat
20641 gcggggcccta ctttttggga acccagaca agcgttccc tggcttctt gccggcgaca
20701 agctggcctg cgccatcgte aacacggccg gccgcgagac cggaggcgtg cactggctcg
20761 cctttgggtg gaatecgcgc tcgcgcacct gctacatgtt cgaccccttt gggttctcgg
20821 accgcccggc caagcagatt tacagcttcg agtacgaggc catgctgcgc cgaagcgcg
20881 ttgcctcctc gcccgaccgc tgtctcagcc tcgagcagtc caccagacc gtgcaggggc
20941 ccgactccgc cgcctgcgga cttttttgtt gcatgttttt gcatgccttc gtgcactggc
21001 ccgaccgacc catggacgga aaccccacca tgaacttgct gacgggggtg ccaaaccgga
21061 tgctacaatc gccacagggt ctgcccaccc tcaggcgcaa ccaggaggag ctctaccgct
21121 tctcgcgcgc ccaactcccct tactttcgat cccacgcgc cgccatcgaa aacgccaccg
21181 cttttgataa aatgaaacaa ctgctgtgat ctcaataaac agcactttat tttacatgca
21241 ctggagtata tgcaagttat ttaaaagtgc aaggggttct cgcgctcgtc gttgtgcgcc
21301 gcgctgggga gggccacggt gcggtactgg tacttgggaa gccacttgaa ctcggggatc
21361 accagtttgg gcactggggt ctcgggggag gtctcgctcc acatgcgccc gctcatctgc
21421 agggcgccca gcatgtccgg gccggagatc ttgaaatcac aattggggcc ggtgctctgc
21481 gcgcgcgagt tgcggtacac ggggttgag cactggaaca ccattagact ggggtacttc
21541 aactggcaa gcacgctctt gtcgctgac tgatccttgt ccaggctcctc ggcgttgctc
21601 aggcgaacg ggtcatctt gcacagctgg cggcccagga agggcacgct ctgaggcttg
21661 tggttacact cgcagtgcac gggcatcagc atcatccccg cgcgcgctg catattcggg
21721 tagaggcct tgacgaaggc cgtgatctgc ttgaaagctt gctgggcctt agccccctcg
21781 ctgaaaaaca ggccgcagct cttcccgtta aactggttat tcccgacccc ggcacatgc
21841 acgcagcagc gcgctcatg gctggctcag tgcaccagc tacgtcccca gcggttctgg
21901 gtcaccttgg ccttgctggg ctgctccttc aacgcgcgct gccggttctc gctggtcaca

FIG. 16A-6

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21961 tccatctcca ccacgtggtc cttgtggatc atcacggtcc catgcagaca cttgagctga
22021 ccctcgacat cgcagcagcc atgatccac agggcgcagc cgggtgcactc ccagttctta
22081 tgcgcgatcc cgctgtggct gaagatgtaa ccttgcaaca ggcgacccat gacgggtgcta
22141 aatgctttct ggggtggtgaa ggtcagttgc agaccgcggg cctcctcgtt catccaggtc
22201 tggcacatct tttggaagat ctccggtctgc tcgggcatga gcttgtaagc atcgcgcagg
22261 ccgctgtcga cgcggtagcg ttccatcagc acgttcatgg tatccatgcc cttctcccag
22321 gacgagacca gaggcagact caggggggtg cgcacgttca ggacaccggg ggtcgcaggc
22381 tcgacgatgc gttttccgtc cttgccttcc ttcaacagaa ccggaggctg gctgaatccc
22441 actcccacga ttacggcatc ttctggggc atctcttctg cggggtctac cttgggtcaca
22501 tgcttggtct ttctggcttg cttctttttt ggagggtgtt ccacggggac cacgtcctcc
22561 tcggaagacc cggagccac ccgctgatac tttcggcgct tgggtggcag aggaggtggt
22621 ggcggcgagg ggctcctctc ctgctccggc ggatagcgcg ccgaccctg gccccggggc
22681 ggagtggcct ctccgtccat gaaccggcgc acgtcctgac tgccgcccgc cattgtttcc
22741 taggggaaga tggaggagca gccgcgttag caggagcagg aggaggactt aaccacccac
22801 gagcaaccca aaatcgagca ggacctgggc ttccaagagc cggctcgtct agaaccccca
22861 caggatgaac aggagcacga gcaagacgca ggccaggagg agaccgacgc tgggctccag
22921 catggctacc tgggaggaga ggaggatgtg ctgctaaaac acttgacgcg ccaatccatc
22981 atcctccggg acgcccctggc cgaccggagc gaaacccctc tcagcgtcga ggagctgtgt
23041 cgggcctacg agctcaacct cttctcgcgc cgcgtgcccc ccaaacgcca gcccaacggc
23101 acctgcgagc ccaacccgcg tctcaacttc tatcccgtct ttgcgggtccc cgaggcccta
23161 gccacctatc acatcttttt caagaaccaa aagatccccg tctcctgccc cgccaaccgc
23221 acccgcgccc acgcgtcctt cgtctgtggg cccggcgcgc gcatacctga tatcgcttcc
23281 ctggaagagg tgcccaagat cttcgaaggg ctccggtcgg acgagacgcg cgcggcaaac
23341 gctctgaaag aaacagcaga ggaagagggt cacactagcg ccctggtaga gttggaaggc
23401 gacaacgcca ggctggccgt gctcaagcgc agcgtcgagc tcaccactt cgcctacccc
23461 gccgtcaacc tcccgcccaa ggtcatgcgt cgcctcatgg atcagctcat catgcccac
23521 atcgaggccc tcgatgaaag tcaggagcag cgcgccgagg acgcccggcc cgtgggtcagc
23581 gacgagcagc tcgcgcgttg gctcgggacc cgcgaccccc aggccttgga acagcggcgc
23641 aagctcatgc tggccgtggt cctgggtcacc ctccagctcg aatgcagcg ccgcttcttc
23701 agcgaccccg agaccctgcg taaggctcag gagaccctgc actacacttt caggcacggt
23761 ttcgtcaggc aggcctgcaa gatctccaac gtggagctga ccaacctggt ctcatgcctg
23821 gggatcctgc acgagaaccg cctgggacag accgtgctcc actctactct gaagggcgag
23881 gcgcgtcggg actatgtccg cgactgtgta tttctcttta tctgccacac ctggcaagca
23941 gccatgggcg tgtggcagca gtgtctcgag gacgaaaatc tgaaggagct ggacaagctt
24001 cttgctagaa accttaaaaa gctgtggacg ggcttcgacg agcgacccgt cgcctcggac
24061 ctggccgaga tcgttttttc agaacgcctg aggcagacgc tgaaaggcgg gctgcccagc
24121 ttcattgagc agagcatggt gcaaaactac cgcactttca ttctcgagcg atctgggatg
24181 ctacccgcca cctgcaacgc attcccctcc gactttgtcc cgctgagcta ccgcgagtgt
24241 cccccgccgc tgtggagcca ctgctatctc ttgcagctgg ccaactacat cgcctaccac
24301 tcggacgtga tcgaggacgt gagcggcgag gggcttctcg agtgccactg ccgctgcaac
24361 ctgtgctccc cgcaccgctc cctgggtctgc aacccccagc ttctgagcga gaccaggtc
24421 atcggtacct tcgagctgca aggtcccgag gagtccaccg ctccgctgaa actcacgccg
24481 ggggtgtgga cttccgcgta cctgcgcaaa tttgtacccg aggactacca cgcccatgaa
24541 ataaagtctt tcgaggacca atcgcgcccc cagcacgcgg atctcacggc ctgcgtcatc
24601 acccagggcg cgatcctcgc ccaattgcac gccatccaaa aatcccgcca agagtttctt
24661 ctaaaaaagg gtagaggggt ctacctggac cccagacggg gcgaggtgct caaccggggt
24721 ctccccccagc atgccgagga agaagcagga gccgctagtg gagcagatgg aagaagaatg
24781 ggacagccag gcagaggagg acgaatggga ggaggagaca gaggaggaag aattggaaga
24841 ggtggaagag gagcaggaaa cagagcagcc cgtcgcgcga ccatccgcgc cggcagcccc
24901 gccggtcacg gatacaacct ccacagctcc ggccaagcct cctcgtagat gggatcgagt
24961 gaaggggtgac ggtaagcacg agcggcaggg ctaccgatca tggagggtcc acaaagcgc
25021 gatcatcgcc tgcttgcaag actgcggggg gaacatcgct ttcccccgc gctacctgct
25081 cttccaccgc ggggtgaaca tcccccgcaa cgtgttgcat tactaccgtc accttcacag
25141 ctaagaaaaa gcaagtaaga ggagtcgccc gaggaggcct gaggatcgcg gcgaacgagc
25201 cctcgaccac cagggagctg aggaaccgga tcttccccac tctttatgcc atttttcagc
25261 agagtgcagg tcagcagcaa gaactgaaag taaaaaaccc gtctctgcgc tcgctcacc
25321 gcagttgctt gtaccacaaa aacgaagatc agctgcagcg cactctcgaa gacgccgagg
25381 ctctgttcca caagtactgc gcgctcactc tttaaagacta aggcgcgccc acccgaaaa
25441 aaggcgggaa ttacctcatc gccaccatga gcaaggagat tcccacccct tacatgtgga
25501 gctatcagcc ccagatgggc ctggccgcgg gcgcctccca ggactactcc acccgcatga
25561 actggctcag tgccggcccc tcgatgatct cacgggtcaa cgggggtccgt aaccatcgaa

FIG. 16A-7

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25621 accagatatt gttggagcag gcgggcgtca cctccacgcc cagggcaaag ctcaacccgc
25681 gtaattggcc ctccaccctg gtgtatcagg aaatccccgg gccgactacc gtactacttc
25741 cgcgtgacgc actggccgaa gtccgcatga ctaactcagg tgtccagctg gccggcggcg
25801 cttcccgggtg cccgctccgc ccacaatcgg gtataaaaac cctgggtgatc cgaggcagag
25861 gcacacagct caacgacgag ttgggtgagct cttcgatcgg tctgcgaccg gacggagtgt
25921 tccaactagc cggagccggg agatcgtcct tcaactccaa ccaggcctac ctgaccttgc
25981 agagcagctc ttcggagcct cgctccggag gcatcggaac cctccagttc gtggaggagt
26041 ttgtgccctc ggtctacttc aacccttct cgggatcgcc aggcctctac ccggacgagt
26101 ttataccgaa cttcgacgca gtgagagaag cgggtggacgg ctacgactga atgtcccattg
26161 gtgactcggc tgagctcgct cggttgaggg atctggacca ctgccgccgc ctgctgctgt
26221 tcgcccggga gagctgcgga ctcactact ttgagtttcc cgaggagcac cccaacggcc
26281 ctgcacacgg agtgccggatc accgtagagg gcaccaccga gtctcacctg gtcaggttct
26341 tcacccagca acccttcctg gtcgagcggg accggggagc taccacctac accgtctact
26401 gcatctgtcc taccgccgag ttgcatgaga atttttgctg tactctttgt ggtgagttta
26461 ataaaagctg aactaagaac cttctttgga atccctgtc atcatcaaat caacaagacc
26521 atcaacttca cctttgagga acaggtgaac tttacctgca agccacacaa gaagtacatc
26581 atctggtttt atcacaacac tactctagca gtagccaaca cctgctcgaa cgacgggtgt
26641 ctctactcta acaatctcac cagtggacta accttctcag ttaaaagggc aaagctaatt
26701 cttcatcgcc ctattgtaga aggaacttac cagtgtcaga gcggacctg cttccacagt
26761 ttacttttgg tgaacgttac cggcagcagc acagccgctc cagaaacatc taaccttctt
26821 tctgatacta acaaacctcg tgcggaggt gagctttggg ttccatctct aacagagggg
26881 gggagtctta ttgaagtggg tgggtatttg attttagggg tgggtcattg tgggtgcata
26941 gcagtgtgt atcaacttcc ttgctgggtc gaaatcaggg tatttatctg ctgggtcaga
27001 cattgtgggg aggaacctg aaggggctct tgctgattat cctttccctg gtgggggggtg
27061 tgctgtcatg ccacgaacag ccacgatgta acattaccac aggcaatgag aggaacgact
27121 gctctgtagt tatcaaatgc gagcaccatt gtctctcaa catcacattc aagaatpaga
27181 ccatgggaaa tgtatgggtg ggattctggc aaccaggaga tgagcagaac tacacgggtca
27241 ctgtccatgg tagcgatggc aatcacactt tcggtttcaa attcattttt gaagtcatgt
27301 gtgatatac actacatgtg gctagacttc atggcttgtg gccccctacc aaggagaaca
27361 tgggtgggttt ttctttggct tttgtgatca tggcctgctt gatgtcaggt ctgctggtag
27421 gggctctagt gtggtttctg aaacgcaagc ccaggtagcg aaatgaggag aaggaaaaat
27481 tgctataaat tctttttctc ttgcacaaac catgaataca gtgttccgta tctgtctgt
27541 ctctcttctt gtagctttcg gtcaggcagg aattcatatt attaatgcta catggtggga
27601 taatataact ttagtgggac cctcagatac tccagttacc tggatgatg gcaagggatt
27661 gcaattttgt gacggaagta cagttaagaa tccgcagatc agacatactt gtaatgatca
27721 aaacttaact ctgattcatg ttaacaaaac ccatgaaaga acatacatgg gttacagaca
27781 tgacagtaag ggaaaagtag actataaggt tacagtcatt ccacctctc ctgctactgt
27841 aaagccacaa ccagatccag aaaatgtctt tgtttatatg ggaaataatg taactttagt
27901 tggacctcca ggaattccag ttagttggta ttatcataat ggcacacagt tctgcgatgg
27961 agataaaaatt attcatccag aattcaacca cacctgtgat aaacaaaacc ttactctgt
28021 gtttgtaaac ttacacatg atggaggcta tcttggattc aattacaaag gtactcagag
28081 aattcagtat gaggttatag ttttagatcg atttccaat tctggtcaga tgaaaattga
28141 agaacaaagt gaggaacag aacagaaaca tactgagcat aataaggctg gacaaaagca
28201 gggatatagat acaaatcaaa agaaagctaa taacagacaa aagccatctc aaaggccatc
28261 aagaagacgg ccgacaaaca ctctgagac aaaacaactt acagtgtcta ttgggtctaa
28321 cttaacttta gttgggtccag atggaaaagt cacttgggtat gatggtgatt taaaagacc
28381 atgtgaagaa caaaactata ggcttccaca tcagtgtagt gctcagaact taactttaat
28441 taatgtaact aaatctcatg agggaaacta ctatggcact aatgacaaag acgaaagcaa
28501 aagatacaga gtgaaagtga acactacaaa ttctcaagct gtaaaaatta accatatac
28561 cagacctact actcctgatc agaaacacag atttgaatta caaattgaaa ataattgcaa
28621 tgatgaagaa tcaaaaattc catctactac tgtggcaatc gtgggtgggag tgattgcggg
28681 cttcataact ataattcattg tcattctgtg ctacatctgc tgccgcaagc gtcccagggc
28741 atacaatcat atggtagacc cactactcag cttctcttac tgagactcag tcactttcat
28801 ttcagaacca tgaaggcttt cacagcttgc gttctgttta acataatcac acttagtgta
28861 gctgcaaattg gtttttaaca tgttaattgt accagattaa gtaatgtaac actgacagga
28921 gctggaatta atactacatg gacagggtat tttaatgagg gtccaaaagg aaaaaatggg
28981 tggatgaata tttgcacatg gggcgatcct agatatgtgt gccatggaaa tagcagtact
29041 attactaatc ttacagttgt ggcacttcta aatttaacca ctaacagaag atttaaagca
29101 gaaagtttta ctagtacga tggttatgaa actaccagtg caaaatttta tgaaattaaa
29161 attattgagc ttccaacaac tagagcacc accacagtta ggacaacaca gcctaccact
29221 gtgcccacta cacatccaac caccacagtc agtacaacta ttgagaccac tactcatact

FIG. 16A-8

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29281 acacagctag acacaacagt gcagaatact actttattga ttgggttttt actgagagga
29341 aatgaaagta ctactgaaca gacagaggct acctcaagtg ccttcagcag cactgcaaact
29401 ttaacttcgc ttgcttggaac taatgaaacc ggagtatcat tgatgaatcg acagccttac
29461 tcagggtttg atattcaaat tacttttctg gttgtctgtg ggatctttat tcttgcggtt
29521 cttctgtact ttgtctgctg caaagccaga gagaaatcta ggcggcccat atacaggcca
29581 gtaatcgggg aacctcagcc tctccaagtg gatggaggct taaggaatct tctcttctct
29641 tttacagtat ggtgatcagc catgattcct aggttcttcc tatttaacat cctgttctgt
29701 ctcttcaaca tctgtgctgc cttcgcggcc gtctcgcacg cctcgcccgga ctgtctaggg
29761 cctttcccaa catacctcct ctttgccctg ctaacctgca cctgcgtctg cagcattgtc
29821 tgcgtggtca tcacctttct gcagctcatc gactgggtgt ggcgcgcgcta caattatctc
29881 caccacagtc ccgaatacag ggacgagaac gtagccagaa tcttaaggct catctgacca
29941 tgcagcctct gctcatgctg atatecctcc tatecctgct ccttgccact tctgtgatt
30001 actctaaatg caaatcgcg gacatatgga atttcttaga ttgctatcag gagaaaattg
30061 atatgccctc ctattacttg gtgattgttg gggtagtcat ggtctgctca tgcactttct
30121 ttgccattat gatctacccc tgttttaate ttggctggaa ctctgttgag gcattcacat
30181 acacactaga aaacagttca ctagecctca cgccaccacc cacaccgctt ccccgagaa
30241 atcagttccc tatgattcag tacttagaag agccccctcc ccggccccct tccactgtta
30301 gctactttca cataaccggc ggcgatgact gaccacctgg acctcgagat ggacggccag
30361 gcctccgagc agcgcaccc gcaactgcgc gtccgacagc agcaggagcg ggccgccaag
30421 gagctcctcg atgccatcaa catccaccag tgcaagaagg gcactcttct cctgggtcaag
30481 caggcaaaga tcacctacga gctcgtgtcc ggccgcaagc agcatcgcct cgcctatgag
30541 ctaccccagc agaagcaaaa gttcacctgc atgggtggcg tcaaccccat agtcatcacc
30601 cagcagtcgg gcgagaccaa cggctgcac cactgctcct gcgaaagccc cgagtgcac
30661 tactccctcc tcaagaccct ttgcggactc cgcgacctcc tcccatgaa ctgatgttga
30721 ttaaaagccc aaaaaccaat caaaccttc ccaattact cataagaata aatcattgga
30781 actaatcatt caataaagat cacttacttg aaatctgaaa gtatgtctct ggtgtagttg
30841 ttcagcagca cctcggaacc ctctcccag ctctggtact ccagtccccg gcgggcccgc
30901 aacttcctcc acaccttgaa agggatgtca aattcctggt ccacaatttt cattgtcttc
30961 cctcagatga caaagaggct ccgggtggaa gatgacttca acccgtcta cccctatggc
31021 tacgcgcgga atcagaatat ccccttcctt actccccct ttgtttcttc cgatggattc
31081 caaaacttcc cacctggggt cctgtcactc aaactggctg acccaatcgc catcactaat
31141 ggggatgttt cactcaagggt gggagggggt cttactgttg aaaaagatag tggaaatcta
31201 aagggtgaacc ctaaggctcc cttgcaagtt acaactgata aacagttgga aattgactg
31261 gcttatccat ttgaagtcag taatggcaag cttggcataa aagcaggtca tggattgaaa
31321 gtcattgaca aaattgctgg tttggaagggt ttggcaggta cgctttagt tttgactgga
31381 aaaggaatag gtactgaaaa tcttgaaaac agtgatgggt caagtagagg agttggtata
31441 aacgtaagac ttgctaaaga tggaggtctg tcttttgata aaaaggggtga tttagttgct
31501 tggataaaac atgatgacag acgcactcta tggacaactc ccgacctatc cccaaattgt
31561 acaatcgatc aggaagggga ttcaaaagctc acttttagtat taacaaaatg tggcagtcaa
31621 attttggtcta atgtctcttt acttgttgta aaaggaaaat ttagtaacat aaacaataat
31681 actaatccaa ctgataaaaa aatcacagta aagctacttt ttaatgaaaa gggagtatta
31741 atggacagtt cgacacttaa gaaagaatat tggaaactaca gaaatgataa ttctactgta
31801 tctcaggcct atgataatgc agttcctttt atgccaacaa taaaagctta tcctaaacct
31861 accacagaca cttcggctaa accagaagat aaaaaaagtg ctgctaaaag atacattgtg
31921 agcaatgtct atattggagg cttgccagat aaaactgttg ttataactat taagtttaat
31981 gcagaaactg aatgtgctta ttcgattacc tttgaattca catgggcaaa aacctttgaa
32041 gatgtgcagt ttgattcctc ctcttttacc ttttctata ttgccaaga aatgaggac
32101 gaagacaaat aaaatgtttt aaaatgaatt catgtatctt tattgatttt tacaccagca
32161 cgggtagtca gtctcccacc accagcccat ttcacagtgt aaacgattct ctcagcacgg
32221 gtggccttaa atagggaat gttctgatta gtgcgggaac tggacttggg gtctataatc
32281 cacacagttt cctggcgagc caaacggggg tcggtgattg agatgaagcc gtctctgaa
32341 aagtcattca agcgggcctc acagtccaag gtcacagtct ggtgaaacga gaagaacgca
32401 cagattcata ctcgaaaac aggatgggtc tgtgcctctc catcagcgcc ctcaacagtc
32461 tctgccgcg gggctcgggt agcatcagtc agatgggac tccgtgtgag cccgcaccc
32521 ctatgatccc cacagccttc agtaagtgc agcacataat caccatgta ttcagcagcc
32581 tgatctcgct catgttctca ccaaaactca tgttggggat gatggaacc acgtgaccat
32641 cataattcag ggtgctccag atcagatgcc tgccctcat gaacacactg cccatataca
32701 cgtaccagat gcggcagtat ctgttcacaa tctgacggta ccagggaag cgctggttga
32761 tgatctcttt gggcatgtct ctcctgaacc acacggccag cagggtgctt cccgcccagc
32821 acatgcaccc gtaaatgact ctcctgaacc aatgcaggat ccagcgctcg taccgctca
32881 actgcaggga gcccggggat gaacagtggc aatgcaggat ccagcgctcg taccgctca

FIG. 16A-9

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32941 ccatctgagc tctcaccaag tccagggtag cggggcacag gcacactgac atacatcttt
33001 ttaaaatttt tatttcctct ggagtcaaga tcatatccca ggggactgga aactcttgga
33061 gcagggtaaa gccagcagca catggtaatc cacggacaga acttacatta tgataatctg
33121 catgatcaca atcaggcaac aggggatgtt gttcagtcag tgaagccctg gtttcctcat
33181 cagatcgtgg taaacgggcc ctgcgatatg gatgatggcg gagcgagctg gattgaatct
33241 cggtttgcat tgtagtggat tctcttgctg accttgctcg acttctgcca gcagaaatgg
33301 gcccttgaac agcagatacc cctcctgcgg ccgtcctttc gctgctgccg ctcagtcac
33361 caactgaagt acatccattc tcgaagattc tggagaagtt cctctgcatc tgatgaaaca
33421 aaaaacccgt ccatgcgaat tcccctcatc acatcagcca ggactctgta ggccatcccc
33481 atccagttaa tgctgccttg tctatcattc agagggggcg gtggcaggat tggagaacc
33541 atttttattc caaacggctc cgaaggacga taaagtgcaa gtcacgcagg tgacagcgtt
33601 cccctccgct gtgctgggtg aaacagacag ccaggtcaaa acccactcta ttttcaaggt
33661 gctcgaccgt ggcttcgagc agtggctcta cgcgtacatc cagcataaga atcacattaa
33721 aggctggccc tccatcgatt tcatcaatca tcaggttaca ttcctgcacc atccccaggt
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33841 acatgtggaa aagctccac agtgccccct ccactttcat aatcaggcag accttcataa
33901 tagaaacaga tcctgctgct ccaccacctg cagcgtgttc aaaacaacaa gattcaataa
33961 ggttctgccc tccgccctga gctcgcgcct caatgtcagc tgcaaaaaat cacttaagtc
34021 ctggggccact acagctgaca attcagagcc agggctaagc gtgggactgg caagcgtaag
34081 ggaaaacttt aatgctccaa agctagcacc caaaaactgc atgctggaat aagctctctt
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34201 cathttgcgt atagaaaagt cctgtaaata agtcactagg accccaggga ccacaatgtg
34261 gtagcttaca ccgcgtcgt gaagcatggt tagtagagat gagagtctga aaaacagaaa
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34501 tggcatggta gtcattcaag gccataaatc tgccctgata tccagtagga accagcacac
34561 tcacttttag gtgaagcaat accaccccat gcggagggaat gtggaaagat tcagggcaaa
34621 aaaaattata tctattgcta gtcccttcct ggacgggagc aatccctcca ggactatcta
34681 tgaaagcata cagagattca gccatagctc agcccgtta ccagtagaca gagagcacag
34741 cagtacaagc gccaacagca gcgactgact acccactgac ccagctccct atttaaaggc
34801 gccttacact gacgtaatga ccaaaggctt aaaaaccccg ccaaaaaaaa acacacacgc
34861 cctgggtgtt ttttgcgaaa acacttccgc gttctcactt cctcgtattg atttcgtgac
34921 ttaacttccg ggttcccacg ttacgtcact tctgccctta catgtaactc agtcgtaggg
34981 cgccatcttg cccacgtcca aaatggcttc catgtccagc cacgcctccg cggcgaccgt
35041 tagccgtgcg tcgtgacgtc atttgcatca tcttctctcg tccaatcagc gctggccccg
35101 ccctaaattc aaaagctcat ttgcatgtta acttttgttt actttgtggg gtatattatt
35161 gatgatc

SEQ ID NO: 5

FIG. 16A-10

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Grp	Vaccine at Wk 0, Wk 4	Monkey ID	Pre		Wk 4		Wk 8		Wk 12	
			Mock	Gag	Mock	Gag	Mock	Gag	Mock	Gag
1	Ad24ΔE1gagΔOrf6Ad5Orf6 10 ¹¹ vp	00C072	3	4	4	381	3	150	3	68
		00C178	3	3	1	559	1	743	0	635
		00C222	0	3	1	369	1	753	0	670
		00D011	1	9	9	211	4	273	0	520
		00D023	0	6	0	295	1	459	1	368
		00D031	15	5	10	103	1	101	1	40
2	Ad24ΔE1gagΔOrf6Ad5Orf6 10 ¹⁰ vp	99C168	4	6	0	118	5	241	3	209
		99C170	10	5	5	241	3	141	3	103
		99C173	1	3	0	23	0	14	0	21
3	Ad24ΔE1gagΔE4Ad5Orf6 10 ¹⁰ vp	99C154	0	3	0	93	0	60	1	53
		99C158	1	0	1	141	0	101	1	120
		99C177	0	0	0	45	0	39	0	79
4	MRKAd5-HIVgag 10 ¹¹ vp	00C018	1	5	13	1025	0	824	3	753
		00C034	0	4	5	219	5	404	0	491
		00C058	4	4	3	1086	0	440	0	439
5	MRKAd5-HIVgag 10 ¹⁰ vp	99C218	0	3	5	2500	0	1580	10	1655
		99C227	6	1	4	529	5	365	5	1004
		99D185	ND	ND	0	425	0	310	0	271

FIG. 17

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Vaccine at Wk 0, Wk 4	Monkey ID	Gag-Specific (Wk 12)	
		%CD4	%CD8
Ad24ΔE1gagΔOrf6Ad5Orf6 10 ¹¹ vp	00C072	0.02	0.02
	00C178	0.05	0.38
	00C222	0.02	0.40
	00D011	0.02	0.27
	00D023	0.01	0.11
	00D031	0.01	0.01
MRKAd5-HIVgag 10 ¹¹ vp	00C018	0.05	0.41
	00C034	0.06	0.18
	00C058	0.02	0.28

FIG. 18

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Grp	Vaccine at Wk 0, Wk 4	Monkey ID	Wk 4	WK 8
1	Ad24ΔE1gagΔOrf6Ad5Orf6 10 ¹¹ vp	00C072	<10	77
		00C178	<10	26
		00C222	<10	423
		00D011	<10	98
		00D023	<10	<10
		00D031	<10	<10
2	Ad24ΔE1gagΔOrf6Ad5Orf6 10 ¹⁰ vp	99C168	<10	<10
		99C170	<10	<10
		99C173	<10	<10
3	Ad24ΔE1gagΔE4Ad5Orf6 10 ¹⁰ vp	99C154	<10	<10
		99C158	<10	<10
		99C177	<10	<10
4	MRKAd5-HIVgag 10 ¹¹ vp	00C018	34	1017
		00C034	14	423
		00C058	46	934
5	MRKAd5-HIVgag 10 ¹⁰ vp	99C218	20	99
		99C227	40	767
		99D185	17	342

FIG. 19

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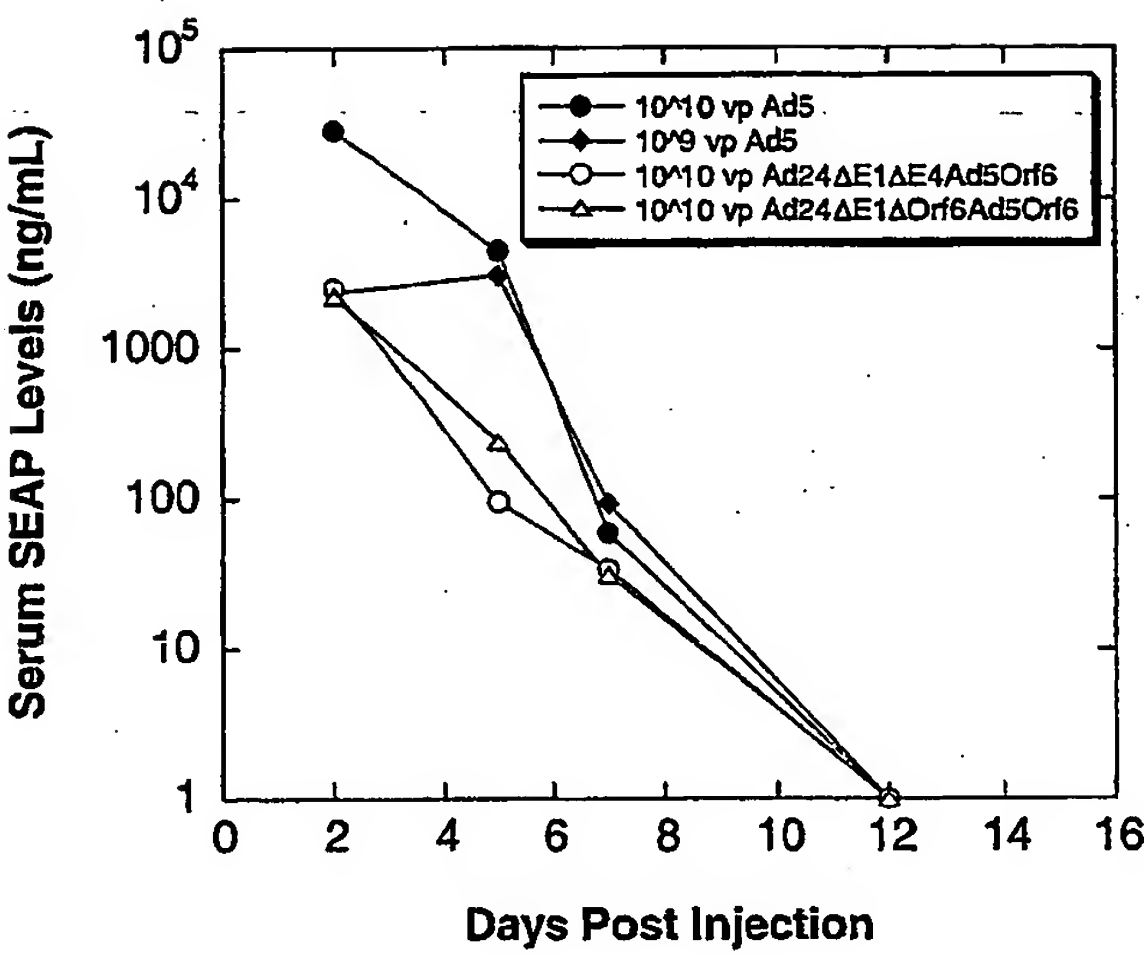


FIG. 20

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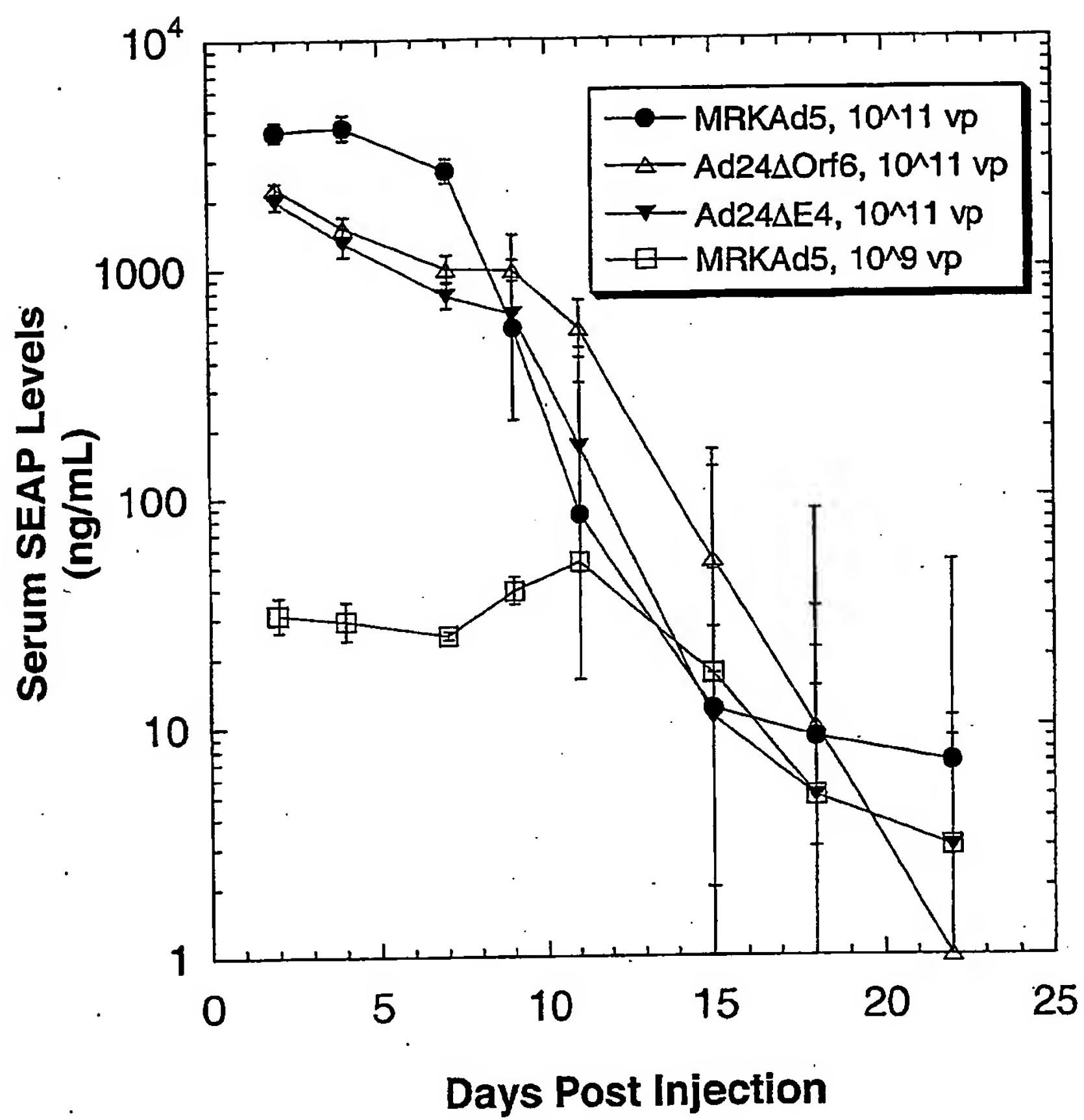


FIG. 21

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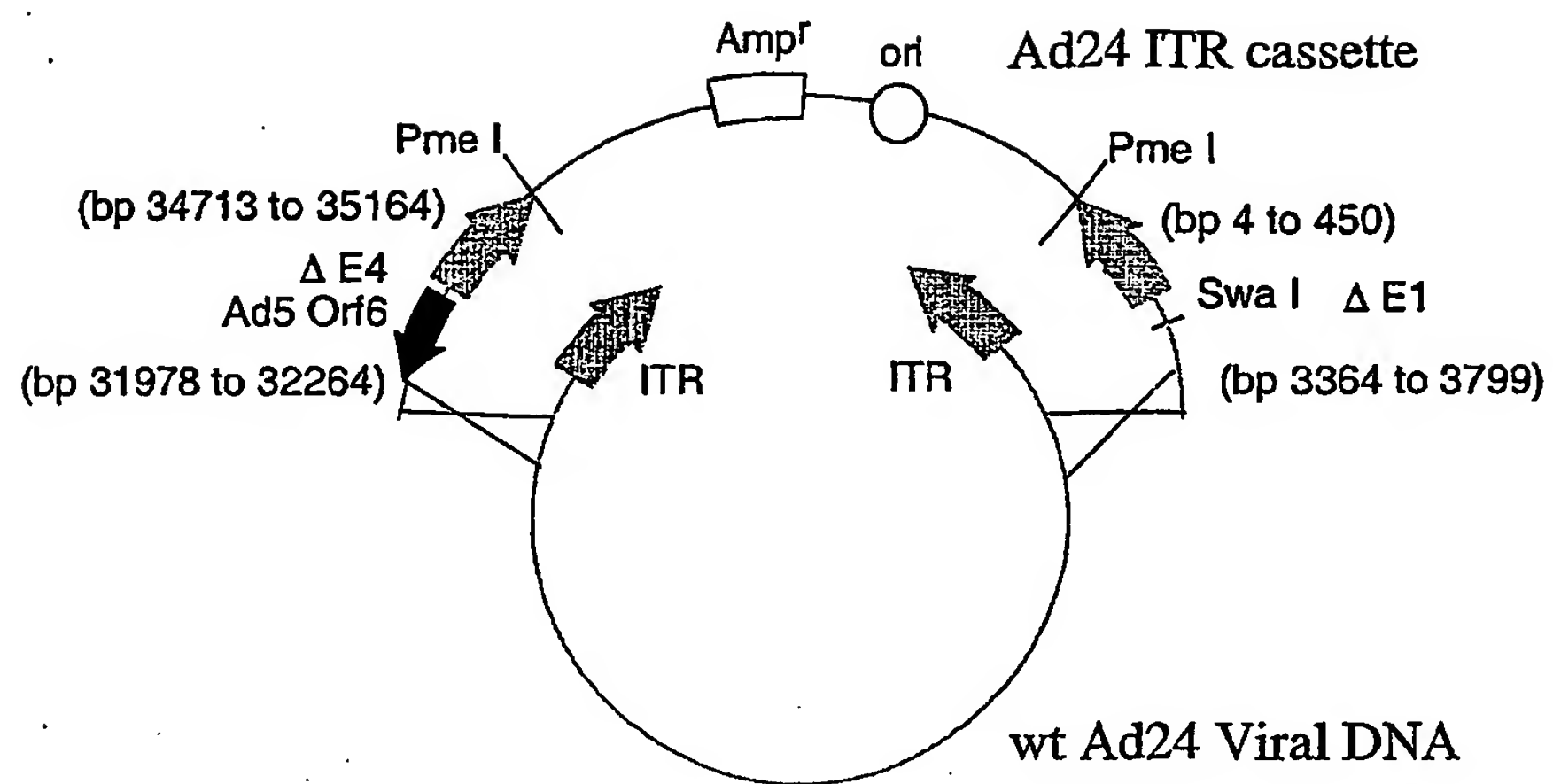


FIG. 22

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Animal	Prime (Wk 0, 4, 26)	Boost (Wk 56)	Pre		Prime ^b		Pre-Boost ^a		Post-Boost ^d	
			Mock ^a	Gag ^a	Mock	Gag	Mock	Gag	Mock	Gag
Monkey 1	10 ⁸ vp MRKAd5-gag	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	18	16	1	244	3	74	3	1235
Monkey 2	10 ⁷ vp MRKAd5-gag	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	10	9	4	83	0	18	0	858
Monkey 3	10 ⁸ vp MRKAd5-gag	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	1	1	0	219	9	69	0	703
Monkey 4	10 ⁷ vp MRKAd5-gag	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	1	1	3	59	1	20	0	419
Monkey 5	none	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	3	4	ND ^c	ND	ND	ND	4	558
Monkey 6	none	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	0	3	ND	ND	ND	ND	1	295
Monkey 7	none	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	1	9	ND	ND	ND	ND	9	103
Monkey 8	none	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	3	3	ND	ND	ND	ND	1	381
Monkey 9	none	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	0	6	ND	ND	ND	ND	0	369
Monkey 10	none	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	15	5	ND	ND	ND	ND	10	211

FIG. 23

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Animal	Prime (Wk 0, 4, 26)	Boost (Wk 56)	Gag-Specific T cells (Wk 60)	
			%CD4	%CD8
Monkey 1	10 ⁸ vp MRKAd5-gag	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	0.06	0.37
Monkey 2	10 ⁷ vp MRKAd5-gag	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	0.01	0.56
Monkey 3	10 ⁹ vp MRKAd6-gag	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	0.07	0.06
Monkey 4	10 ⁷ vp MRKAd6-gag	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	0.04	0.20

FIG. 24

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Animal	Prime (Wk 0, 4)	Boost (Wk 24)	Pre		Prime ^b		Pre-Boost ^c		Post-Boost ^d	
			Mock ^a	Gag ^a	Mock	Gag	Mock	Gag	Mock	Gag
Monkey 11	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	10 ⁷ vp MRKAd5-gag	3	4	3	150	4	28	0	188
Monkey 12	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	10 ⁷ vp MRKAd5-gag	0	3	1	753	4	554	0	1029
Monkey 13	10 ¹¹ vp Ad24ΔE1gagΔOrf6Ad5Orf6	10 ⁷ vp MRKAd5-gag	1	9	4	273	0	370	0	1520
Monkey 14	none	10 ⁷ vp MRKAd5-gag	0	0	ND ^e	ND	ND	ND	4	94
Monkey 15	none	10 ⁷ vp MRKAd5-gag	0	0	ND	ND	ND	ND	1	168
Monkey 16	none	10 ⁷ vp MRKAd5-gag	8	3	ND	ND	ND	ND	8	149

FIG. 25

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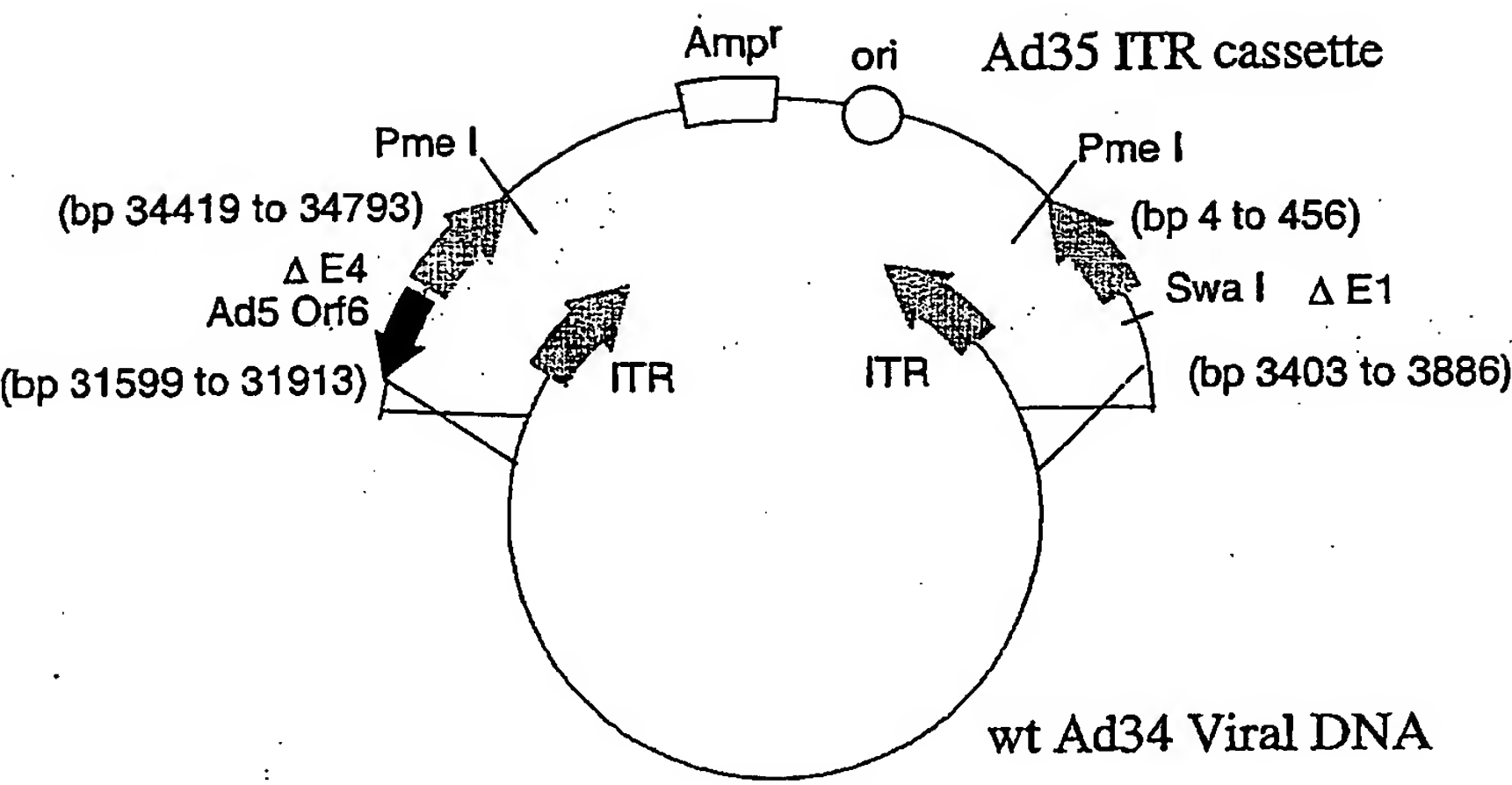


FIG. 26

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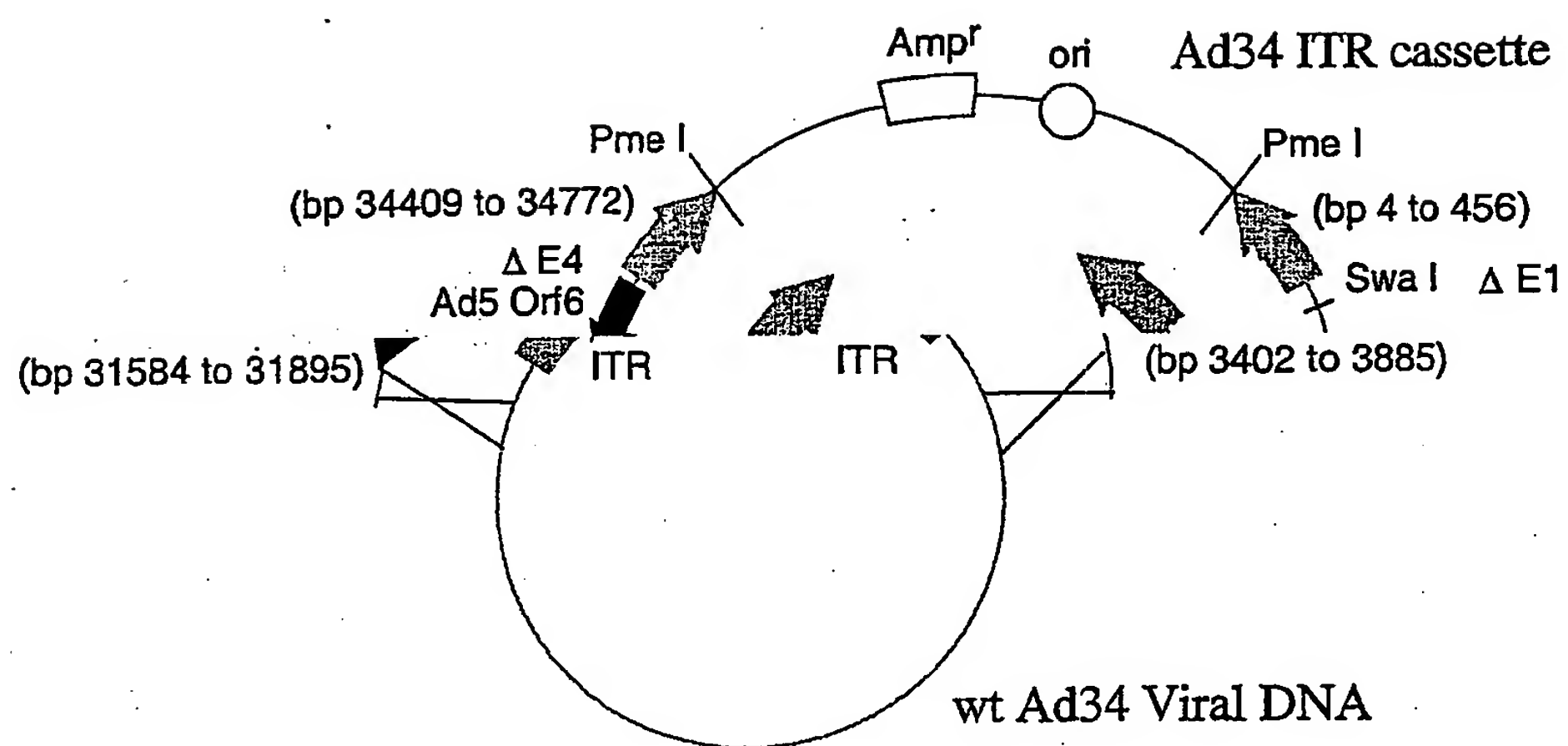


FIG. 27

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1 catcatcaat aatatacctt atagatggaa tgggtgccaat atgtaaatga ggtgatttta
61 aaaattgtgg ggtgtgtggt gattggctgt ggggttaacg gctaaacggg gcggcgcggc
121 cgtgggaaaa tgacgttttg tgggggtgga gtttttttgc aagttgtcgc gggaaatgtg
181 acgcataaaa aggccttttt tctcacggaa ctactgactt tccccacggt atttaacagg
241 aaatgaggta gttttgaccg gatgcaagtg aaaattgctg atttgcgcgc gaaaactgaa
301 tgaggaagtg tttttctgaa taatgtggta tttatggcag ggtggagtat ttgttcaggg
361 ccaggtagac tttgacccat tacgtggagg tttcgattac cgtgtttttt acctgaattt
421 ccgcgtaccg tgtcaaagtc ttctgttttt acgtagggtg cagctgatcg ctacggtatt
481 tatacctcag ggtttgtgtc aagaggccac tcttgagtgc cagcgagaag agttttctcc
541 tctgcgcggg cagtttaata ataaaaaaat gagagatttg cgatttctgc ctcaggaaat
601 aattttctgt gagactggaa atgaaatact ggagcttgtg gtgcacgccc tgatgggaga
661 cgatccggag ccacctgtgc agcttttttg gcctcctacg cttcaggaac tgtatgattt
721 agaggtagag ggatcggagg attctaataa ggaagctgtg aatggccttt ttaccgattc
781 tatgctttta gctgctaata aaggattaga attagatccg cctttggaca ctttcgatac
841 tccaggggtg attgtggaaa gcggtacagg tgtaagaaaa ttacctgatt tgggttccgt
901 ggactgtgat ttgcactgct atgaagacgg gtttcctccg agtgatgagg aggaccatga
961 aaaggagcag tctatgcaga ctgcagcggg tgagggagtg aaggctgcca gtgttggttt
1021 tcagttggat tgcccggagc ttccctggaca tggctgtatg tcttgtgaat ttcacaggaa
1081 aaatactgga gtaaaggaaac tgttatgttc gctttgttat atgagagcgc actgccactt
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1321 caagcccatt cctgtgaagc ttaagcctgg gaaacgtcca gcagtggaaa aacttgagga
1381 cttgtttacag ggtggggacg gacctttgga cttgagtaca cggaaacggc caagacaata
1441 agtgttccat atccgtgttt acttaagggtg acgtcaatat ttgtgtgaga gtgcaatgta
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2161 gtctcctgaa ctgcaacggg tgcttactgg atctacgtcc actggacggg ataggggcgt
2221 taagagggag agggcatcta gtggtactga tgctagatct gagttggctt taagttaaat
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3781 agttacttgt ccttttggcc cagctggagg ctttgacca acgtctgggt gaactttatc
3841 agcaggtggc cgagttgcga gtacaaactg agtctgctgt cggcacggca aagtctaaat

FIG. 28A-1

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3901 aaaaaaaaaat tccacaatca atgaataaat aaacgagctt gttgttgatt taaaatcaag
3961 tgttttttatt tcattttttcg cgcacggtat gccctagacc accgatctcg atcattgaga
4021 acacggtgga ttttttccag aatcctatag aggtgggatt gaatgttttag atacatgggc
4081 attaggccat ctttgggggtg gagatagctc cattgaagggt attcatgctc cggggtagtg
4141 ttgtaaatca cccagtcata acaaggctgc agtgcattggt gttgcacaat atcttttaga
4201 agtaggctga ttgccacaga taagcccttg gtgtagggtg ttacaaaccg gttgagctgg
4261 gaggggtgca ttccgggtga aattatgtgc attttggatt ggatttttaa gttggcaata
4321 ttgccgcaa gatctcgtct tgggttcatt ttatgaagga ccaccaagac ggtgtatccg
4381 gtacatttag gaaatttatc gtgtagcttg gatggaaaag cgtggaaaaa tttggagaca
4441 cccttgtgtc ctccgagatt ttccatgcac tcatccatga taatagcaat ggggcccgtg
4501 gcagcagcgc gggcaaacac gttccgtggg tctgacacat catagtattg ttccctgagt
4561 aaatcatcat aagccatttt aatgaatttg gggcggagag tacccgattg gggtagaat
4621 gttccttcgg gccccggagc atagtccccc tcacagattt gcatttccca agctttcagt
4681 tccgatggtg gaatcatgtc cacctggggg gctatgaaga acaccgtttc tggggcgggg
4741 gtgattagtt gggatgatag caagtctctg agcaattgag atttgccaca tccgggtggg
4801 ccataaatga ttccgattac aggttgccag tggtagttaa gggaaacggc actgccgtct
4861 tctcgaagca agggggccac ctccgttcac atttccctta catgcatatt ttcccgacc
4921 aaatccatta ggaggcgtc tccctcctagt gatagaagtt cttgtagtga ggaaaagttt
4981 ttcagcggtt ttagaccgtc agccatgggc attttgaga gagtttgctg caaaagttct
5041 agtctgttcc acagttcagt gatgtgttct atggcatctc gatccagcag acctcctcgt
5101 ttccgcggtt tggacggctc ctggagtagg gtatgagacg atgggcgtcc agcgctgcca
5161 ggggttcggc cttccagggg ctccagtgtt gcgcttcag actcattctg ctgggtggga
5221 ggtgtgcgcc tgcttgggcg cttgccaggg tgtatgtcgg ccaagtagca gtttaccatg agttcgtagt
5281 acttctgtcg cttggcgccc tgtatgtcgg cctttggcgc ggagcttacc tttggaagtt ttcttgcata
5341 tgagcgcctc ggctgcgtgg cctttggcgc gcttgggcgc aaggaaaatg gattctgggg
5401 ccgggcagta taggcatttc agcgcataca gcttgggcgc agttttcaca ttccaccagc caggttaaat
5461 agtatgcac tgcccgccag gaggcgcaaa cgccatattt tttgatgcgt ttcttacctt
5521 ccggttcatt ggggtcaaaa acaagttttc tgacaaacag gctgtccgta tcccctaga
5581 tgggtctccat gagttcgtgt cctcgttgag tgcctcggtc ttcttcgtac aggaactctg
5641 ctgattttac aggcctcttc tccagtggag ccagcacaaa ggaggctatg tgggaggggt
5701 accactctga tacaaaggcg cgcgtccagg tttccaaagt atgcaaacac atgtcacctt
5761 agcgatcggt gtcaaccagg ggggtccact aggtgtattt cacgtgacct ggggtccccg
5821 cttcaacatc caggaatgtg attggcttgt gctcttcctc actgtcttcc ggatcgctgt
5881 ctgggggggt ataaaagggg gcggttcttt cctctcga ggcgggcag acctctgcac
5941 ccaggaacgt cagctgttgg ggtaggtatt cctctcga ggcgggcag acctctgcac
6001 tcaggttgct agtttctaag aacgaggagg atttgatatt gacagtgccg gttgagatgc
6061 ctttcatgag gttttcgtcc atttggtcag aaaacacaaat ttttttattg tcaagtttgg
6121 tggcaaatga tccatacagg gcgttggata aaagtttggc aatggatcgc atgggttggg
6181 tcttttctct gtccgcgcgc tctttggcag cgatgttgag ttggacatac tcgcgtgcta
6241 ggcacttcca ttccgggaag atagtgttca attcatctgg cacgattctc acttgccacc
6301 ctcgattatg caaggtaatt aaatccacac tgggtggccac ctccgctcga aggggttcgt
6361 tgggtccaaca gagcctacct cctttcctag aacagaaagg gggaaagtggg tctagcataa
6421 gttcatcggg aggggtctga tccatggtaa agattcccgg aagtaaatcc ttatcaaaat
6481 agctgatggg agtggggtca tctaaggcca tttgccattc tcgagctgcc agtgcacgct
6541 catatgggtt aaggggactg ccccagggca tgggatgggt gagtgcagag gcatacatgc
6601 cacagatgtc atagacgtag atgggatcct caaagatgcc tatatagggt ggatagcatc
6661 gccccctct gatacttgct cgcacatagt catatagtct atgtgatggc gctagcaacc
6721 ccggacccaa gttggtgcga ttgggttttt ctgttctgta gacaatctgg cgaaagatgg
6781 cgtgagaatt ggaagagatg gtgggtcttt gaaaaatgtt gaaatgggca tgaggtagac
6841 ctacagagtc tctgacaaag tgggcataag attcttgaag cttggttacc agttcggcgg
6901 tgacaagtac gtctagggcg cagtagtcaa gtgtttcttg aatgatgtca taacctggtt
6961 ggtttttctt ttcccacagt tcgcggttga gaaggtattc ttccgactcc ttccagtact
7021 cttctagcgg aaaccctctt ttgtctgcac ggtaagatcc tagcatgtag aactgattaa
7081 ctgccttgta agggcagcag cccttctcta cgggtagaga gtatgcttga gcagcttttc
7141 gcagcgaagc gtgagtaagg gcgaagggtg ctctgaccat gactttgaga aattgggtatt
7201 tgaagtccat gtcgtcacag gctccctgtt cccagagttg gaagtctacc cgtttcttgt
7261 aggcgggggt gggcaaagcg aaagtaacat cgttgaagag aatcttaccg gctctgggca
7321 taaaattgcg agtgatgcgg aaaggctgtg gtacttccgc tcgattgttg atcacctggg
7381 cagctaggac gatctcgtcg aaaccgttga tgttgtgtcc tacgatgtat aattctatga
7441 aacgcggcgt gcctttgacg tgaggtagct tattgagctc atcaaaggtt aggtctgtag
7501 ggtcagataa ggcgtagtgt tcgagagccc attcgtgcag gtgaggattt gcatgtagga
7561 atgatgacca aagatccacc gccagtgtg tttgtaactg gtcccatac tgacgaaaat
7621 gctggccaat tgccattttt tctggagtga cacagtagaa ggttctgggg tcttgttgcc
7681 atcgatccca ctttagttta atggctagat cgtgggcat gttgacgaga cgctcttctc
7741 ctgagagttt catgaccagc atgaaaggaa ctagtgtgtt gccaaaggac cccatccagg

FIG. 28A-2

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7801 tgtaagtttc cacatcgtag gtcaggaaga gtctttctgt gcgaggatga gagccgatcg
7861 ggaagaactg gatttcctgc caccagttgg aggattggct gttgatgtga tggaagtaga
7921 agtttctgcg gcgcgcgcag cattcgtgtt tgtgcttgta cagacggccg cagtagtcgc
7981 agcgttgcaac gggttgtatc tcgtgaatga gctgtacctg gcttcccttg acgagaaatt
8041 tcagtgggaa gccgaggcct ggcgattgta tctcgtgctc ttctatatcc gctgtatcgg
8101 cctgttcatc ttctgtttcg gtggtggtca tgctgacgag ccccgcgggg aggcaagtcc
8161 agacctcggc gcgggagggg cggagctgaa ggaccagagc gcgcaggctg gagctgtcca
8221 gaggctcgag acgctgcgga ctcaggttag taggtaggga cagaagatta acttgcataa
8281 tcttttccag ggcgtgcggg aggttcagat ggtacttgat ttccacaggt tcgtttgtag
8341 agatgtcaat ggcttgccag gtccgtgtc ctttggggcg cactaccgta cctttgtttt
8401 ttcttttgat cgggtggtggc tctctgtgct cttgcatgct cagaagcgat gacggggacg
8461 cgcgcgcggc ggaagcgggt gttccggacc cggaggcatg gctggtagtg gcacgtcggc
8521 gccgcgcacg ggcaggttct ggtactgcgc tctgagaaga cttgctgctg ccaccacgcg
8581 tcgattgacg tcttgtatct gacgtctctg ggtgaaagct accggccccg tgagcttgaa
8641 cctgaaagag agttcaacag aatcaatttc ggtatcgta acggcagctt gtctcagtat
8701 ttcttgtacg tcaccagagt tgccttggtg ggcgatctcc gccatgaact gctcgatttc
8761 ttcttctcga agatctccgc gaccgcctct ctcgacggtg gccgcgaggt cattggagat
8821 accggcccatg agttgggaga atgcagtcac gccgcctcgc ttccagacgc ggctgtaaac
8881 cacggccccc tcggagtctc ttgcgcgcac caccacctga gcgaggttaa gctccacgtg
8941 tctggtgaag accgcatagt tgcataggcg ctgaaaaagg tagttgagtg tgggtggcaat
9001 gtgttcggcg acgaagaaat acatgatcca tcgtctcagc ggcatttcgc tgacatcgcc
9061 cagagcttcc aagcgtccca tggcctcgta gaagtccacg gcaaaattaa aaaactggga
9121 gtttcgcgcg gacacgggtca attcctcctc gagaagacgg atgagttcgg ctatggtggc
9181 ccgtacttcg cgttcgaagg ctcccgggat ctcttcttcc tcttctatct cttcttccac
9241 taacatctct tcttcgtctt caggcggggg cggagggggg acacggcgac gtcgacggcg
9301 cacggggcaa cggtcgatga atcgttcaat gacctctcgc cggcggcggc gcatggttcc
9361 agtgacggcg cggccgttct cgcgcggtcg cagagtaaaa acaccgcccgc gcatctcctt
9421 aaagtgggtg ctgggaggtt ctccgtttgg gagggagagg gcgctgatta tacattttat
9481 taattggccc gtagggactg cgcgcagaga tctgatcgtg tcaagatcca cgggatctga
9541 aaacctttcg acgaaagcgt ctaaccagtc acagtcacaa ggtaggctga gtacggcttc
9601 ttgtggggcg ggggtggttat gtgttcggtc tgggtcttct gtttcttctt catctcggga
9661 aggtgagacg atgctgctgg tgatgaaatt aaagtaggca gttctaagac ggcggatggt
9721 ggcgaggagc accaggtctt tgggtccggc ttgctggata cgcaggcgat tggccattcc
9781 ccaagcatta tcctgacatc tagcaagatc tttgtagtag tcttgcatga gccgttctac
9841 gggcacttct tcctcaccgc ttctgccatg catacgtgtg agtccaaacc cgcgcattgg
9901 ttgtaccagt gccaaagtcag ctacgactct ttcggcgagg atggcttgct gtacttgggt
9961 gaggggtggct tgaaagtcac caaaatccac aaagcgggtg taagccccgg tattaatggt
10021 gtaagcacag ttggccatga ctgaccagtt aactgtctgg tgaccagggc gcacgagctc
10081 ggtgtattta aggcgcgaat aggcgcgggt gtcaaagatg taatcgttgc aggtgcgcac
10141 cagatactgg taacctataa gaaaatgcgg cgggtggttg cggtagagag gccatcgttc
10201 tgtagctgga gcgcggggg cgaggtcttc caacataagg cggtagatag cgtagatgta
10261 cctggacatc caggtgatcc ctgcggcggt agtagaagcc ctaggaaact cgcgtacgcg
10321 gttccaaatg ttgcgtagcg gcatgaagta gttcattgta ggcacgggtt gaccagttag
10381 gcgcgcgcag tcattgatgc tctatagaca cggagaaaat gaaagcgttc agcgactcga
10441 ctccgtagcc tggagggaac tgaacgggtt gggtcgcggt gtaccccggt tcgagacttg
10501 tactcgagcc ggccggagcc gcggctaacc tgggtattggc actcccgtct cgaccagacc
10561 tacaaaaatc caggatacgg aatcgagtcg ttttgctggg tgccgaatgg caggggaagtg
10621 agtcctatct tttttttttg ccgctcagat gcatcccgtg ctgacgacaga tgcgtcccca
10681 acaacagccc ccctcgcagc agcagcaacc acaaaaggct gtccctgcaa ctactgcaac
10741 tgccgctgtg agcgggtgcg gacagcccg cctatgatct gacttggaag agggcggaagg
10801 actggcacgt ctaggtgcgc cttcgcgcga gcggcatccg cgagttcaac tgaaaaaaga
10861 ttctcgcgag gcgtatgtgc cccaacagaa cctatttaga gacagaagcg gcgaggagcc
10921 ggaggagatg cgagcttccc gctttaacgc gggtcgtgag ctgctgcacg gtttggacag
10981 aagacgagtg ttgcgggacg aggttttcca agttgatgaa gtgacaggga tcagtcctgc
11041 cagggcacac gtggctgcag ccaaccttgt atcggcttac gaacagacag taaaggaaga
11101 gcgtaatttc caaaagtctt ttaataatca tgtgcgaacc ctcattgccc gcgaagaagt
11161 cacccttggg ttgatgcatt tgtgggattt gatggaagct atcattcaga accctactag
11221 caaacctctg accgcacagc tgtttctggt ggtgcaacac agcagagaca atgaggcttt
11281 cagagaggcg ctgctcaaca tcaccgaacc cgaggggaga tgggtgtatg atcttatcaa
11341 cattctacag agtatcatag tgcaggagcg gagcctgggc ctggccgaga aggtggctgc
11401 catcaattac tcggttttga gcttgggaaa gtattacgct cgcaagatct acaagactcc
11461 atacgttccc atagacaagg aggtgaagat agatgggttc tacatgcgca tgacgtgaa
11521 ggtgttgacc ctgagcgatg atcttggggg gtaccgcaat gacagaatgc atcgcgcggt
11581 gagcgccagc agggagcgcg agttaagcga cagggaaact atgcacagtt tgcaagagc
11641 tctaactgga gctggaaccg aggtgagaa ttactttgat atgggagctg acttgcagt

FIG. 28A-3

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11701 gcagcctagt cgcagggctc tgaacgcccgc gacggcagga tgtgagcttc cttacataga
11761 agaggcggat gaaggcgagg aggaagaggg cgagtacttg gaagactgat ggcacaaccc
11821 gtgttttttg ctagatggaa cagcaagcac cggatcccgc aatgcgggcg gcgctgcaga
11881 gccagccgtc cggcatatac tcctcggacg attggacca ggccatgcaa cgtatcatgg
11941 cgttgacgac tcgcaacccc gaagccttta gacagcaacc ccaggccaac cgtctatcgg
12001 ccatcatgga agctgtagtg ccttcccgtc ctaatcccac tcatgagaag gtcctggcca
12061 tcgtgaacgc gttggtggag aacaaagcta ttcgtccaga tgaggccgga ctggtataca
12121 acgctctctt agaacgcgtg gctcgttaca acagtagcaa tgtgcaaacc aatttggacc
12181 gtatgataac agatgtacgc gaagccgtgt ctcagcgcga aaggttccag cgcgatgcca
12241 acctgggttc gctggtggcg ttaaattgctt tcttgagtac tcagcctgct aatgtgccgc
12301 gtggtcaaca ggattatact aacttttttaa gtgctttgag actgatggta tcagaagtac
12361 ctcagagcga agtatatcag tccgggtcctg attacttctt tcagactagc agacagggct
12421 tgcagacggg aaatctgagc caagcttttta aaaaccttaa aggtttgtgg ggagtgcag
12481 ccccggtagg agaaagagca accgtgtcta gcttggttaac tccgaactcc cgcctattat
12541 tactgttggg agctcctttc accgacagcg gtagcatcga ccgtaattcc tatttgggtt
12601 acctactaaa cctgtatcgc gaagccatag ggcaaagtca ggtggacgag cagacctatc
12661 aagaaattac ccaagtcagt cgcgctttgg gacaggaaga cactggcagt ttggaagcca
12721 ctctgaactt cttgcttacc aatcgggtctc aaaagatccc tectcaatat gctcttactg
12781 cggaggagga gaggatcctt agatatgtgc agcagagcgt gggattgttt ctgatgcaag
12841 agggggcaac tccgactgca gcactggaca tgacagcgcg aaatatggag cccagcatgt
12901 atgccagtaa ccgacctttc attaacaac tgctggacta cttgcacaga gctgccgcta
12961 tgaactctga ttatttcacc aatgccatct taaacccgca ctggctgccc ccacctggtt
13021 tctacacggg cgaatatgac atgcccagcc ctaatgacgg atttctgtgg gacgacgtgg
13081 acagcgatgt tttttcacct ctttctgac atcgcacgtg gaaaaaggaa ggcggcgata
13141 gaatgcattc ttctgcatcg ctgtccgggg tcattggtgc taccgcggt gagcccaggt
13201 ctgcaagtcc ttttcctagt ctaccctttt ctctacacag tgtacgtagc agcgaagtgg
13261 gtagaataag tcgcccaggt ttaatgggcg aagaggagta cctaaacgat tccttgctca
13321 gaccggcaag agaaaaaat ttcccaaaca atggaataga aagtttgggt gataaaatga
13381 gtagatggaa gacttatgct caggatcaca gagacgagcc tgggatcatg gggactacaa
13441 gtagagcgag ccgtagacgc cagcgccatg acagacagag ggtcttgtg tgggacgatg
13501 aggattcggc cgatgatagc agcgtattgg acttgggtgg gagaggaagg ggcaaccggt
13561 ttgctcatth gcgccctcgc ttgggtggta tgttgtaaaa aaaaataaaa aagaaaaaac
13621 tcaccaagge catggcgacg agcgtacgtt cgttcttctt tattatctgt gtctagtata
13681 atgaggcgag tcgtgctagg cggagcgggt gtgtatccgg agggctcctc tccttcgtac
13741 gagagcgtga tgcagcagca gcaggcgacg gcggtgatgc aatccccact ggaggctccc
13801 tttgtgcctc cgcgatacct ggcacctacg gagggcagaa acagcattcg ttactcgga
13861 ctggcacctc agtacgatac caccagggtg tatctggtgg acaacaagtc ggcggacatt
13921 gcttctctga actatcagaa tgaccacagc aacttcttga ccacggtggt gcaaaacaat
13981 gactttaccc ctacggaagc cagcacccag accattaact ttgatgaacg atcgcggtgg
14041 ggcggtcagc taaaaacccat catgcatact aacatgcccc acgtgaacga gtatatgttt
14101 agtaacaagt tcaaagcgcg tgtgatggtg tccagaaaac ctcttgaggg tgttagagta
14161 gacgataatt atgatcataa gcaagatatt ctaaaaatac agtggttcga gtttactttg
14221 ccagaaggca acttttcggt cactatgact atcgacttga tgaacaatgc catcatagac
14281 aattacttga aagtgggcag acagaatgga gtgttggaaa gtgacattgg tgttaagttc
14341 gacactagga acttcaagtt gggatgggat ccagaaacta agttgatcat gcctgggggt
14401 tacacctatg aggccttcca tcctgacatc gtattgctgc ctggctgcgg agtggaactt
14461 accgaaagcc gtctgagcaa ccttcttggc attagaaaga aacacccatt ccaagagggt
14521 tttatagatc tgtatgagga tttagaagga ggaatatatt cagccctttt ggatgtagat
14581 gcttatgaga acagcaagaa agatcaaaaa gccaaaatag aagctgctgc agaagctaaa
14641 gcaaacatag ttgccaacga tccggttaagg gtggctaacg ctagtgaat caggggagac
14701 agttttgccc caacatccgt tccgactaaa gaatcattat tggatgatgt gtctcaaac
14761 atagagttaa aactcactat taagcctgtg gaaaaagatg gcaaaaacag aagttacaat
14821 gtgttggaag ataaaaatcaa cacggcctat cgcagttggt acctttcgta caattatggc
14881 gaccccgaaa aaggagtgcg ttcttggaac ttgctacca cctcagatgt cacctgcgga
14941 gcggagcagg tctactggtc gcttccagac atgatgcagg atcctgtcac tttccgctcc
15001 actagacaag tcagtaacta ccctgtggtg ggtgcagagc ttatgcccgt cttttcaaag
15061 agcttctaca acgaacaagc tgtgtactcc cagcagctcc gccagtccac ctgccttacg
15121 caccgtctca accgctttcc tgagaaccag attttaatcc gtccgcccgc gccacaatt
15181 accaccgtca gtgaaaacgt tcctgctctc acagatcacg ggaccctgcc gttgcgcagc
15241 agtatccggg gagtccaacg tgtgaccgtt actgacgcca gacgccgcac ctgtccctac
15301 gtgtacaagg cactgggcat agtcgcaccg cgcgtccttt caagccgcac tttctaaaaa
15361 aaaaaaaaaa atgtccgttc ttatctcgcc cagtaataac accggttggg gtctgcgcgc
15421 tcccagcaag atgtacggag gcgcacgcaa acgttctacc caacatcccg tgcgtgttcg
15481 cgggcatttt cgcgtcccat ggggtgccct caagggccgc actcgcgttc gaaccacgt
15541 cgatgatgta atcgatcagg tggttgccga cgcgccgta tatactccta ctgcgcctac

FIG. 28A-4

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15601 atctactgtg gacgcagtta ttgacagtgt agtggctgac gctcgcaact atgctcgacg
15661 taagagccgg cgaaggcgca ttgccagacg tcaccgagct accactgcca tgcgagcagc
15721 aagagctctg ctacgaagag ctagacgcgt ggggcgaaga gccatgctta gggcgccag
15781 acgtgcagct tcgggcgcca gcgcggcgag gtcccgcagg caagcagccg ctgtcgcagc
15841 ggcgactatt gccgacatgg cccaatcgcg aagaggcaat gtatactggg tgcgtgacgc
15901 tgccaccggt caacgtgtac ccgtgcgcac ccgtccccct cgcacttaga agatactgag
15961 cagtctccga tgttgtgtcc cagcggcgag gatgtccaag cgcaaataca aggaagaaat
16021 gctgcagggt atcgcacctg aagtctacgg ccaaccgttg aaggatgaaa aaaaaccccg
16081 caaaatcaag cgggtaaaaa aggacaaaaa agaagaggaa gatggcgatg atgggctggc
16141 ggagtttgtg cgcgagtttg cccacggcg acgctgcaa tggcgtgggc gcaaagttcg
16201 acatgtgttg agacctggaa cttcgggtgt ctttacaccc ggcgagcgtt caagcgctac
16261 ttttaagcgt tcctatgatg aggtgtacgg ggatgatgat attcttgagc aggcagctga
16321 ccgattaggg gagtttgctt atggcaagcg tagtagaata aatcccaagg atgaaacagt
16381 gtccataccc ttggatcatg gaaatccac ccctagtctt aaaccggtca ctttgcagca
16441 agtgttaccg gtaactccgc gaacagggtg taaacgcgaa ggtgaagatt tgtatccac
16501 tatgcaactg atggtgcccc aacgccagaa gttggaggac gttttggaga aagtaaaagt
16561 ggatccagat attcaacctg aggttaaaagt gagaccatt aagcaggtag cgcctggtct
16621 gggagtacaa actgtagaca ttaaaattcc cactgaaagt gcaaaccggc ccatggatgc
16681 cgcaaagcct actgccacct ccactgaagt gcaaaccggc ccatggatgc ccatgcctat
16741 tacaactgac gccgtcggtc ccactogaag atcccgcagc aagtacggtc cagcaagtct
16801 gttgatgccc aactatgtcg tacacccatc tattattcct actcctgggt accgaggcac
16861 tcgctactat cgcagccgaa acagtacttc ccgcgtcgc cgcaagacac ctgcaaatcg
16921 cagtcgtcgc cgtagacgca caagcaaacc gattcccggc gccctgggtg ggcaagtgtg
16981 ccgcaatggg agtgcggaac ctttgacact gccgcgtgcg cgttaccatc ctagtatcat
17041 cacttaatac atgttgccgc tgcctccttg cagatatggc cctcacttgt cgccttcgcg
17101 ttcccatcac tggttaccga ggaagaaact cgcccgtag aagagggatg ttggggcgcg
17161 gaatgcgacg ctacaggcgca cggcgtgcta tccgcaagca attgcggggg ggttttttgc
17221 cagccttaat tccaattatc gctgctgcga ttggcgcaat accaggcata gcttccgtgg
17281 cggttcaggg ctcgcaacga cattgacatt ggaaaaaaa aaaacgtata aataaaaaat
17341 acaatggact ctgacactcc tggtagcttg actatgtttt cttagagatg gaagacatca
17401 atttttcatc cttggctccg cgacacggca cgaagccgta catgggcacc tggagcgaca
17461 tcggcacgag ccaactgaac gggggcgctc tcaattggag cagtatctgg agcgggctta
17521 aaaattttgg ctcaaccata aaaacatacg ggaacaaagc ttggaacagc agtacaggac
17581 aggcgcttag aaataaactt aaagaccaga acttccaaca aaaagtagtc gatgggatag
17641 cttccggtat caatggagtg gtagatttgg ctaaccaggc tgtgcagaaa aagataaaca
17701 gtcgtttgga cccgccgcca gcaaccccag gtgaaatgca agtggaggaa gaaattcctc
17761 cgccagaaaa acgaggcgac aagcgtccgc gtcccgatgt ggaagagacg ctggtgacgc
17821 gcgtagatga accgccttct tatgaggaag caacgaagct tggaaatgcc accactagac
17881 cgatagcccc tatggccacc ggggtgatga aaccttctca gttgcacga cccgtcacct
17941 tggatttgcc ccctcctcct gctgctactg ctgtaccgcg ttctaagcct gtcgctgccc
18001 cgaaaccagt cgccgtagcc aggtcacgtc ccggggggcg tcctcgtcca aatgcacact
18061 ggcaaaatac tctgaacagc atcgtgggtc taggcgtgca aagtgtaaaa cgccgtcgct
18121 gcttttaatt aaatatggag tagcgcttaa cttgcctatc tgtgtatatg tgtcattaca
18181 cgccgtcaca gcatcagagg aaaaaaggaa gaggtcgtgc gtcgacgctg agttacttct
18241 aagatggcca ccccatcgat gctgccccaa tgggcataca tgcacatcgc cggacaggat
18301 gcttcggagt acctgagtcg ggtctggtg cagttcgccc gcgccacaga cacctacttc
18361 aatctgggaa ataagtttag aaatcctacc gtagcgccga cccacgatgt gaccaccgat
18421 cgtagccagc ggctcatggt gcgcttcgtg cccgttgacc gggaggacaa tacatactct
18481 tacaagtgcc ggtacacctt ggccgtgggc gacaacagag tgctggatat ggccagcacg
18541 ttctttgaca ttaggggcgt gttggacaga ggtcccagtt ttaaacccta ttctggtacg
18601 gcttacaact ccctggctcc taaaggcgct ccaaatgcat ctcagtgggt ggataagggg
18661 gttacaagca ctggcctagt ggacgacggc aatactgatg atggggaaga agccaaaaaa
18721 gcaacataca ctttttggtg tgctccagta aaagccgagg ctgaaatcac aaaagacgga
18781 ttgccgggtg gcttggaagt ttcaactgaa ggtcctaaac caatctatgc tgataagctt
18841 tatcagccag aacctcaagt gggagacgaa acttggactg acctagacgg aaaaaccgaa
18901 gagtatggag ggagggttct taaacctgaa actaaaatga aaccctgcta cggatctttt
18961 gctaaacctc ctaatatata aggaggtcag gcaaaggtaa aaccaaaaga agacgatggc
19021 actaacaaca tcgagtatga cattgacatg aacttctttg acttaagatc acaaagatca
19081 gaactcaaac ctaaaattgt aatgtatgca gaaaatgtgg acctggaatg tccagatact
19141 catgttgtgt acaaacctgg agtttcagat gctagtcttg agaccaatct tggacaacag
19201 tctatgcccc acagacccaa ctacattggc ttacagagata acttcatcgg acttatgtac
19261 tataacagta ctggcaacat gggggtactg gctggccaag cgtctcagtt gaatgcagtg
19321 gttgacttgc aggacagaaa cacagaactg tcttaccacac tcttgcttga ctctctgggc
19381 gacagaacca gatactttag catgtggaat caggctgtgg acagtatatga tcctgatgta
19441 cgtgttattg aaaatcatgg tgtggaagat gaacttccca actattgttt tccgttggat

FIG. 28A-5

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19501 ggtgtcggtc cgcgaaacaga tagttacaag gagattaagc caaatggaga ccaatctact
19561 tggacaaatg tagacccaac tggcagcagt gaacttgcta agggaaatcc atttgccatg
19621 gaaattaacc ttcaagccaa tctatggcga agtttccctt attccaatgt ggctctatat
19681 ctcccagact cgtacaaata cccccgtcc aatgtcactc ttccagaaaa caaaaacacc
19741 tacgactaca tgaacgggcg ggtgggtgccc ccattctctag tagacaccta tgtgaacatt
19801 ggtgccagggt ggtctctgga tgccatggac aatgtcaacc cattcaacca ccaccgtaac
19861 gctggcttgc gttaccgatc catgcttctg ggtaacggac gttatgtgcc tttccacata
19921 caagtgcctc aaaaattctt cgctgttaaa aacctgctgc ttctcccagg ctctacact
19981 tatgagtga actttaggaa ggatgtaaac atgggtctac agagtccct cggtaacgac
20041 ctacgggtag atggcgccag catcagtttt acgagcatca acctctatgc tacttttttc
20101 cccatggctc acaacaccgc ttccaccctt gaagccatgc tgcggaatga caccaatgat
20161 cagtcattca acgactacct atctgcagct aacatgctct accccattcc tgccaatgca
20221 accaatattc ccatttccat tcttctcgc aactgggccc ctttcagagg ctggtcattt
20281 accagactga aaaccaaaga aactccctct ttggggctct gatttgacct ctacttcgtc
20341 tattctgggt ctattcccta cctggatggt acctctacc tgaaccacac ttttaagaag
20401 gtttccatca tgtttgactc ttcagtgage tggcctggaa atgacagggt actatctcct
20461 aacgaatttg aaataaagcg cactgtggat ggcgaaggct acaacgtagc ccaatgcaac
20521 atgaccaaag actggttctt ggtacagatg ctgcaccaact acaacatcgg ctatcagggc
20581 ttctacattc cagaaggata caaagatcgc atgtattcat ttttcagaaa cttccagccc
20641 atgagcaggc aggtggttga tgaggtcaat tacaaagact tcaaggccgt cgccataccc
20701 taccaacaca acaactctgg ctttgtgggt tacatggctc cgaccatgcg tcaaggtaaa
20761 ccctatccc ctaactatcc ctatccactc attggaacaa ctgccgtaaa tagtggtacg
20821 cagaaaaagt tcttgtgtga cagaaccatg tggcgcatat cgttctcaag caacttcatg
20881 tctatgggag cccttacaga cttgggacag aacatgctct atgccaactc agctcatgct
20941 ctggacatga cctttgaggt ggatcccatg gatgagccca ccctgcttta tcttctctc
21001 gaagttttcg acgtggtcag agtgcacatg ccacaccgcg gcatcatcga ggcagtctac
21061 ctgcgtagac cgttctcggc cggtaacgct accacgtaag aagcttcttg cttcttgcaa
21121 acagcagctg caaccatggc ctgcccgatcc caaaacggct ccagcgagca agagctcaga
21181 gccattgtcc aagacctggg ttgcccagca ttttttttgg gaacctttga taagcgcttc
21241 ccgggggttca tggcccccga taagctcgcc tgtgccattg taaatacggc cggacgtgag
21301 acgggggggag agcactgggt ggctttcggg tggaaacccac gttctaacac ctgctacctt
21361 tttgatecct ttggattctc ggatgatcgt ctcaaacaga tttaccagtt tgaatatgag
21421 ggtctcctgc gccgcagcgc tcttgctacc aaggaccggt gtattacgct ggaaaaatct
21481 acccagaccg tgcaggggcc ccgttctgoc gcctgcccag tttctgctg catgttctt
21541 catgcctttg tgcactggcc tgacogtccc atggacggaa accccaccat gaaattgcta
21601 actggagtgc caaacaacat gcttcattct cctaaagtcc agcccaccct gtgtgacaat
21661 caaaaagcac tctaccattt tctcaatacc cattcgctt attttcgctc tcatcgtaaa
21721 cacatcgaaa gggccactgc gttcgaccgt atggatgtgc aataatgatt catgtaaaca
21781 acgtgttcaa taaacagcac tttatttttt acatgtatcg aggtcttga ttacttattt
21841 atttacaagt cgaatgggtt ctgacgagaa tcagaatgac ccgacggcag tgatacgttg
21901 cggaactgat acttgggttg ccacttgaat tegggaaatca ccaacttggg aaccgggtata
21961 tcgggcagga tgtcactcca cagctttctg gtcagctgca aagctcccag caggtcagga
22021 gccgaaatct tgaaatcaca attaggacca gtgctctgag cgcgagagtt gcggtapacc
22081 ggattgcagc actgaaacac catcagcgac ggatgtctta cgcttgccag cacggtggga
22141 tctgcaatca tgcccacatc cagatcttca gcattggcaa tgctgaacgg ggtcatcttg
22201 caggtctgcc taccatggc gggcacccaa ttaggcttgt ggttacaatc gcagtgcagg
22261 gggatcagta tcatcttggc ctgatcctgt ctgattcctg gatacacggc tctcatgaaa
22321 gcatcatatt gcttgaaagc ctgctgggct ttactaccct cgggtataaaa catcccgcag
22381 gacctgctcg aaaactgggt agctgcgcag ccggcatcat tcacacagca gcgggcgtca
22441 ttgttggcta tttgcaccac acttctgccc cagcggtttt ggggtgattt ggttcgctcg
22501 ggattctcct tcaaggctcg ttgtccgttc tcgctggcca catccatctc gataatctgc
22561 tccttctgaa tcataatatt gccatgcaag cacttcagct tgccctcata atcattgcag
22621 ccatgaggcc acaacgcaca gcctgtacat tcccaattat ggtgggcat ctgagaaaaa
22681 gaatgtatca ttccctgcag aaatcttccc atcatcgtgc tcagtgtctt gtgactagtg
22741 aaagttaact ggatgcctcg gtgctcctcg ttcacgtact ggtgacagat gcgcttgat
22801 tgttcgtgct gctcaggcat tagtttaaaa gaggttctaa gttcgttate cagcctgtac
22861 ttctccatca gcagacacat cacttccatg cctttctccc aagcagacac caggggcaag
22921 ctaatcggat tcttaacagt gcaggcagca gctcctttag ccagagggtc atctttggcg
22981 atcttctcaa tgcttctttt gccatccttc tcaacgatgc gcacggcggt gtagctgaaa
23041 cccactgcta caagttgcgc ctcttctctt tcttctcgc tgtcttgact gatgtcttg
23101 atggggacat gtttgggtct ccttggcttc tttttcgggg gtatcggagg aggaggactg
23161 tcgctccggt ccggagacag ggaggattgt gacgtttcgc tcaccattac caactgactg
23221 tcggtagaag aacctgacc cacacggcga caggtgtttc tcttcggggg cagaggtgga
23281 ggcgattgag aagggtcggt gtccgacctg gaaggcggat gactggcaga accccttccg
23341 cgttcggggg tgtgctccct gtggcggtcg ctttaactgat ttccttcgcg gctggccatt

FIG. 28A-6

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23401 gtgtttctcct aggcagagaa acaacagaca tggaaactca gccattgctg tcaacatcgc
23461 cacgagtgcc atcacatctc gtcctcagcg acgaggaaaa ggagcagagc ttaagcattc
23521 caccgcccag tcctgccacc acctctaccc tagaagataa ggaggtcgac gcattctcatg
23581 acatgcagaa taaaaaagcg aaagagtctg agccagacat cgaacaagac ccgggctatg
23641 tgacaccggt ggaacacgag gaagagttga aacgctttct agagagagag gatgaaaact
23701 gcccaaaaca gcaagcggat aactatcacc aagatgctgg aaatagggat cagaacaccg
23761 actacctcat agggcttgac ggggaagacg cgctccttaa acatctagca agacagtcac
23821 tcatagtcaa ggatgcatta ttggacagaa ctgaagtgcc catcagtgtc gaagagctca
23881 gccgcgccta cgagcttaac ctatttttcac ctctactccc ccccaaactg cagccaaacg
23941 gcacctgcga gccaaatcct cgcttaaaact tttatccagc ttttgctgtg ccagaagtac
24001 tggctacctt tcacatcttt tttaaaaatc aaaaaattcc agtctcctgc cgcgctaate
24061 gcacccgcgc cgatgcccta ctcaatctgg gacctgggtc acgcttacct gatatagtt
24121 ccttggaaga ggttccaaag atcttcgagg gtctgggcaa taatgagact cgggcccga
24181 atgctctgca aaagggagaa aatggcatgg atgagcatca cagcgttctg gtggaattgg
24241 aagcgcgataa tgccagactc gcagctactc agcgaagcgt cgaggtcaca cactttgcat
24301 acccgcgtgt caacctgccc cctaaagtca tgacggccgt catggaccag ttactcatta
24361 agcgcgcaag tcccctttca gaagacatgc atgaccaga tgcctgtgat gagggtaa
24421 cagtggtcag tgatgagcag ctaacccgat ggctgggac cgactctccc cgggatttgg
24481 aagagcgtcg caagcttatg atggccgtgg tgctgggtac cgtagaacta gagtgtcttc
24541 ggcgtttctt taccgattca gaaaccttgc gcaaactcga agagaatctg cactacactt
24601 ttagacacgg ctttgtgcgg caggcatgca agatatctaa cgtggaactc accaacctgg
24661 tttcctacat gggatttctg catgagaatc gcctaggaca aagcgtgctg cacagcacc
24721 ttaaggggga agcccgccgt gattacatcc gcgattgtgt ttatctctac ctgtgccaca
24781 cgtggcaaac cggcatgggt gtatggcagc aatgtttaga agaacagAAC ctgaaagagc
24841 taaacaagct cttacagaaa tctcttaagg ttctgtggac agggttcgac gagcgcaccg
24901 tcgcttccga cctggcagac ctcatcttcc cagagcgtct cagggttact ttgcgaaacg
24961 gactgcctga ctttatgagc cagagcatgc ttaacaattt tctgtcttcc atcctggaac
25021 gctccggtat cctgcccgcg acctgctgcg cactgccctc cgactttgtg cctctcacct
25081 accgcgaatg cccccgcg ctatggagtc actgctacct gttccgtctg gccaactacc
25141 tctcctacca ctcggtatg atcgaggatg tgagcggaga cggcttgctg gagtgtcact
25201 gccgctgcaa tctgtgcag cccaccggt ccctagcttg caacccccag ttgatgagcg
25261 aaaccagat aataggcacc tttgaattgc aaggccccag cagccaaggc gatgggtctt
25321 ctccctggga aagtttaaaa ctgaccccg gactgtggac ctccgcctac ttgcgcaagt
25381 ttgccccgga agattaccac ccctatgaaa tcaagttcta tgaggacca tccagcctc
25441 cgaaagccga actttcggcc tgcgtcatca cccagggggc aattctggcc caattgcaag
25501 ccattccaaa atcccgcga gaatttctac tgaaaaagg taagggggtc taccttgacc
25561 cccagaccgg cgaggaactc aacacaagg tccctcagga tgtcccaacg acgagaaagc
25621 aagaagttga aggtgcagcc gccgccccca gaagatatg aggaagattg ggacagtcag
25681 gcagaggaag cggaggagga ggacagtctg gaggacagtc tggaggaaga cagtttggag
25741 gaggaaaacg aggaggcaga ggaggtggaa gaagtaaccg ccgacaaaca gttatcctcg
25801 gctgcggaga caagcaacag cgctaccatc tccgctccga gtcgaggaac ccggcggcgt
25861 cccagcagta gatgggacga gaccggacgc ttcccgaaac caaccagcgc ttccaagacc
25921 ggtaagaagg atcggcaggg atacaagtcc tggcgggggc ataagaatgc catcatctcc
25981 tgcttgcatg agtgcggggg caacatatcc ttacgcggc gctacttgct attccatcat
26041 ggggtgaact ttccgcgcaa tgttttgc atactaccgt acctccacag cccctactat
26101 agccagcaaa tcccggcagt ctgcacagat aaagacagcg gcggcgacct ccaacagaaa
26161 accagcagcg gcagttagaa aatacacaac aagtgcagca acaggaggat taaagattac
26221 agccaacgag ccagcgcaaa cccgagagtt aagaaatcgg atctttccaa ccctgtatgc
26281 catcttccag cagagtcggg gccaaagagc ggaactgaaa ataaaaaacc gatctctgcg
26341 ttcgctcacc agaagttggt tgtatcaca gagcgaagat caacttcagc gcactctcga
26401 ggacgcgag gctctcttca acaagtactg cgcgctgact cttaaagagt aggcagcgac
26461 cgcgcttatt caaaaaaggc gggaattaca tcatcctcga catgagtaaa gaaattccca
26521 cgccttacat gtggagttat cagcccaaaa tgggattggc ggcaggcgcc tcccaggact
26581 actccacccg catgaattgg ctacgcggc ggccttctat gatttctcga gttaatgata
26641 tacgcgccta ccgaaaccaa atacttttgg aacagtcagc tcttaccacc acgccccgc
26701 aacaccttaa tcccagaaat tggcccgccg ccctagtgt aacaggaaag cccgctccca
26761 ccactgtatt acttctcga gacgcccagg ccgaagtcca aatgactaat gcaggtgcgc
26821 agttagcgg cggctccacc ctatgtcgtc acaggcctcg gcataatata aaacgcctga
26881 tgatcagagg ccgaggtatc cagctcaacg acgagtcggt gagctctccg cttgggtctac
26941 gaccagacgg aatctttcag attgcgggct gcgggagatc ttccttcacc cctcgtcagg
27001 ctgttctgac tttggaaagt tctgtctcgc aaccccgctc gggcggaate gggaccgttc
27061 aatttgtgga ggagtttact ccctctgtct acttcaacc cttctccgga tctcctgggc
27121 actaccgga cgagttcata ccgaacttcg acgcgattag cgagtcagt gacggctacg
27181 attgatgtct ggtgacgcgg ctgagctatc tgggctgcga catctagacc actgccgcg
27241 ctttcgctgc tttgcccggg aactcattga gttcatctac ttcgaactcc ccaaggatca

FIG. 28A-7

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27301 ccctcaaggt ccggcccacg gagtgcggat tactatcgaa ggcaaaatac actctcgct
27361 gcaacgaatt ttctcccagc ggcccgtgct gatcgagcga gaccagggaa acaccacggt
27421 ttccatctac tgcatttgta atcaccccg attgcatgaa agcctttgct gtcttatgtg
27481 tactgagttt aataaaaact gaattaagac tctcctacgg actgccgctt cttcaaccg
27541 gatttttaca ccagaagaac gaaacttttc ctgtcgtcca ggactctgtt aacttcacct
27601 ttctactca caaactagaa gctcaacgac tacaccgctt ttccagaagc attttcccta
27661 ctaatactac ttcaaaaacc ggaggtgagc tccaaggtct tcctacagaa aacccttggg
27721 tggaagcggg ccttgtagtg ctaggaattc ttgcgggtgg gcttggtgatt attctttgct
27781 acctatacac accttgcttc actttcctag tgggtgtgtg gtattggttt aaaaaatggg
27841 gcccatacta gtcttgcttg ttttactttc gcttttgtaa ccgggttctg ccaattacga
27901 tccatgtcta gacttcgacc cagaaaactg cacacttact ttgcacccg acacaagccg
27961 catctgtgga gttcttatta agtgcggatg ggaatgcagg tccgttgaaa ttacacacaa
28021 taacaaaacc tggaacaata ccttatccac cacatgggag ccaggagttc ccgagtggta
28081 cactgtctct gtccgaggtc ctgacgggtc catccgcatt agtaacaaca ctttcatttt
28141 ttctgaaatg tgcgatctgg ccatgttcat gagcaaacag tattctctat ggctcctag
28201 caaggacaac atcgtaacgt tctccattgc ttattgcttg tgcgcttgcc ttcttactgc
28261 tttactgtgc gtatgcatac acctgcttgt aaccactcgc atcaaaaacg ccaataacaa
28321 agaaaaaatg ccttaacctc tttctgttta cagacatggc ttctcttaca tctctcatat
28381 ttgtcagcat tgtcactgcc gctcacggac aaacagtcgt ctctatccct ctaggacata
28441 attacactct cataggacc ccaatcactt cagaggtcat ctggaccaaa ctgggaagcg
28501 ttgattactt tgatataatc tgcaacaaaa caaaaccaat aatagtaact tgcaacatac
28561 aaaatcttac attgattaat gttagcaaa tttacagcgg ttactattat gggtatgaca
28621 gatacagtag tcaatataga aattacttgg ttcgtgttac ccagttaaaa accacgaaaa
28681 tgccaaatat ggcaaagatt cगतccgatg acaattctct aatgattgc aattgttgca gcggtggcag
28741 ccacaccgga cgaaaaaac atcccagatt ttttatatgc ttgtcgctac aaaaagtttc
28801 tggatgatgg actaataata atatgcattg ttaacattta atttcttttt atacagccat
28861 atcctaaaaa acaagatctc ctactaaggc ttatgcttac actctgactt ctgctcgtc
28921 gggttccact accacattcc gctcaaactg cacactaaaa ggacctcaag gtggtcatgt
28981 acacctcact gtaactatag acaatggatg gtttacaata ccatgtgacc aacctggtag
29041 cttttgggtg agaatatatg acctaaccat tatcaacgtg acagcaaatg acaaaggctt
29101 atttttctgc aacggcagag aaagtatgtt agattataac attattgtac tgccatctac
29161 ctattatgga accgactata ctactttctc tagcagcagt gtcgctaaca atacaatttc
29221 cactccagca ccccgacaaa tttttaaacy cactgtgaat aattctacaa cttcacatac
29281 caatecaacc tttgcccgcg tcagcattat cgctgcagtg acaattggaa tatctattct
29341 aacaatttcc acttcaacaa acgcctgctg ctatagaaaa gacaaacata aaggtgatcc
29401 tgtttttacc ataacctact aatttgttct tttttttttt atttacagta tggatgaacac
29461 attacttaga tttgatattt ttcttcttca ccatactcat ttgtgcattt aatgtttgcg
29521 caatcatggt acctagaaat acagcaaccc cagactgtat aggagcattt gcttcctatg
29581 ctactttcac agcagtagcc ttttggtact tgcactctgc tatgtagcat agtctgcctg gttattaatt
29641 cactttttgc tctagactgg atccttgtgc gaattgccta cctgcgccac catcccgat
29701 ttttccaact aaatatcgcg gcaacttcta gactcatcta aaaccatgca ggctatacta
29761 accgcaacca aatatctatt gcttccctac gctgtctcaa cccagctgc ctatagtact
29821 ccaatatttt tgcttctatt atgcaaattc caacaaccgt ggtcatttct tgcttgctat
29881 ccaccagaac accttagaaa cccaaattta ataattgatt ctggaataat taatataatc
29941 cgagaaaaat cagaaattcc tttgatatac cccctatttg attttggctg gaatgctccc
30001 tgttgacaca atcatccaca agaccagag gaacacattc ccctacaaaa catgcaacat
30061 aatgcacatg taatagatta cgaaagtga ccaacacccc cactactccc tgctattagt
30121 ccaatagcgc taaccggcgg agatgactga aacactcacc acctccaatt ccgcccagga
30181 tacttcaacc taaccggcgg gcgtctcaga acagcgactt gcccaactac gcatccgcca
30241 tctgctcgat atggacggcc aagagctcag agatgtcatc caaattcacc aatgcaaaaa
30301 gcagcaggaa cgcgcgcca aacaagccaa gatatactac gagatcaccg ctactgacca
30361 aggcataatc tggttggtaa gcccacaacg acaaaaattt acctgcatgg tgggaatcaa
30421 tgcctctctc tacgaacttg aaagtggaga tactaagggt tgcattcact gctcctgcga
30481 ccccatagtt atcacccagc ccctgctgaa gaccctatgc ggcctaagag acctgctacc
30541 ttccatcgag tgcacctaca aaaaatgatta acttacttga aatcagcaat aaggtctctg
30601 aatgaattaa aaatgatta ctcccagcag cactcactt cctcttccc aactctggta ttctaaacc
30661 ttgaaatttt catactttct ccatacttta aaggggatgt caaattttag ctctctcct
30721 cgttcagcgg tcttcatgtc tttcttccca gatgaccaag agagtccggc tcagtgactc
30781 gtaccacaaa gtctaccct atgaagatga aagcacctcc caacaccct ttataaacc
30841 cttcaaccct tccccaaatg gcttcacaca aagcccagac ggagttctta ctttaaaatg
30901 agggtttatt ttaaaccca ctaacaacca caggcggatc tctacagcta aaagtgggag ggggacttac
30961 ttaaaccca actgatggt ccttacaaga aaacatacgt gctacagcac ccattactaa
31021 agtgatgac tctgtagaac tatccattgg aatatgatta gaaactcaaa acaataaact
31081 aaataatcac tctgtagaac ggttaaaatt taacaacggt gacatttgta taaaggatag
31141 atgtgcaaaa ttgggaaatg

FIG. 28A-8

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31201 tattaacacc ttatggactg gaataaaccc tccacctaac tgtcaaattg tggaaaacac
31261 taatacaaat gatggcaaac ttactttagt attagtaaaa aacggagggc ttgttaatgg
31321 ctacgtgtct ctagtgtgtg tatcagacac tgtgaaccaa atgttcacac aaaagacagc
31381 aaacatccaa ttaagattat attttgactc ttctggaaat ctattaactg atgaatcaga
31441 cttaaaaatt ccacttaaaa ataaatcttc tacagcgacc agtgaaactg tagccagcag
31501 caaagccttt atgccaagta ctacagctta tcccttcaac accactacta gggatagtga
31561 aaactacatt catggaatat gttactacat gactagttaa gatagaagtc tatttccctt
31621 gaacatttct ataattgctaa acagccgtat gatttcttcc aatgttgcct atgccatata
31681 atttgaatgg aatctaaatg caagtgaatc tccagaaagc aacatagcta cgctgaccac
31741 atcccccttt ttcttttctt acattacaga agacgacaac taaaataaag ttttaagtgtt
31801 tttattttaaa atcacaaaat tctgagtgtt attttgcctc caccttccca tttgacagaa
31861 tacaccaatc tctccccacg cacagcttta aacatttggg taccattaga gatagacatt
31921 gtttttagatt ccacattcca aacagtttca gagcgagcca atctgggggc agtgatagat
31981 aaaaatccat cgcgatagtc ttttaaagcg ctttcacagt ccaactgctg cggatgcgaa
32041 tccggagtct ggatcacggg catctggaag aagaacgatg ggaatcataa tccgaaaacg
32101 gtatcggacg attgtgtctc atcaaaccga caagcagccg ctgtctgcgt cgctccgtgc
32161 aactgctgtt tatgggatca ggggtccacg tgtcctgaag catgatttta atagccctta
32221 acatcaactt tctggtgcga tgcgcgcagc aacgcattct gatttcactc aaatctttgc
32281 agtaggtaca acacattatt acaatattgt ttaataaacc ataattaaaa gcgctccagc
32341 caaaactcat atctgatata atcgccccctg catgaccatc ataccaaagt ttaatataaa
32401 ttaaatgacg ttccctcaaa aacacactac ccacatacat gatctctttt ggcattgtgca
32461 tattaacaat ctgtctgtac catggacaac gttgggttaat catgcaaccc aatataacct
32521 tccggaacca cactgccaac accgctcccc cagccatgca ttgaagtga cctgtctgat
32581 tacaatgaca atgaagaacc caattctctc gaccgtgaat cacttgagaa tgaaaaatat
32641 ctatagtggc acaacataga cataaatgca tgcattctct cataattttt aactcctcag
32701 gatttagaaa catatcccag ggaataggaa gctcttgtag aacagtaaag ctggcagaa
32761 aaggaagacc acgaacacaa cttacactat gcatagtcat agtatcacia tctggcaaca
32821 gcgggtgggc ttcagtcata gaagctcggg tttcattttc ctcacaacgt ggtaactggg
32881 ctctggtgta aggggtgatgt ctggcgcatg atgtcgagcg tgcgcgcaac cttgtcataa
32941 tggagttgct tcttgacatt ctcgtatttt gtatagcaaa acgcgccctt ggcagaacac
33001 actcttcttc gccttctatc ctgcccgtta gcgtgttccg tgtgatagtt caagtacagc
33061 cacactctta agttgggtcaa aagaatgctg gcttcagttg taatcaaaac tccatcgcat
33121 ctaattgttc tgaggaaatc atccacggta gcatatgcaa atcccaacca agcaatgcaa
33181 ctggattgag tttcaagcag gagaggagag ggaagagacg gaagaacat gttaattttt
33241 attccaaacg atctcgcagt acttcaaatt gtagatcgcg cagatggcat ctctcgcccc
33301 cactgtgttg gtgaaaaagc acagctaaat caaaagaaat gcgattttca aggtgctcaa
33361 cgggtggcttc caacaaagcc tccacgcgca catccaagaa caaaagaata ccaaagaag
33421 gagcattttc taactcctca atcatcatat tacattcctg caccattccc agataatttt
33481 cagctttcca gccttgaatt attcgtgtca gttcttgtgg taaatccaat ccacacatta
33541 caaacaggtc ccggagggcg ccctccacca ccattcttaa acacaccctc ataattgcaa
33601 aatatcttgc tctgtgtgca cctgtagcga attgagaatg gcaacatcaa ttgacatgcc
33661 cttggctcta agttcttctt taagttctag ttgtaaaaac tctctcatat tatcaccaaa
33721 ctgcttagcc agaagcccc ccgggaacaag agcaggggac gctacagtgc agtacaagcg
33781 cagacctccc caattggctc cagcaaaaac aagattggaa taagcatatt gggaaccgcc
33841 agtaatatca tccaagttgc tggaaatata atcaggcaga gtttcttcta aaaattgaat
33901 aaaagaaaaa tttgccaaaa aaacattcaa aacctctggg atgcaaatgc aatagggttac
33961 cgcgctgcgc tccaacattg ttagttttga attagtctgc aaaaataaaa aaaaaaaca
34021 gcgtcatatc atagtagcct gaggaacagg tggataaatc agtctttcca tcacaagaca
34081 agccacaggg tctccagctc gaccctcgta aaacctgtca tgggtattaa acaacagcac
34141 cgaaagttcc tgcggtgac cagcatgaat aattcttgat gaagcatata atccagacat
34201 gttagcatca gttaacgaga aaaaacagcc aacatagcct ttgggtataa ttatgcttaa
34261 tctgaagtat agcaaagcca cccctcgcgg atacaaagta aaaggcacag gagaataaaa
34321 aatataatta tttctctgct gctgttcagg caacgtcgcc cccggctccct ctaaatacac
34381 atacaaagcc tcatcagcca tggcttacca gacaaagtac agcggggcacg cacaagctct
34441 aaagtcactc tccaacctct ccacaatata tatacacaag ccctaaactg acgtaatggg
34501 agtaaagtgt aaaaaatccc gccaaaccca acacacaccc cgaaactgag tcaccagggg
34561 aaagtacagt ttcacttccg caatcccaac aagcgtcact tctcttttct cagggtagct
34621 cacatcccat taacttgcaa cgtcattttc ccacggccgc gccgccccgt ttagccgtta
34681 accccacagc caatcaccac acaccccaca atttttaaaa tcacctcatt tacatattgg
34741 caccattcca tctataaggt atattattga tgatg

SEQ ID NO: 12

FIG. 28A-9

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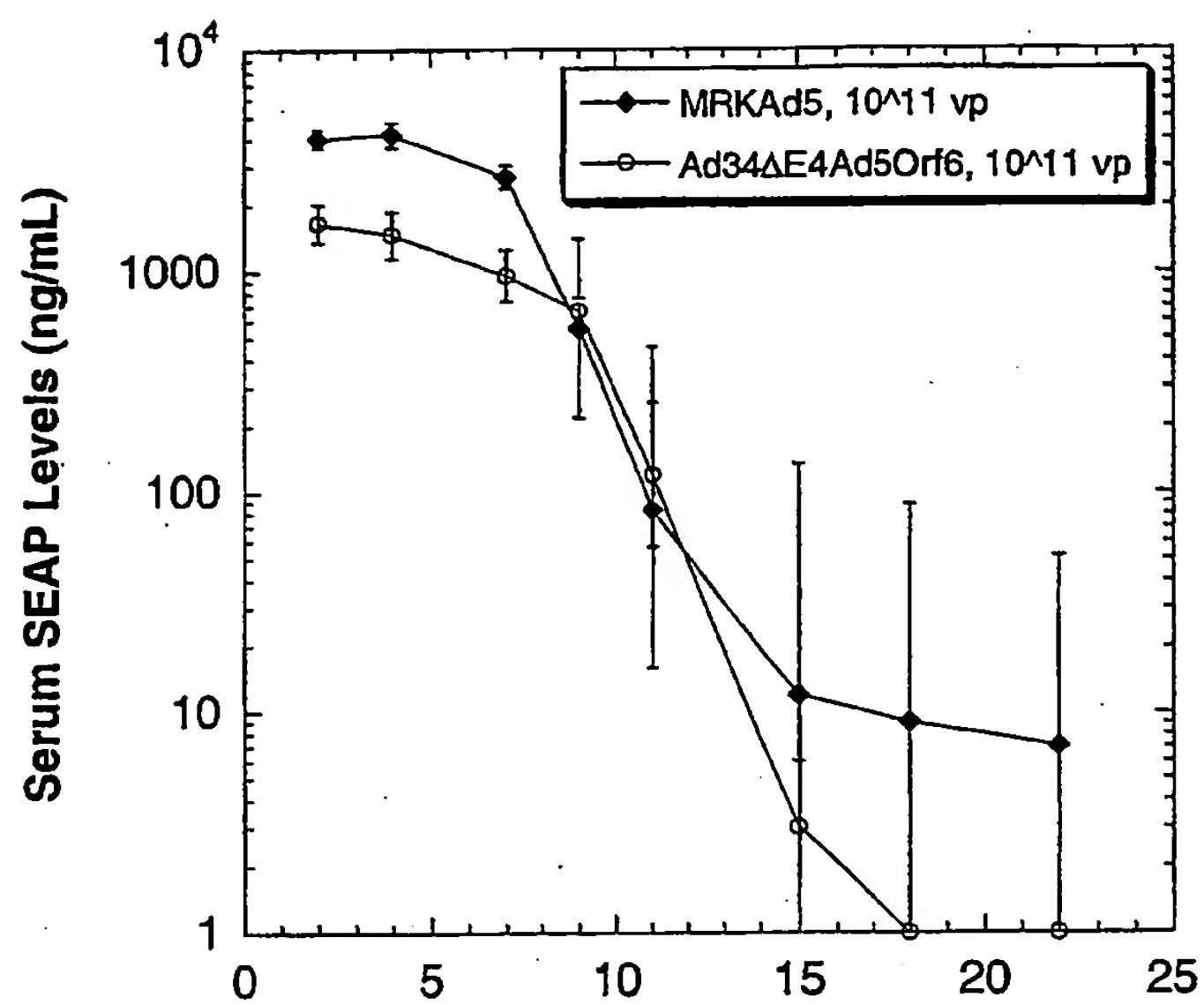


FIG. 29

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Vaccine Wk 0, 4, 24	Monkey ID	Pre		Wk 4		Wk 8		Wk 24		Wk 28		Wk 36	
		Mock	Gag ^a	Mock	Gag	Mock	Gag	Mock	Gag	Mock	Gag	Mock	Gag
MRKAd5gag, 10 ⁶ 11 vp	00C018	1	5	13	1025	0	824	8	756	0	474	0	383
MRKAd5gag, 10 ⁶ 11 vp	00C034	0	4	5	219	5	404	3	445	3	339	0	216
MRKAd5gag, 10 ⁶ 11 vp	00C058	4	4	3	1086	0	440	4	1439	0	2338	0	840
Ad34ΔE1gagΔE4Ad5Orf6, 10 ⁶ 11 vp	00D038	6	8	5	111	1	301	0	224	1	535	0	233
Ad34ΔE1gagΔE4Ad5Orf6, 10 ⁶ 11 vp	00D042	6	30	4	89	4	264	1	73	0	181	0	69
Ad34ΔE1gagΔE4Ad5Orf6, 10 ⁶ 11 vp	00D055	3	18	1	118	1	816	0	429	0	439	0	273

FIG. 30

Vaccine	Monk ID	IFN- γ ⁺ CD4 ⁺ CD3 ⁺ per 10 ⁶ Lymphocytes		IFN- γ ⁺ CD8 ⁺ CD3 ⁺ per 10 ⁶ Lymphocytes	
		Mock	Gag ^a	Mock	Gag ^a
Ad34 Δ E1gag Δ E4Ad5Orf6	00D038	22	154	130	450
	00D042	32	118	96	171
	00D066	12	238	150	442

FIG. 31

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Vaccine T=0, 4 wks	Vaccine T=24 wks	Monkey ID	Pre		T=4 wks		T=8 wks		T=24 wks		T=28 wks		T=32 wks	
			Mock	Gag ^a	Mock	Gag	Mock	Gag	Mock	Gag	Mock	Gag	Mock	Gag
Ad34ΔE1gagΔE4Ad5Orf6, 10 ⁶ 11 vp	Ad35ΔE1gagΔE4Ad5Orf6, 10 ⁶ 10 vp	00D018	4	8	1	84	5	334	5	89	0	308	3	244
Ad34ΔE1gagΔE4Ad5Orf6, 10 ⁶ 11 vp	Ad35ΔE1gagΔE4Ad5Orf6, 10 ⁶ 10 vp	00D044	1	1	8	79	0	374	8	138	0	453	1	253
Ad34ΔE1gagΔE4Ad5Orf6, 10 ⁶ 11 vp	Ad35ΔE1gagΔE4Ad5Orf6, 10 ⁶ 10 vp	00D084	4	8	1	125	8	655	8	145	0	351	1	236
NaNa		00D087	1	1	3	3	8	54	8	8	5	5	3	0

FIG. 32

Vaccine (T=0, 4 Wks)	Vaccine (T=24 Wk)	Monkey ID	IFN- γ ⁺ CD4 ⁺ CD3 ⁺ per 10 ⁶ Lymphocytes		IFN- γ ⁺ CD8 ⁺ CD3 ⁺ per 10 ⁶ Lymphocytes	
			Mock	Gag	Mock	Gag
Ad34 Δ E1gag Δ E4Ad5Orf6, 10 ¹¹ vp	Ad35 Δ E1gag Δ E4Ad5Orf6, 10 ¹⁰ vp	00D016	62	433	176	1288
Ad34 Δ E1gag Δ E4Ad5Orf6, 10 ¹¹ vp	Ad35 Δ E1gag Δ E4Ad5Orf6, 10 ¹⁰ vp	00D044	136	593	323	1871
Ad34 Δ E1gag Δ E4Ad5Orf6, 10 ¹¹ vp	Ad35 Δ E1gag Δ E4Ad5Orf6, 10 ¹⁰ vp	00D064	188	785	292	892

FIG. 33

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